



DISSERTATION

Titel der Dissertation

“Total Syntheses of (-)-Kendomycin and
(+)-Echinopine A and B”

Verfasser

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*Für meine Eltern Karl und Irmgard
und für Nadine*

*DIE GEFÄHRLICHSTE WELTANSCHAUUNG IST DIE WELTANSCHAUUNG
DERJENIGEN, DIE DIE WELT NICHT ANGESCHAUT HABEN*

(Alexander von Humboldt)

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Publications, Conference Contributions and Awards Resulting from this Thesis

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Peer-Reviewed

“Total Synthesis of the Antibiotic Kendomycin by Macrocyclization via Photo-Fries Rearrangement and Ring Closing Metathesis (RCM)”, T. Magauer, H. J. Martin, J. Mulzer, *Angew. Chem., Int. Ed.* **2009**, *48*, 6032–6036.

“Ring Closing Metathesis and Photo-Fries Reaction for the Construction of the Ansamycin Antibiotic Kendomycin. Development of a Protecting Group Free Oxidative Endgame”, T. Magauer, H. J. Martin, J. Mulzer, *Chem. Eur. J.* **2009**, *Accepted*.

“Total Synthesis of (+)-Echinopine A and B: Determination of Absolute Stereochemistry”, T. Magauer, J. Mulzer, K. Tiefenbacher, *Org. Lett.* **2009**, *ASAP*.

Press-Releases

Radio

„Neuartiges Antibiotikum gegen resistente Keime“, E. Schütz, *Dimensionen-Magazin*, Österreich 1 (Ö1), 11.09.2009.

Journals

“Neue Leitstrukturen für Antibiotikum entwickelt“, A. Geipel-Kern, *PROCESS PharmaTEC*, Vogel Business Media, Würzburg **2009**, *5*, 14.

Online

„Neuartiges Antibiotikum gegen resistente Keime“, W. Weitlaner, Presstext Austria, <http://presstext.at>, www.derstandard.at, www.wien-heute.at (08.09.2009); V. Schallhart, Universität Wien, <http://public.univie.ac.at> (08.09.2009); R. Müller, Portal für Organische Chemie, www.organische-chemie.ch (09.09.2009), M. Mössmer, Österreich Journal, www.oesterreichjournal.at Ausgabe 76, 60 (07.10.2009).

„Neue Hoffnung gegen resistente Bakterien: Wiener Chemiker synthetisieren Wirkstoff“, Austria Presse Agentur, www.news.at, www.oe24.at, www.springermedizin.at (08.09.2009).

Conference Contributions

Posters

Poster presented at the “**Gordon Research Conference – Natural Products**”. Title of presented poster: “Total Synthesis of the *ansa*-Polyketide Kendomycin via Photo-Fries Rearrangement & Ring Closing Metathesis”, Tilton, NH, USA (26–31 July 2009).

Poster presented at the “**10th Tetrahedron Symposium**”. Title of presented poster: “Total Synthesis of the *ansa*-Polyketide Kendomycin via Photo-Fries Rearrangement & Ring Closing Metathesis”, Paris, France (13–26 June 2009).

Poster presented at the “**Synthesefest of the Ludwig-Maximilian University**”. Title of presented poster: “Total Synthesis of Kendomycin”, Munich, Germany (17–18 March 2009).

Poster presented at the “**21. Irseer Naturstofftage der DECHEMA e.V.**”. Title of presented poster: “Total Synthesis of Kendomycin”, Irsee, Germany (25–27 February 2009).

Poster presented at the “**IASOC 2008 – Ischia Advanced School of Organic Chemistry**”. Title of presented poster: “Towards the Total Synthesis of Kendomycin”, Ischia, Italy (27 September–02 October 2008).

Poster presented at the “**BOSS XI – 11th Belgium Organic Synthesis Symposium**”. Title of presented poster: “Towards the Total Synthesis of the *ansa*-Macrocyclic Kendomycin”, Ghent, Belgium (13–18 July 2008).

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Invited oral presentation at the “**38. Doktorandenworkshop Naturstoffe: Chemie, Biologie und Ökologie Leibniz-Institut für Pflanzenbiochemie**”. Title of presentation: “Neue Projekte aus der Wiener Naturstoffsynthese”, Halle (Saale), Germany (16 October 2009).

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Invited oral presentation at the “**Symposium of Organic Chemistry at ESPCI**”. Title of presentation: “Total Synthesis of Kendomycin”, Paris, France (6 February 2009).

Awards

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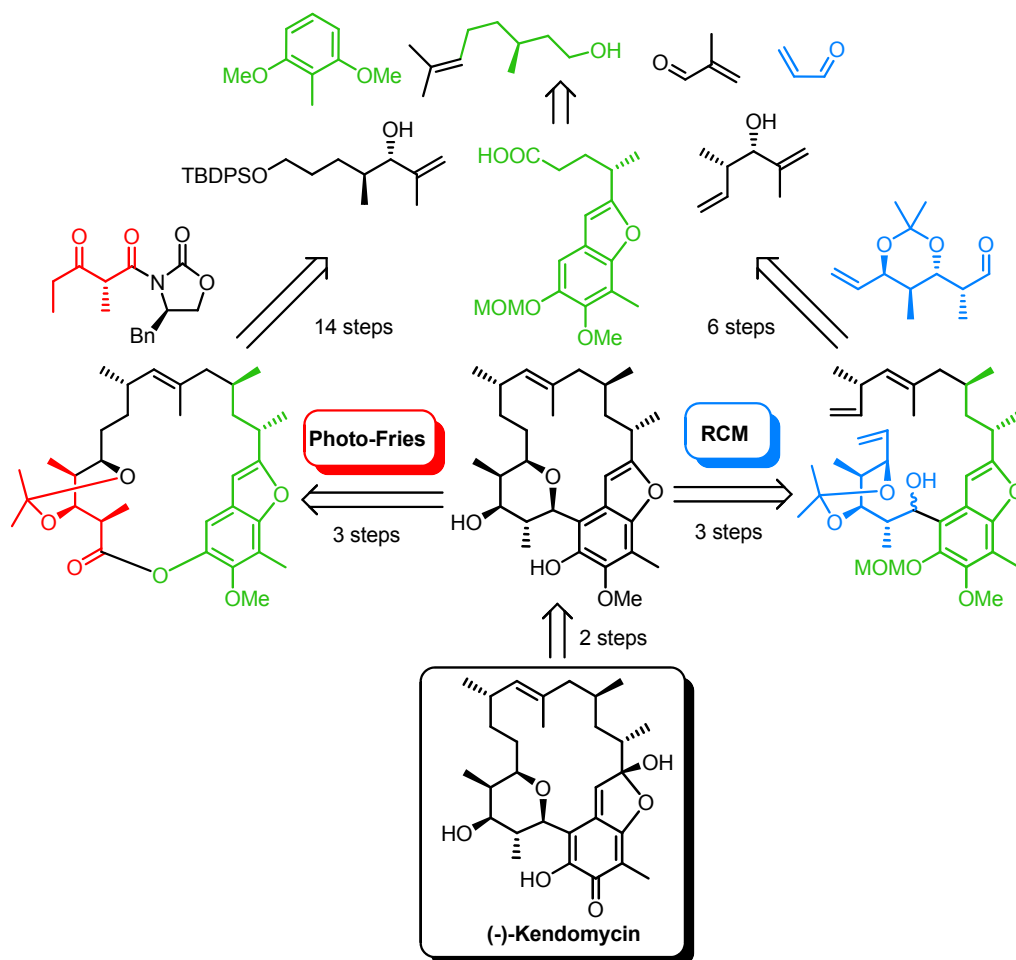
Title of presented poster: “Total Synthesis of Kendomycin”, Munich, Germany (17–18 March 2009).



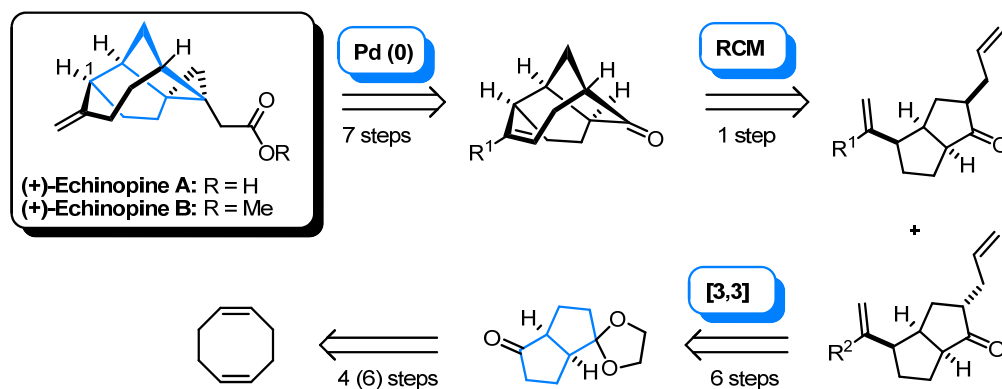
The “kendomycin–family”: T. Magauer, A. B. Smith III and C. Lee at the Gordon Natural Products Research Conference 2009 (Tilton, NH, USA).

A. Graphical Abstract

Total Synthesis of (-)-Kendomycin



Total Syntheses of (+)-Echinopine A and B: Determination of Absolute Stereochemistry



B. Abstract

This Ph.D. thesis describes the first total syntheses of (+)-echinopine A and B and two total syntheses of the antibiotic (-)-kendomycin.

Kendomycin, an unprecedented *ansa*-polyketide was first isolated in 1996 (in 1999 re-isolation from *Streptomyces violaceoruber*) and has been fascinating (bio)chemists due to its promising biological activity (endothelin receptor antagonist, antiosteoporotic and antibacterial activity, cytotoxic activity comparable to cisplatin) and remarkable complex molecular framework (fully carbogenic *ansa*-chain, nine stereocenters, pentasubstituted tetrahydropyran ring, unique *p*-quinone methide chromophore).

The total syntheses focused on two convergent strategies, whereas both took advantage of diastereoselective Claisen-Ireland rearrangements of unusual complexity to construct the north-east domain. Formation of the fully substituted carbon skeleton was either accomplished via a highly efficient photo-Fries rearrangement or ring closing metathesis (RCM) reaction. Apart from the application of the powerful RCM reaction, the so far underestimated photo-Fries rearrangement was extended to the construction of macrocycles. The final steps to install the quinone-methide-lactol unit could be performed by a chemoselective oxidation-hydrolysis sequence, thus avoiding additional protective groups manipulations.

Echinopine A and **B**, novel tetracyclic sesquiterpenoids from *Echinops spinosus* were isolated in 2007 and have attracted great interest because of their unique 3,5,5,7-membered ring skeleton. Limitation of material and scarcity of sample source prevented proper biological testing and determination of the absolute stereochemistry. These limitations, the complex and novel molecular architecture render the title compounds ideal targets for total synthesis.

The concise total syntheses are characterized by (1) 1,5-cyclooctadiene as an inexpensive, easily available starting material, (2) the extension of Myers' [3,3]-sigmatropic protocol to the stereoselective installation of vinyl and isopropenyl units, (3) RCM as an efficient tool to close the strained seven-membered ring and (4) the extension of an unusual Pd(0)-C₂-homologation of phenyltriflates to vinyltriflates. In addition, the proposed structures were validated and the absolute stereochemistry could be determined.

C. Zusammenfassung

Die vorliegende Dissertation beschreibt die Totalsynthesen von (+)-Echinopine A und B, sowie zwei Totalsynthesen des Antibiotikums (-)-Kendomycin.

Kendomycin, ein neuratiges *ansa*-Polyketid, wurde 1996 erstmals isoliert (Reisolierung 1999 aus *Streptomyces violaceoruber*) und hat seitdem (Bio)Chemiker aufgrund der vielversprechenden biologischen Aktivität (Endothelin-Rezeptor-Antagonist, antiosteoporotische/antibakterielle Aktivität, zytotoxische Aktivität vergleichbar mit Cisplatin) und der komplexen molekularen Struktur (vollständig kohlenstoffhaltige *ansa*-Kette, neun Stereozentren, fünffach substituierter Tetrahydropyran Ring, einzigartiger *p*-chinoider Chromophor) fasziniert.

Die Totalsynthesen stützten sich auf zwei konvergente Strategien, wobei beide auf einer ungewöhnlich komplexen, diastereoselektiven Claisen-Ireland Umlagerung für den Aufbau der nordöstlichen Domäne basierten. Die Bildung des vollständig substituierten Kohlenstoffskelettes wurde einerseits durch eine äußerst effiziente Photo-Fries Reaktion beziehungsweise durch ringschließende Metathese (RCM) erreicht. Abgesehen von der effizienten RCM Reaktion, wurde die bis jetzt unterschätzte Photo-Fries Reaktion für den Aufbau von Makrozyklen erweitert. Der Aufbau der Chinon-Laktol Einheit konnte mit Hilfe einer chemoselektiven Oxidations-Hydrolyse Sequenz, ohne Hinzunahme von Schutzgruppen, bewältigt werden.

Echinopine A und B, neuartige tetrazyklische Sesquiterpene, wurden 2007 aus *Echinops spinosus* isoliert und lenkten aufgrund ihres einzigartigen 3,5,5,7- Ringgerüsts die Aufmerksamkeit auf sich. Aufgrund der geringen Verfügbarkeit der Naturstoffe und der Probenquelle konnten weder angemessene biologische Tests, noch eine Bestimmung der Absolutkonfiguration durchgeführt werden. Diese Einschränkungen und die komplexe molekulare Architektur machen die Titelverbindungen zu idealen Zielstrukturen für die Totalsynthese.

Die Totalsynthesen bestechen durch (1) die Verwendung von 1,5-Cyclooctadien als billiges, leicht zugängliches Startmaterial, (2) die Erweiterung von Myers' [3,3]-sigmatropen Umlagerungsprotokolls für die stereoselektive Anlegung von Vinyl- und Isopropenyleinheiten, (3) eine RCM um den gespannten sieben-gliedrigen Ring aufzubauen und (4) die Erweiterung einer ungewöhnlichen Pd(0)-C₂-Verlängerung von Phenyltriflaten auf Vinyltriflate. Zusätzlich konnten die ursprünglich vorgeschlagenen Strukturen validiert und die absolute Stereochemie bestimmt werden.

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PART A: Total Synthesis of (-)-Kendomycin

1. Introduction and Background

1.1. Isolation and Biological Activity

The novel *ansa*-macrolide kendomycin (**1**) was originally isolated by Funahashi¹ from *Streptomyces* sp. AL-71389 as (-)-TAN 2162 and in 2000, Zeeck's group from Göttingen disclosed the re-isolation of **1** from *Streptomyces violaceoruber*.²

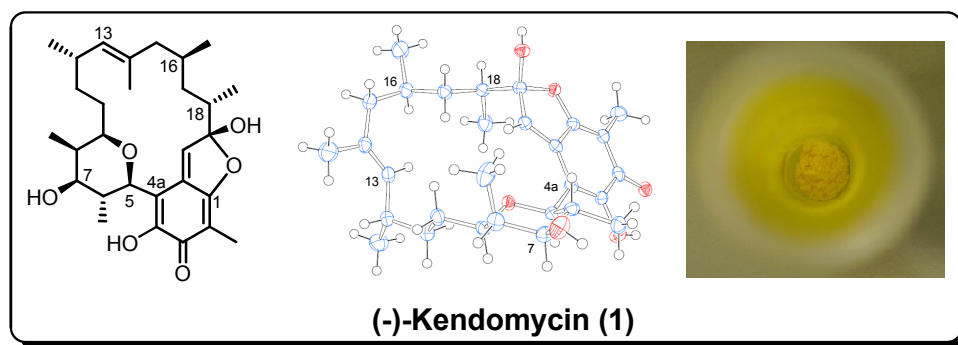


Figure 1. 2D-view and X-ray structure of kendomycin (**1**). Re-crystallization from dichloromethane provided yellow crystals suitable for X-ray analysis.

The patent literature describes compound **1** as a highly active endothelin receptor antagonist with promising antiosteoporotic activity.^{1,3} *In vitro* assays by Zeeck and co-workers revealed that **1** and **2** (see Scheme 1) also efficiently act against different human tumor cell lines (HMO2, HEP-G2 and MCF7). The cytotoxic activity is comparable to the established drugs cisplatin and doxorubicin, whereas the mode of action is mainly attributed to proteasome inhibition.⁴ In addition, kendomycin exhibits a remarkable antibiotic activity against Gram-positive and Gram-negative bacteria, including multidrug-resistant *Staphylococcus aureus* (MRSA) strains. In 2008, Boyce reported that kendomycin and several

¹ (a) Y. Funahashi, N. Kawamura, T. Ishimaru, *Jap. Pat.* 08 231 551 [A2 960 910], **1996**; *Chem. Abstr.* **1997**, 126, 6553; (b) Y. Funahashi, T. Ishimaru, N. Kawamura, *Jap. Pat.* 08 231 552 [A2 960910], **1996**; *Chem. Abstr.* **1996**, 125, 326518.

² (a) H. B. Bode, A. Zeeck, *J. Chem. Soc., Perkin Trans. I* **1999**, 323–328; (b) H. B. Bode, A. Zeeck, *J. Chem. Soc., Perkin Trans. I* **2000**, 2665–2670.

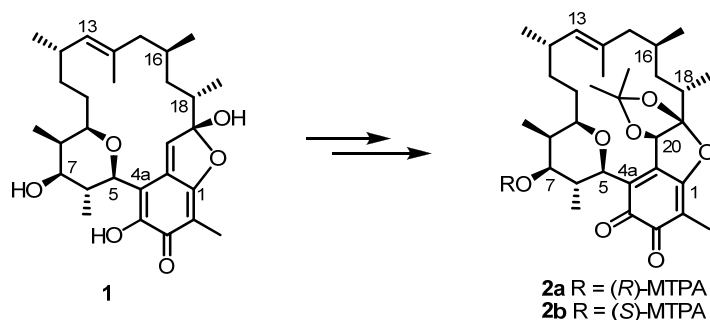
³ M. H. Su, M. I. Hosken, B. J. Hotovec, T. L. Johnston, *US Pat.* 5728727 [A980317], **1998**; *Chem. Abstr.* **1998**, 128, 239489.

⁴ Y. A. Elnakady, M. Rohde, F. Sasse, C. Backes, A. Keller, H.-P. Lenhof, K. J. Weissman, R. Mueller, *ChemBioChem* **2007**, 8, 1261–1272.

analogues inhibit the Bcl-xl-Bak interaction, which plays a significant role in apoptosis.⁵ These salient qualities render compound **1** an interesting and promising agent for further drug development.

1.2. Structure Elucidation and Molecular Framework

Based on careful 2D-NMR experiments (HMBC, COSY), HRMS and X-ray analysis, Zeeck's group was able to unambiguously assign the relative stereochemistry of **1**.² Determination of the absolute stereochemistry was achieved by means of advanced Mosher's ester analysis of stable ketal **2**, which was obtained from **1** by oxidation with FeCl₃ in acetone and subsequent derivatization (Scheme 1).



Scheme 1. Determination of the absolute stereochemistry of (-)-**1** via Mosher's advanced ester methodology. The C7-stereocenter was designated to be (S)-configured.

The outstanding and unprecedented molecular structure of **1** is manifested in nine stereocenters, a fully carbogenic *ansa*-chain, a highly substituted tetrahydropyran ring and a unique *p*-quinone methide chromophor. The latter, which is susceptible to nucleophilic 1,6-addition at the C20 position, seems to be responsible for the instability of compound **1** in aqueous DMSO and methanol.⁵ Figure 2 emphasizes the synthetically most challenging domains of kendomycin.

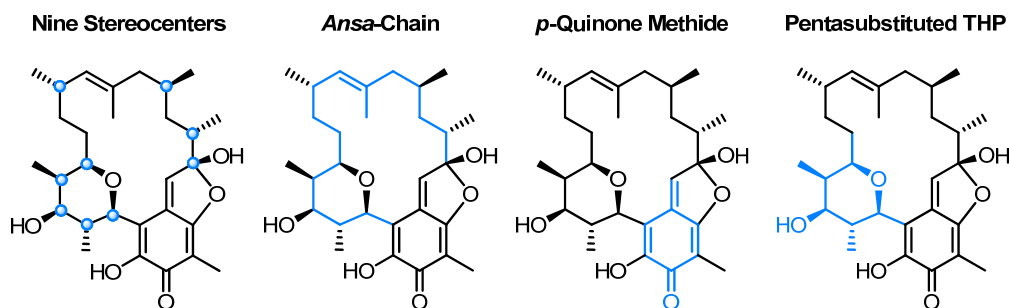


Figure 2. Structural features of **1**.

⁵ C. O. Janssen, S. Lim, E. P. Lo, K. F. Wan, V. C. Yu, M. A. Lee, S. B. Ng, M. J. Everett, A. D. Buss, D. P. Lane, R. S. Boyce, *Bioorg. Med. Chem. Lett.* **2008**, *18*, 5771-5773.

1.3. Biosynthesis

The general biosynthesis of polyketides starts with the polyketide synthase (PKS; classified into type I, II and III) catalyzed linkage of short-chain activated fatty acids such as acetyl CoA, propionyl CoA, (methyl)malonyl CoA and butyryl CoA.⁶ Several Claisen-type reactions for chain elongation followed by additional transformations provide a great variety of polyketides (secondary metabolites).

The introductorily mentioned PKS, responsible for the assemblage of the acyl CoA components, is a massive multienzyme complex and consists of several enzyme domains. The latter are grouped into modules, each responsible for an individual chain propagation step.⁷ The acyl transferase (AT) domain, which selects the acyl CoA extender unit, an acyl carrier protein (ACP) for binding and further reaction and the ketoacyl synthase (KS), which catalyzes the decarboxylative Claisen condensation are the minimum requirements of one extension module. In addition to the essential domains of the loading module ("load" in Figure 3), further modifications are realized in the extension modules (modules 1 to 8) by β -ketoacyl reductase (KR), β -hydroxyacyl dehydratase (DH), enoyl reductase (ER) and methyl transferase (MT) domains. The thioesterase (TE) domain (termination module) is located in the last module and catalyzes the release of the nascent polyketide chain from the enzyme complex with concomitant macrocyclization.

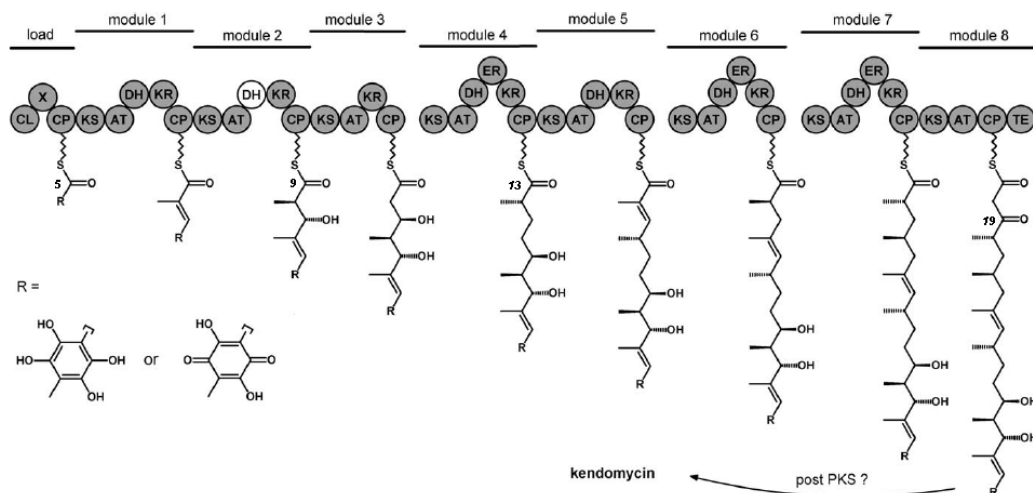
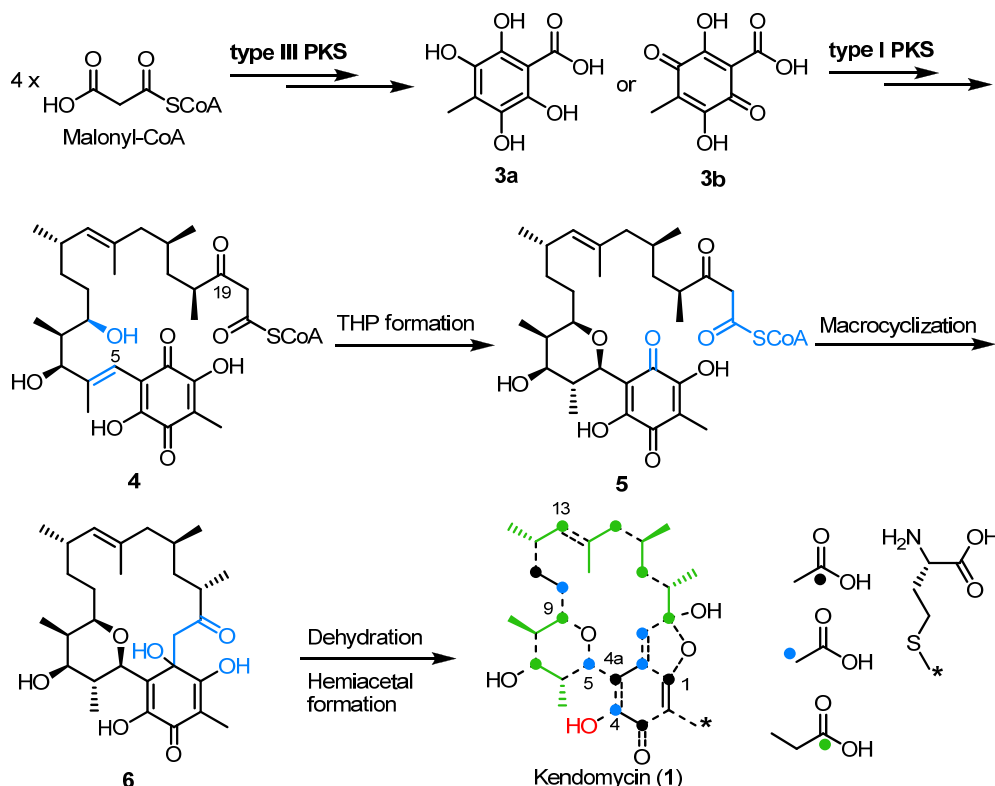


Figure 3. Type I PKS mediated biosynthesis of kendomycin (**1**) in a clockwise manner from C5 to C19. The unusual starter unit R (shown on the left) is provided from the type III PKS depended pathway shown on page 4 (reproduced with minor alterations from ref. 8). CL = CoA ligase, X = so far unknown quinone formation domain.

⁶ F. J. Leeper, J. C. Vederas, Eds., *Biosynthesis: Aromatic Polyketides, Isoprenoids, Alkaloids* (Topics in Current Chemistry), Springer, 1st edition **2000**, 1–53.

⁷ J. E. McMurry, T. P. Begley, *The Organic Chemistry of Biological Pathways*, Roberts & Company Publishers, **2005**, 369–378.

In contrast to other polyketides, an unusual loading unit and a biochemically unknown termination step are involved in the biosynthesis of **1**. Both, kendomycin type I PKS and type III PKS, act as a hybrid system during the biosynthesis.⁸ The biosynthesis of the benzoic acid starter unit **3**, which in turn is processed by a type I PKS, is addressed to the ACP independent type III PKS ("chalcon synthase").



Scheme 2. Biosynthesis and labeling pattern of **1**.

Condensation of four molecules of malonyl CoA (Scheme 2), incorporation of molecular oxygen to install the C4–OH functionality and further modifications give **3a** or/and **3b**, which in turn are activated by a CoA ligase (CL) domain and linked to the ACP domain of the type I PKS. After incorporation of eight extender units (six molecules of methylmalonyl CoA and two of malonyl CoA) the stage is set for termination. Pyran ring formation between the benzylic C5 carbon atom and the C9 hydroxyl group in **4** probably initiates the final steps. Compared to the usual macrolactonization step, a decarboxylative thioesterase (TE) mediates an aldol-like condensation to give the fully carbogenic *ansa*-macrocycle **6**. Finally, spontaneous dehydration and hemiacetal formation completes the biosynthesis of **1**, even

⁸ S. C. Wenzel, H. B. Bode, I. Kochems, R. Mueller, *ChemBioChem* **2008**, *9*, 2711–2721.

though the detailed biochemical mechanisms of the latter steps are not fully understood. Detailed labelling studies disclosed the origin of all oxygen and carbon atoms.²

2. Previous Synthetic Work

The pioneering work of the Mulzer group strongly influenced all syntheses and fragment preparations developed so far. Several synthetic strategies, which were directed by their preliminary results, in particular with regard to the observed atropisomerism, will be covered in the following chapters.

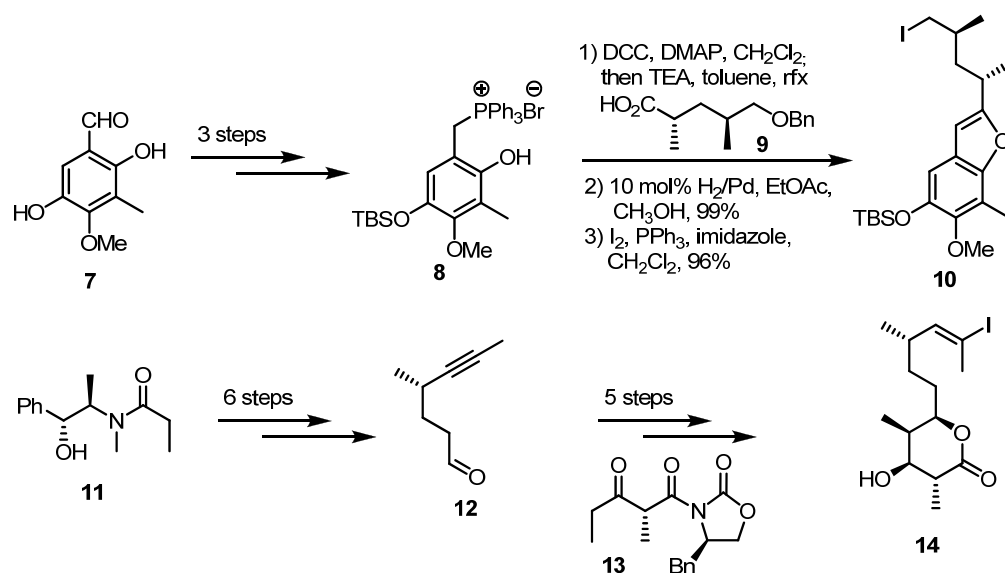
2.1. Reported Total Syntheses of Kendomycin

2.1.1. Lee's Total Synthesis of Kendomycin

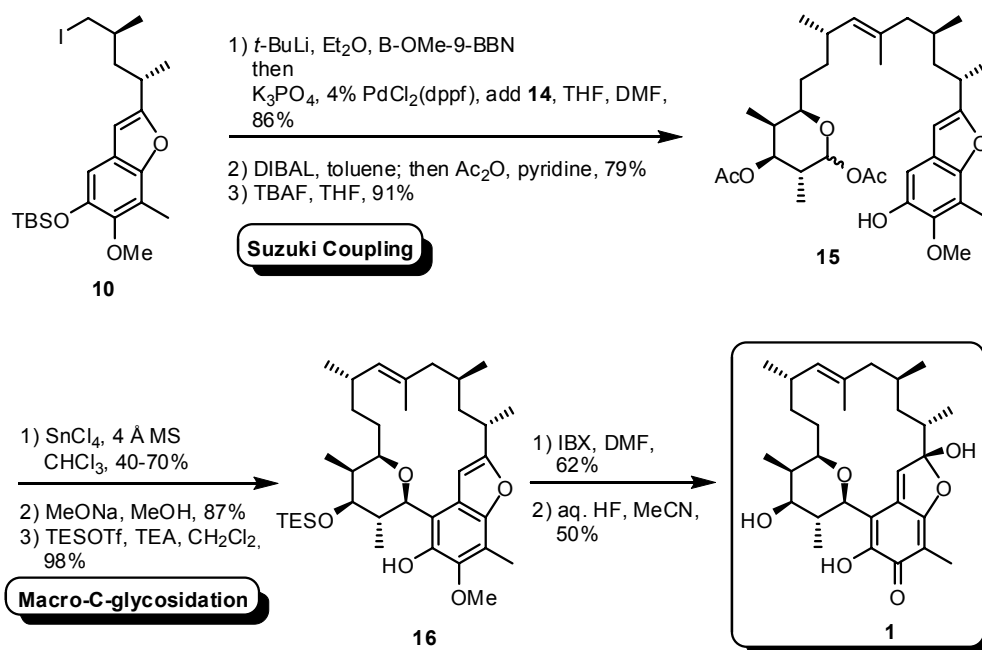
Strategy: Macro-C-Glycosidation: Avoiding the problem of atropisomerism.

In 2004, the first total synthesis of kendomycin was reported by Lee and co-workers.⁹ Their elegant and elaborated strategy focused on the Suzuki-Miyaura coupling partners **10** and **14** (Scheme 3), and furthermore key-intermediate **15** (Scheme 4), the macroglycosidation precursor. Installation of benzofuran **10** was accomplished by one pot esterification of phosphonium salt **8** with acid **9** and Wittig reaction, followed by hydrogenolysis and Appel reaction. Preparation of vinyl iodide **14** started with Myers' asymmetric alkylation of **11** to give, after five steps, aldehyde **12**. Elongation with Evans extended auxiliary **13** and further functional group manipulations afforded lactone **14** in eleven steps from compound **11**.

⁹ Y. Yu, H. Men, C. Lee, *J. Am. Chem. Soc.* **2004**, *126*, 14720–14721.

Scheme 3. Lee's syntheses of the eastern and western portion of **1**.

A high yielding three steps sequence, including the Suzuki–Miyaura coupling of **10** and **14** gave rapid access to phenol **15** (Scheme 4). Formation of an intermediate O–glycoside and subsequent rearrangement to the C–glycoside with SnCl_4 provided, after exchange of the secondary acetate, sufficient amounts of macrocycle **16**. All efforts to close the macrocycle by Friedel–Crafts acylation with the corresponding TBS protected phenol failed.



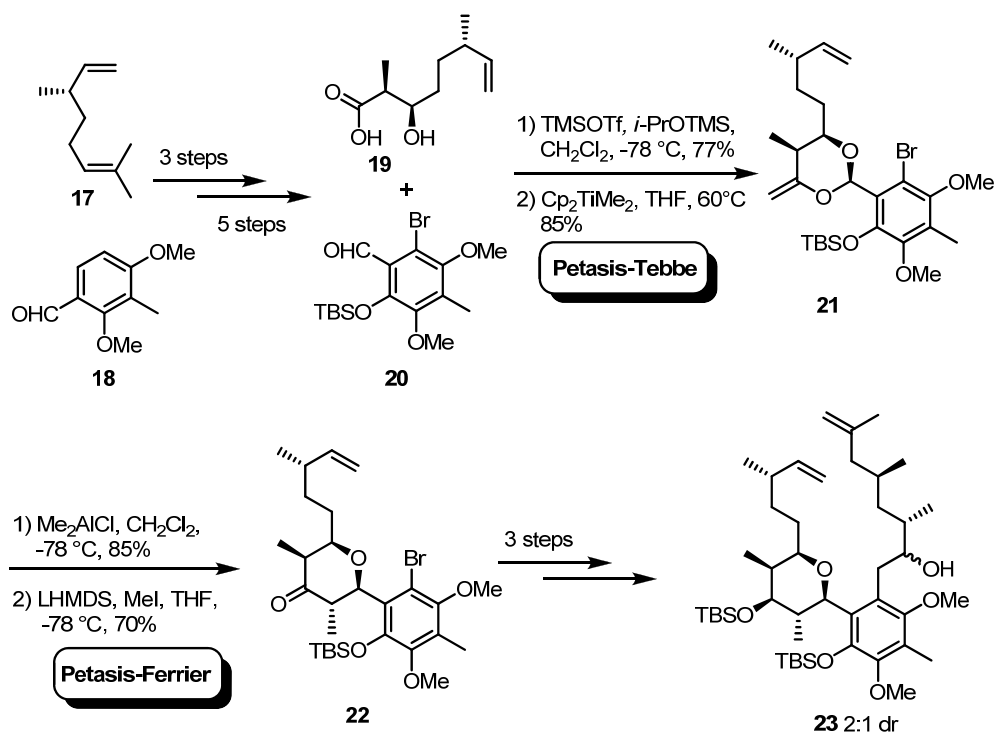
Scheme 4. The key-steps of Lee's synthesis.

Formation of the unstable *o*-quinone chromophor with IBX, followed by one pot deprotection and hydrolysis with aqueous HF gave **1** in 20 linear steps.

2.1.2. Smith's Total Synthesis of Kendomycin

Strategy: Petasis–Ferrier rearrangement and RCM: The hunt for the (*E*)-double bond geometry.

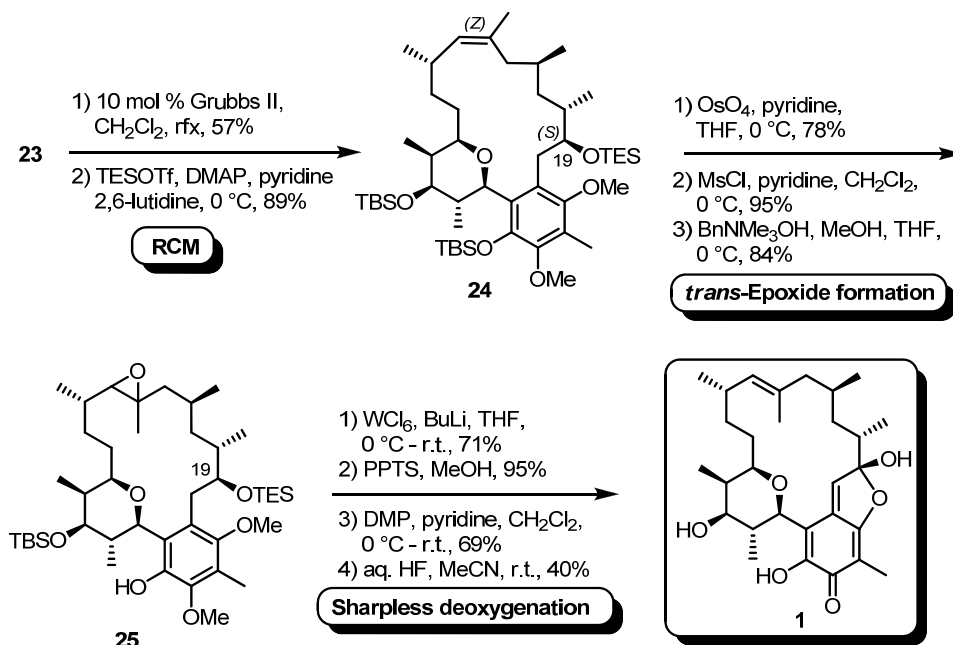
The second total synthesis of (-)-**1**, relying on a more conventional approach, was achieved by Smith and co-workers in 2005.¹⁰ The synthetic plan focused on a Petasis–Ferrier rearrangement to construct the tetrahydropyran unit and a RCM reaction to build up the macrocyclic skeleton. Despite the negative RCM results from the Mulzer group (see page 16) they were convinced, that TBS protected phenol **23** would be a superior and more productive RCM precursor (Scheme 5). Preparation of enol acetal **21** started with the condensation of β -hydroxy acid **19** and benzaldehyde **20**. The so obtained dioxanone intermediate was converted to **21** via Petasis–Tebbe methylenation. Lewis acid induced Petasis–Ferrier rearrangement followed by diastereoselective methylation, presumably via a skewed-boat-like transition state (TS), generated tetrahydropyran **22**. After three more steps the stage was set for the envisaged RCM reaction.



Scheme 5. Smith's approach to RCM precursor **23**.

¹⁰ A. B. III Smith, E. F. Mesaros, E. Meyer, *J. Am. Chem. Soc.* **2005**, *127*, 6948–6949.

Careful experimentation showed, that on treatment of the 2:1 diastereomeric mixture **23** with Grubbs second generation catalyst, only the major (*S*)-alcohol participated in the RCM reaction (Scheme 6). The unexpected and undesired formation of the (*Z*)-olefin geometry required a rather laborious, four steps isomerization sequence of macrocycle **24**, relying on formation of *trans*-epoxide **25** and Sharpless deoxygenation with tungsten(IV).



Scheme 6. Smith's final steps towards **1**: Macrocyclization and double bond isomerization.

Selective desilylation and Dess–Martin oxidation of the secondary C19 alcohol as well as the phenol gave an intermediate *o*-quinone, which was hydrolyzed to **1**. The second total synthesis of **1** was accomplished with a longest linear sequence of 21 steps and with an overall yield of 0.49%.

2.1.3. Panek's Total Synthesis of Kendomycin

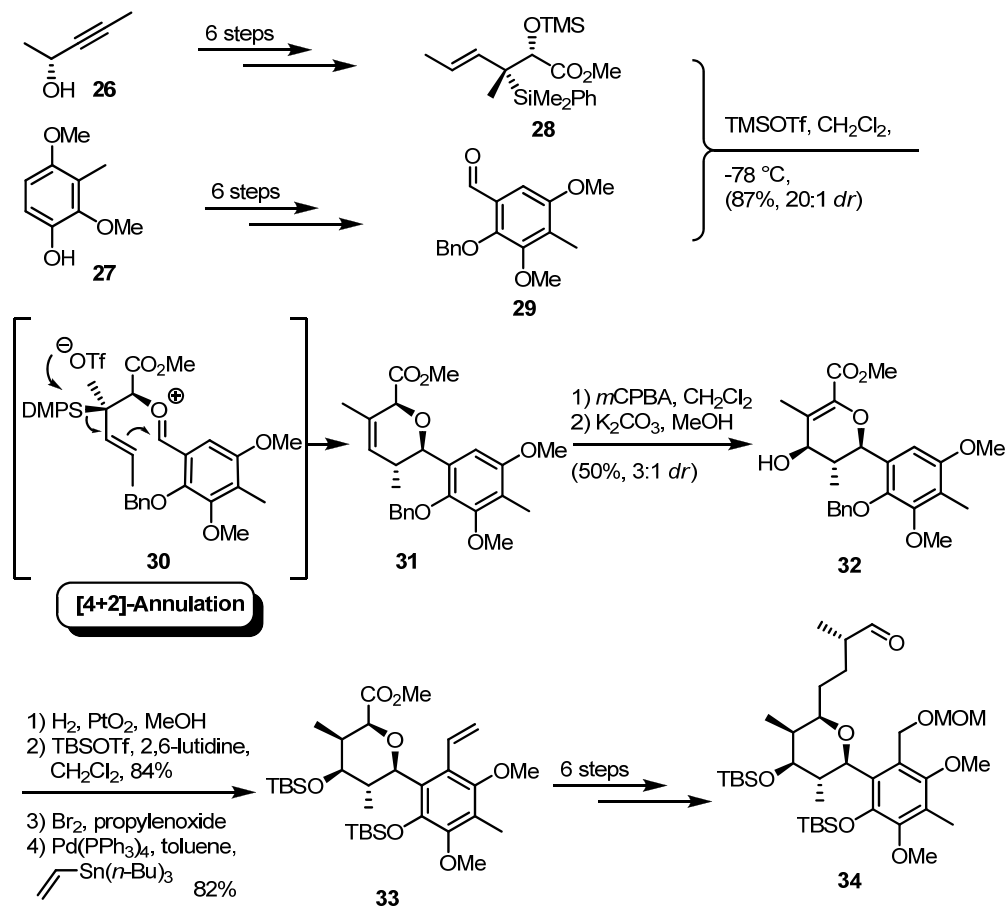
Strategy: [4+2]–Annulation and intramolecular Barbier–type reaction: A linear approach.

Panek's stereocontrolled [4+2]–annulation strategy¹¹ in combination with an intramolecular Barbier–type cyclization culminated in the third total synthesis of kendomycin (Scheme 7).¹² *Syn*–selective [4+2]–annulation of (*E*)-vinyl silane **28** with benzaldehyde **29** gave the desired dihydropyran **31** in good yield and diastereoselectivity. Epoxidation and regioselective base induced epoxide opening delivered

¹¹ J. T. Lowe, J. S. Panek, *Org. Lett.* **2005**, *7*, 1529–1532.

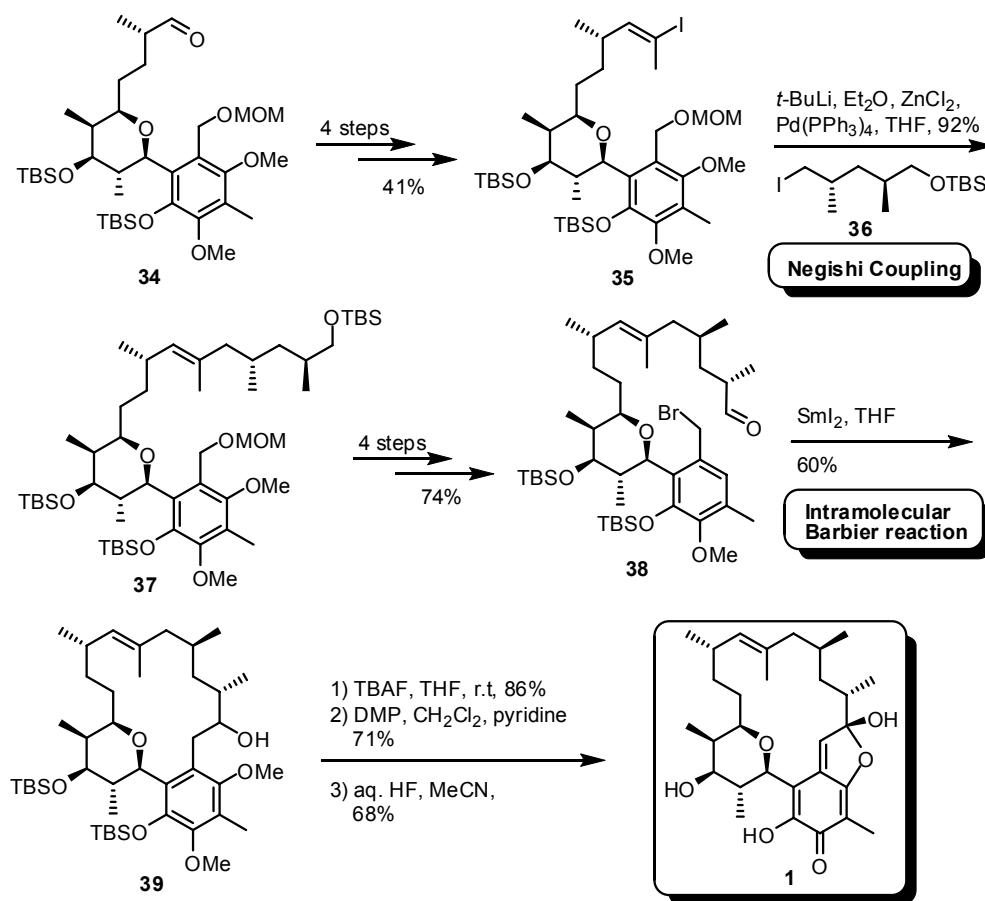
¹² J. T. Lowe, J. S. Panek, *Org. Lett.* **2008**, *10*, 3813–3816.

intermediate **32**. Further functional group manipulations and introduction of the styrene unit by a Stille coupling gave the fully substituted arene-THP fragment **33**. Aldehyde **34** was obtained after additional six steps.



Scheme 7. Panek's [4+2]-annulation strategy.

Negishi type coupling of vinyl iodide **35**, easily obtained from **34** in four steps, and alkyl iodide **36** provided the complete kendomycin carbon scaffold **37** (Scheme 8). Release of the aldehyde and the benzylbromide function within four steps paved the way for the final sequence.



Scheme 8. Completion of Panek's total synthesis.

Intramolecular SmI_2 -mediated Barbier macrocyclization of **38** produced secondary alcohol **39** in good yield. Selective cleavage of the phenolic TBS protecting group, oxidation and subsequent treatment with aqueous HF resulted in complete deprotection with simultaneous formation of **1**.

A major drawback is the high linearity of the synthesis, which proceeds in 32 steps (1.1% overall yield) from alcohol **26**.

2.2. Reported Formal Syntheses of Kendomycin

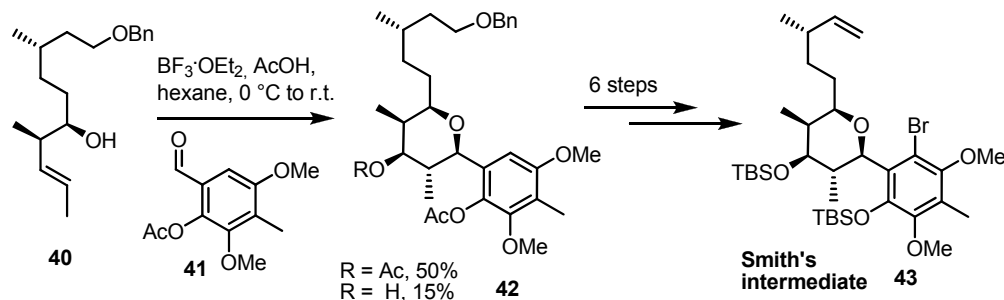
2.2.1. Rychnovsky's Formal Syntheses of Kendomycin

Strategy: Intramolecular-Prins cyclization: Low selectivity for the desired macrocycle.

The key steps of Rychnovsky's formal syntheses of kendomycin (**1**), published in 2006¹³ and 2008¹⁴, were both based on Prins-reactions. The desired Lewis acid ($\text{BF}_3\text{-OEt}_2$) promoted intermolecular cyclization of

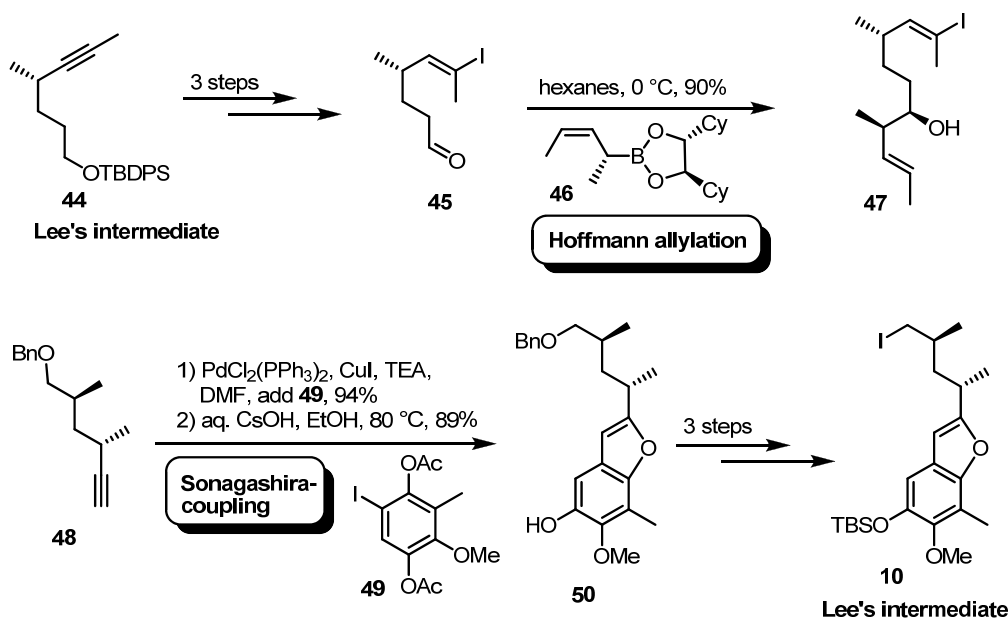
¹³ K. B. Bahnck, S. D. Rychnovsky, *Chem. Commun.* **2006**, 2388–2390.

homoallylic alcohol **40** with benzaldehyde **41** was achieved after some optimization (Scheme 9). THP fragment **42** was obtained in moderate yield and transformed to bromide **43** within six steps, a fragment of Smith's total synthesis.



Scheme 9. Rychnovsky's first formal synthesis.

In 2008, they revisited the Prins–cyclization and extended it to an intramolecular version (Scheme 10). Homoallylic alcohol **47** was obtained in four steps from known compound **44** via Hoffmann's allylation of vinyl iodide **45**. The modified synthesis of Lee's intermediate **10** started from alkyne **48** by Sonagashira coupling with iodide **49** and subsequent base induced 5–*endo*-dig cyclization to benzofuran **50**. A three step procedure was then necessary to complete the synthesis of **10**.

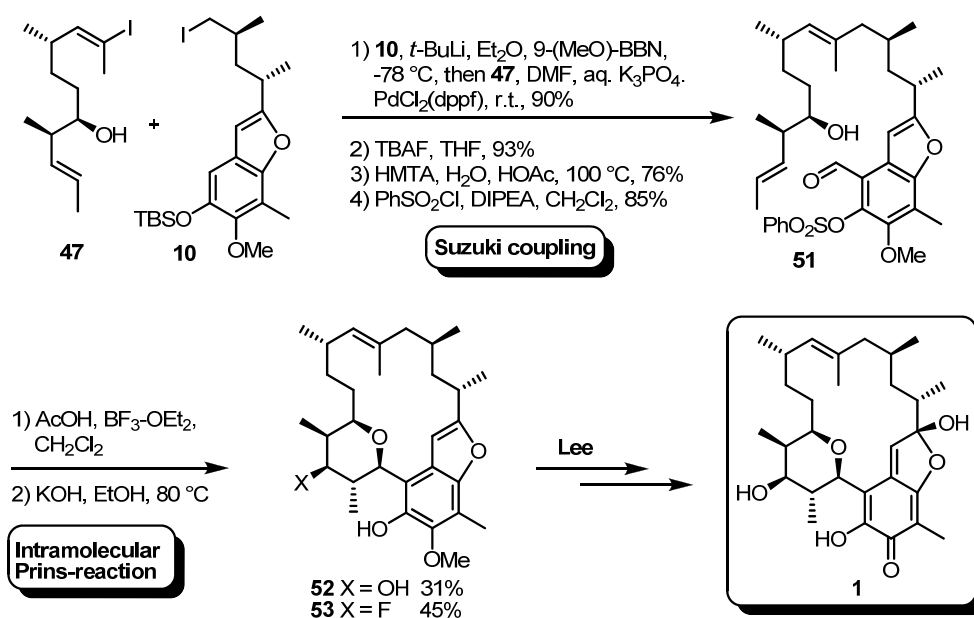


Scheme 10. Alternative synthesis of Lee's right-hand portion of **1**.

¹⁴ K. B. Bahnck, S. D. Rychnovsky, *J. Am. Chem. Soc.* **2008**, *130*, 13177–13181.

Linkage of **47** and **10** was achieved via Lee's established Suzuki Miyuara coupling sequence (Scheme 11). Desilylation, Duff formylation and installation of an electron withdrawing benzenesulfonyl protecting group, essential for efficient cyclization, provided the desired Prins precursor **51**.

Acid induced intramolecular cyclization under high dilution conditions followed by saponification gave a mixture of Lee's intermediate **52** and the corresponding fluorine derivative **53**. However, due to the unfavorable ratio of **52** and **53**, the synthetic practicability seems to be rather restricted. Efforts to reproduce Lee's final steps failed.



Scheme 11. Rychnovsky's formal synthesis via intramolecular Prins-cyclization.

A chronological summary of all total and formal syntheses reported so far is given in the following table. Lee's elegant synthesis is outstanding and the most efficient one published so far. Our second synthesis is just four steps longer and features a higher overall yield compared to Smith's synthesis. Panek's synthesis from 2008 features interesting chemistry but a major drawback is the lack of convergence. Rychnovsky's first formal synthesis, reported in 2006, describes only a very basic building block from Smith's synthesis.

| | Total Syntheses | | | Formal Syntheses | |
|------------|-----------------|------|--------|------------------|-------|
| | Year | LLS | Yield | LLS | Yield |
| Lee | 2004 | 20 | 1.3% | | |
| Smith | 2005 | 21 | 0.5% | | |
| Panek | 2008 | 32 | 1.1% | | |
| Rychnovsky | 2008 | (20) | (2.6%) | 17 | 8.8% |
| Rychnovsky | 2006 | (22) | (0.5%) | 12 | 13.5% |
| Own Work | 2009 | 32 | 0.4% | | |
| Own Work | 2009 | 24 | 1.0% | | |

Table 1. Overview about the reported total and formal syntheses of (-)-kendomycin. The number of steps and quoted yields were determined by starting from commercially available starting materials. LLS = Longest linear sequence.

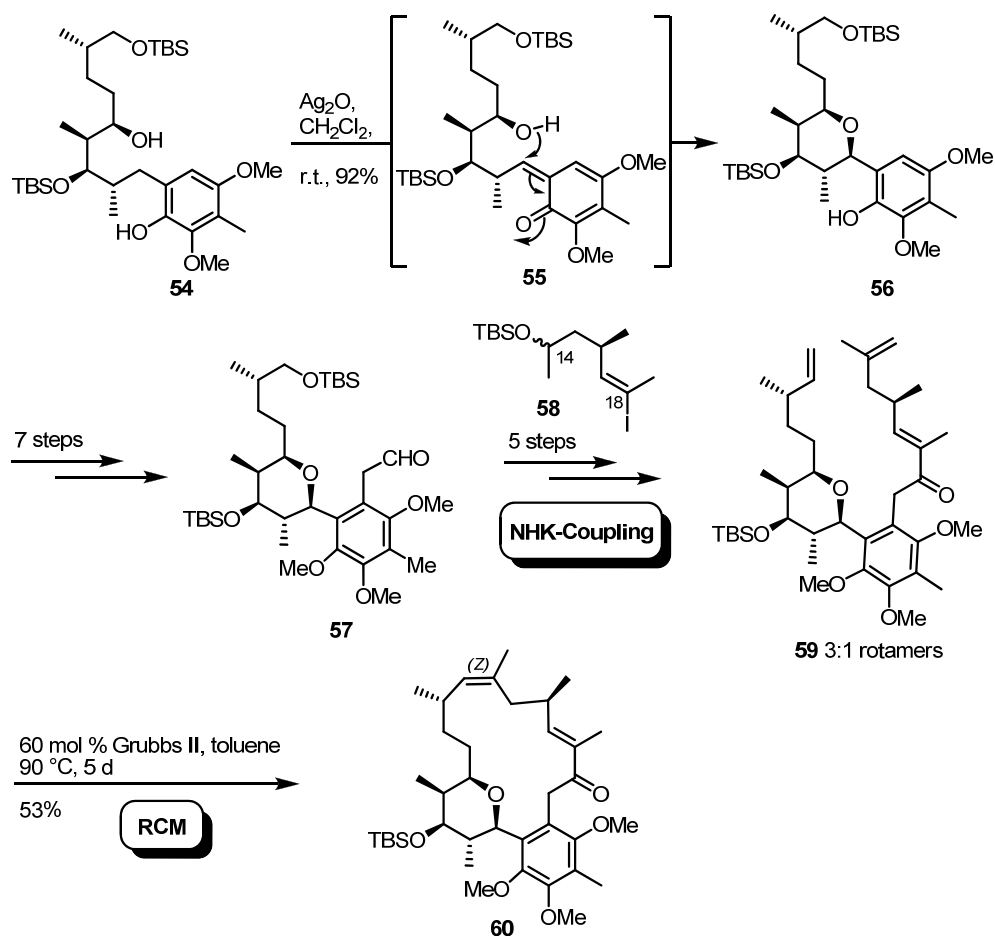
2.3. Fragment Preparations of Kendomycin

2.3.1. Arimoto's Approach to the Macrocyclic Core of Kendomycin

Strategy: RCM: Problems similar to Smith and Mulzer.

Arimoto's synthesis of the south-western domain of kendomycin was already described in 2004 (Scheme 12).¹⁵ Preparation of intermediate **54** was accomplished in 18 linear steps via standard chemistry. A silver(I) promoted oxidative cyclization of **54**, presumably involving *o*-quinone methide **55**, delivered the desired THP moiety in excellent yield. Transformation of homobenzylic aldehyde **57** to pentasubstituted arene **56** was realized in another seven steps.

¹⁵ T. Sengoku, H. Arimoto, D. Uemura, *Chem. Commun.* **2004**, 10, 1220–1221.

Scheme 12. Arimoto's synthesis of a major portion of **1**.

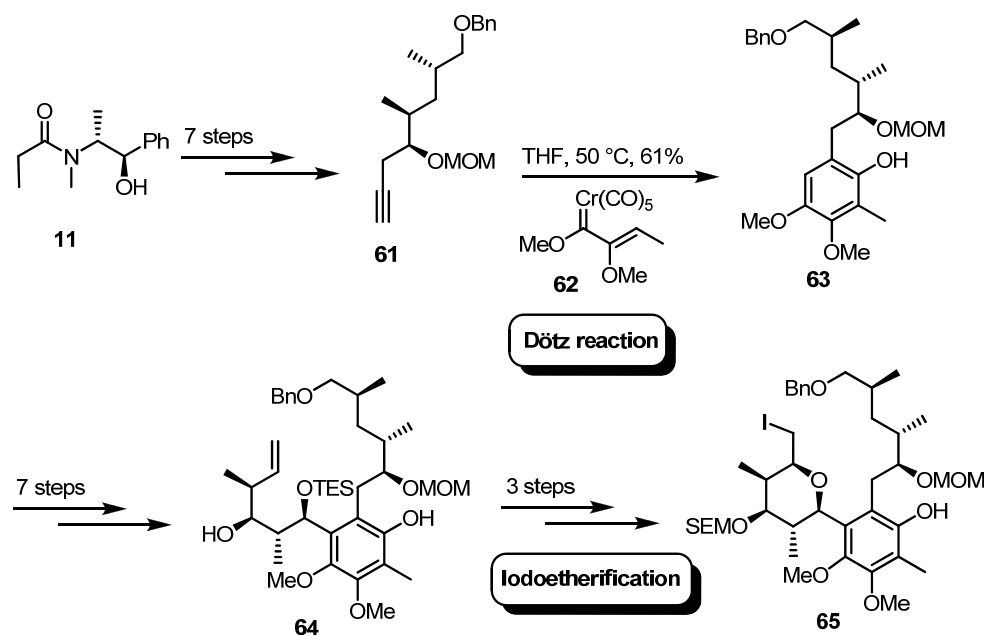
Linkage of the eastern C14–C18 domain, which was described in 2007, consumed additional five synthetic operations, whereas the key-step consisted of a Nozaki–Hiyama–Kishi reaction between **57** and **58**.¹⁶ Subjecting the 3:1 rotameric mixture (the kendomycin like rotamer proved to be favored) of **59** to a variety of RCM conditions was disappointing. A substoichiometric amount of Grubbs' second generation catalyst (60 mol %) and long reaction times were necessary to construct the undesired (*Z*)-configured macrocycle **60**. Apart from this drawback, a 35 steps sequence to **60** would not necessarily facilitate the synthesis of **1**.

2.3.2. White's Approach to Kendomycin

Strategy: Dötz–annulation reaction: *De novo* synthesis of the aromatic core.

¹⁶ T. Sengoku, D. Uemura, H. Arimoto, *Chem. Lett.* **2007**, 36, 726–727.

White's group reported the *de novo* synthesis of kendomycin's arene core via the Dötz annulation reaction in 2005 (Scheme 13).¹⁷ A high yielding seven steps sequence, starting from Myers' *N*-propionylpseudoephedrine **11**, lead to alkyne **61**. Preparation of the Fischer-type alkenylchromium carbenoid **62** was realized in three steps from 1-methoxyprop-1-yne. Highly regioselective Dötz annulation, followed by oxidative workup provided intermediate **63** in good yield. Transformation of **63** to **64**, which possesses the fully functionalized arene core, was achieved by seven synthetic operations and paved the way for the owing THP formation. The three remaining steps were realized by initial protecting group manipulations and final iodocyclization to give **65**.



Scheme 13. White's Dötz annulations strategy.

2.3.3. Williams' Synthesis of Kendomycin's *Ansa*-Chain

Strategy: Asymmetric conjugate addition: Linear construction of the *ansa*-chain.

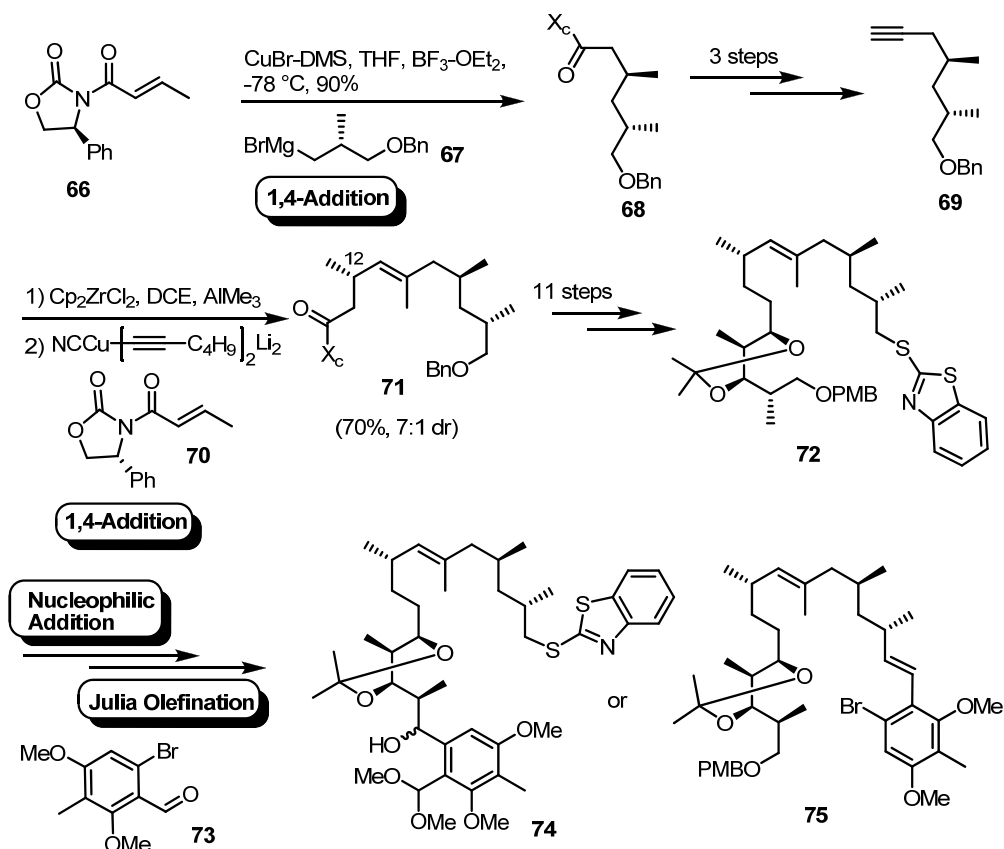
An auxiliary based approach of the *ansa*-chain was disclosed by Williams in 2005 (Scheme 14).¹⁸ Conjugate addition of **67** to *N*-enoyl oxazolidine **66** gave the 1,3-*anti*-fragment **68**. Negishi's *syn*-carboalumination protocol followed by *in situ* transmetalation with an higher-order cuprate set the stage for the second 1,4-addition. The application of **70** secured the diastereoselective installation of

¹⁷ J. D. White, H. Smits, *Org. Lett.* **2005**, *7*, 235–238.

¹⁸ D. Williams, K. Shamim, *Org. Lett.* **2005**, *7*, 4161–4164.

the C12 methyl group. Additional eleven steps from **71** were necessary to create the fully substituted *ansa*-chain **72**.

Connection to the aromatic moiety was accomplished via two pathways. First, nucleophilic addition of lithiated arene **73**, which was protected as its dimethoxyacetale prior to halogen-metal exchange, delivered **74** in three steps from **72**. Second, (*E*)-selective Julia olefination of benzaldehyde **73** with **72** derived sulfone provided **75** in two steps and excellent yield.



Scheme 14. Williams' approach of the fully substituted *ansa*-chain.

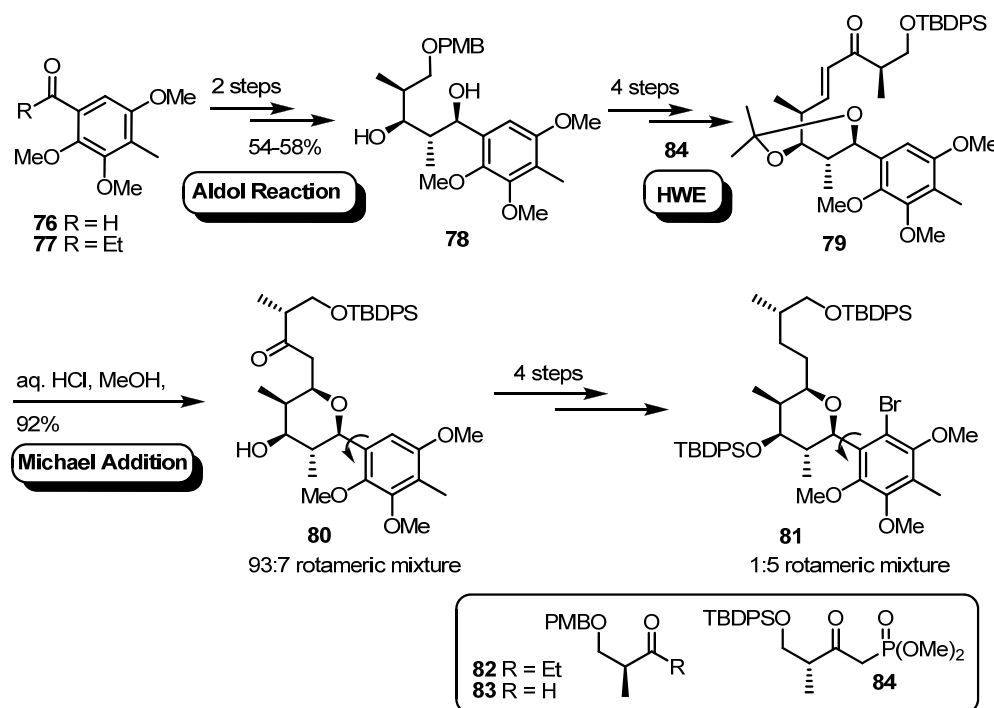
2.4. Mulzer's Approaches to Kendomycin

2.4.1. 1st Generation Approach: Michael-Addition for the Preparation of the THP Ring

The first generation approach culminated in the synthesis of the south-west domain of kendomycin (**1**).¹⁹ The preparation of diol **78** was either achieved via *anti*-selective Peterson (**76** and **82**, Cy₂BCl) or

¹⁹ H. J. Martin, M. Drescher, H. Kählig, S. Schneider, J. Mulzer, *Angew. Chem., Int. Ed.* **2001**, *40*, 3186–3188.

syn-selective titanium enolate aldol reaction (**77** and **83**, $\text{Ti}(\text{O}i\text{Pr})_3\text{Cl}$) and subsequent stereoselective Evans–Carreira 1,3-*anti* reduction (Scheme 15). Four additional steps delivered α,β -unsaturated ketone **79**, which upon treatment with HCl cyclized to the desired tetrahydropyran **80**. Careful 2D-NMR experiments with **80** and **81** disclosed the first aryl-C-glycoside atropisomerism, which was depended on the degree of arene substitution. NOE experiments confirmed the preferred kendomycin-like orientation of **80** and revealed that in **81** the opposite rotamer is predominant. The latter resulted in great steric hindrance of the bromide function and all efforts to install the western side chain failed.



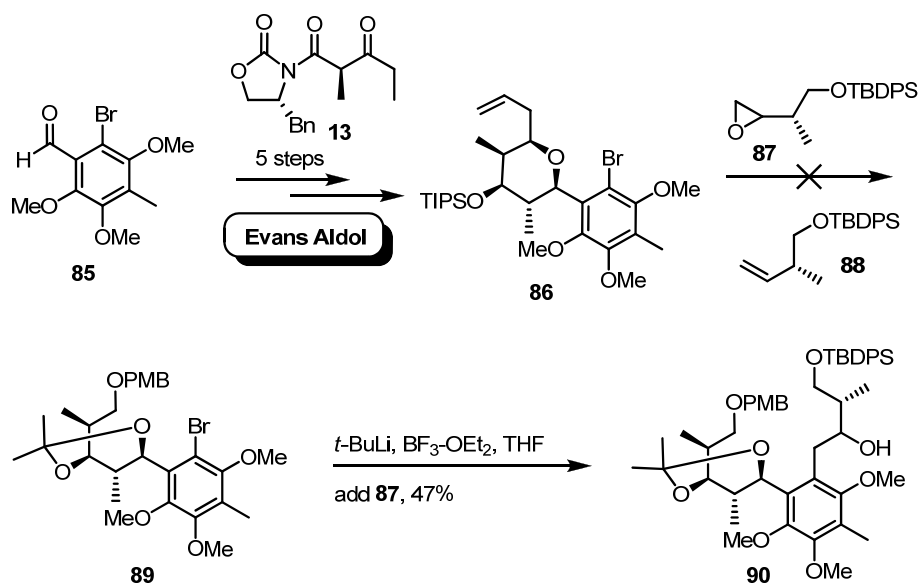
Scheme 15. Mulzer's 1st generation approach.

2.4.2. 2nd Generation Approach: Preparation of the *hexa*-Substituted Arene Core & Improvement of Tetrahydropyran Formation

Installation of the fully substituted arene **85** prior to THP formation revealed the Peterson *anti*-aldol chemistry to be low diastereoselective.²⁰ This problem could be solved by the well established extended Evans' aldol methodology. In that manner, the preparation of the fully substituted tetrahydropyran **86** was achieved in 5 steps from **85** with excellent diastereoselectivity (Scheme 16). Albeit all attempts to connect the side chains **87** and **88**, either via epoxide opening or via Heck coupling, to the rotameric mixture **86** failed, a moderate yield of the hexasubstituted arene **90** could be obtained with metallated **89**.

²⁰ M. M. B. Marques, S. Pichlmair, H. J. Martin, J. Mulzer, *Synthesis* **2002**, 2766–2770.

The former results and additional experiments strongly suggested the installation of the eastern domain prior to the western side chain.

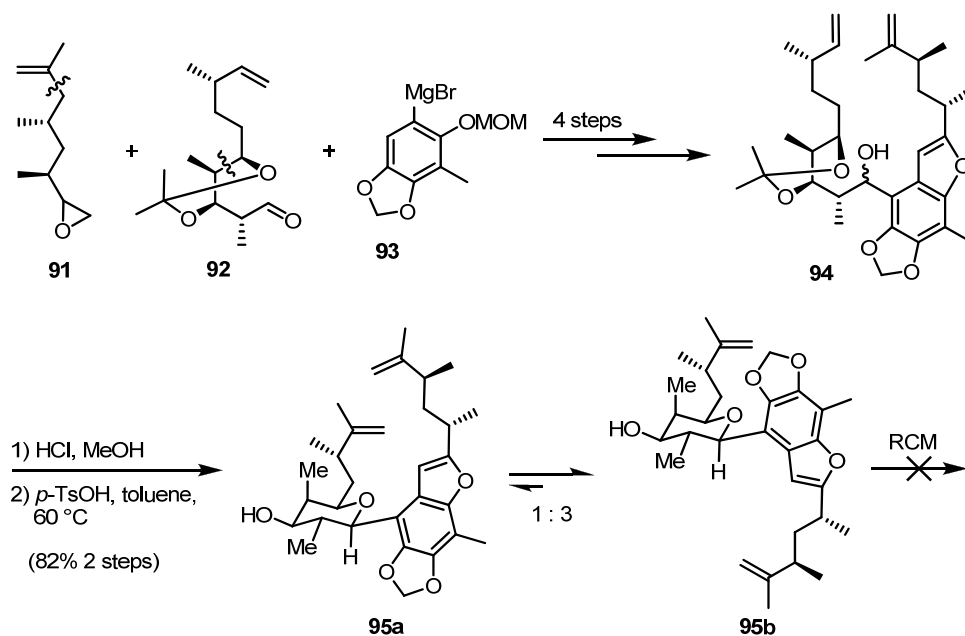


Scheme 16. Mulzer's 2nd generation approach.

2.4.3. 3rd Generation Approach: Preparation of the Full Carbon Skeleton and Attempted RCM

In 2003, a revised and superior strategy particularly with regard to prior benzofuran formation lead to highly advanced precursor **94** (Scheme 17).²¹ The convergent and concise synthesis focused on well elaborated building blocks **91**, **92** and **93**. Epoxide **91**, which was easily prepared via auxiliary chemistry and Schlosser–Fouquet coupling to introduce the isopropenyl functionality, and Grignard **93** served for the right hand portion of kendomycin. Aldehyde **92** was built up by Evans *syn*-aldol methodology and chiral pool building block β -(+)-citronellene. Linkage of **91** to **93**, followed by benzofuran formation and connection of aldehyde **92** via an *o*-lithiation – nucleophilic addition sequence, provided benzylic alcohol **94** in four steps. Deprotection and $\text{S}_{\text{N}}1$ -cyclization afforded a 1:3 rotameric mixture of **95a** and **95b**, whereas the disadvantageous rotamer **95b** prevailed. Not surprisingly, intermediate **95** did not undergo the desired RCM reaction.

²¹ S. Pichlmair, M. M. B. Marques, M. P. Green, H. J. Martin, J. Mulzer, *Org. Lett.* **2003**, *5*, 4657–4659.

Scheme 17. The 3rd generation approach.

In order to circumvent the unfavorable atropisomerism, which seemed to be the major hindrance for successful cyclization, several analogs were designed and tested for the RCM reaction (Figure 4). Intermediates **96** to **98** were prepared in an analogous manner to that of **94** (for **98** compare **101**). The desired RCM reaction was never observed, albeit the conformation of **98** (R = MOM) was fixed in a kendomycin-like orientation.²²

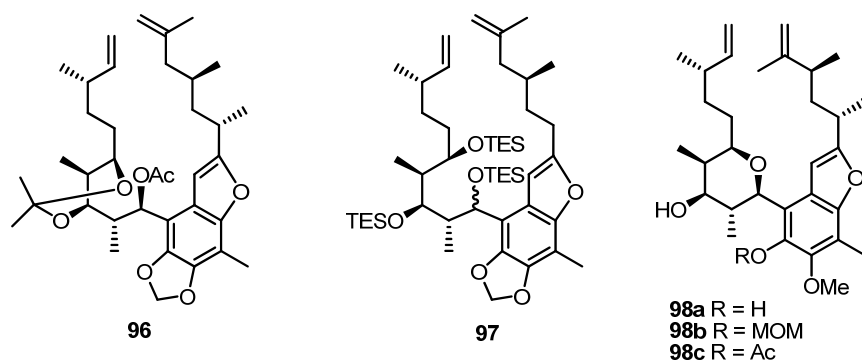
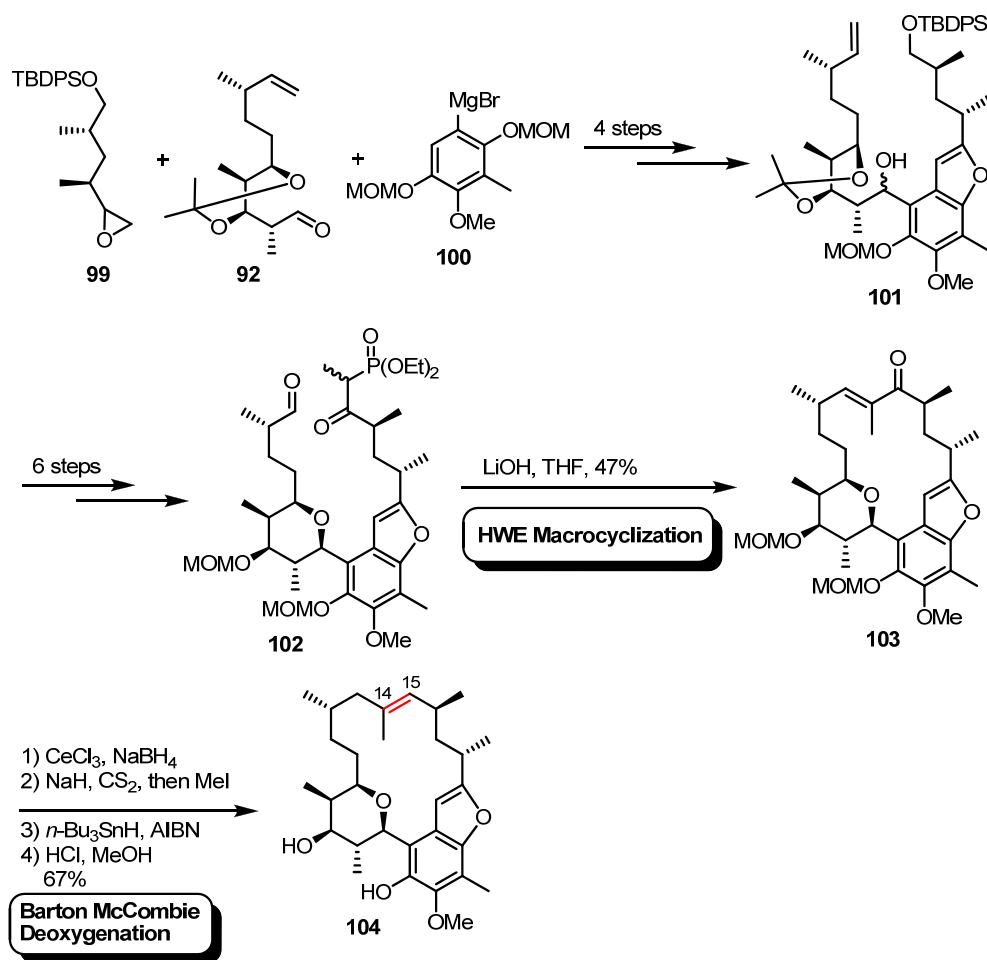


Figure 4. Alternative RCM precursors.

²²J. Mulzer, S. Pichlmair, M. P. Green, M. M. B. Marques, H. J. Martin, *Proc. Nat. Acad. Sci.* **2004**, *101*, 11980–11985.

2.4.4. 4th Generation Approach: Macrocyclisation via Horner–Wadsworth–Emmons Reaction

Since RCM macrocyclization proved to be impossible for several benzofuran analogs, it was time for a new cyclization strategy.²² The construction of more versatile intermediate **101** was executed in a similar manner to that of **94** (Scheme 18). Finally, the reliable intramolecular Horner–Wadsworth–Emmons (HWE) reaction showed to be the method of choice and preparation of β -keto phosphonate **102** could be performed straight forward from **101**. Indeed, on treatment with LiOH, **102** smoothly cyclized to ketone **103**.



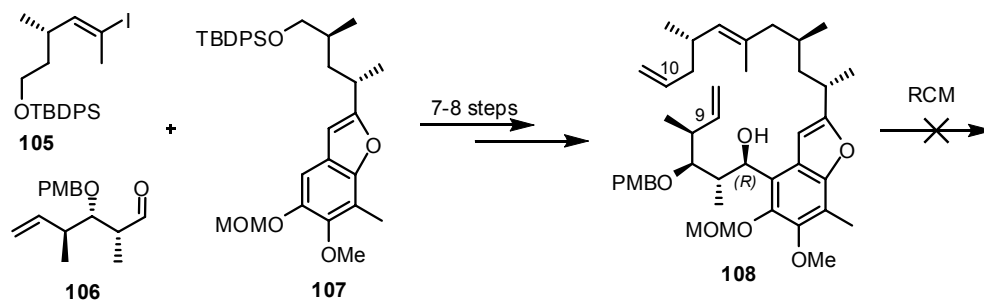
Scheme 18. The 4th generation approach: Successful macrocyclization.

After numerous unsuccessful attempts to remove the undesired ketone function in **103**, deoxygenation was achieved with Barton McCombie's protocol. Unfortunately, extensive 2D-NMR studies of **104** showed that the double bond entirely migrated to the C14–C15 position. Having arrived at the most

disappointing stage of the synthesis they decided to give up the ring closure between C13 and C14 completely.

2.4.5. 5th Generation Approach: RCM – Etherification Approach

Disconnection of the C9–C10 bond was the key strategy of the fifth generation approach, which is described in more detail in the appended full paper.²³



Scheme 19. The 5th generation approach. Attempted RCM between C9 and C10.

²³ (a) T. Magauer, H. J. Martin, J. Mulzer, *Chem. Eur. J.* **2009**, *Accepted*; (b) T. Magauer, *Towards the Total Synthesis of the ansa-Macrocycle Kendomycin*, Diploma Thesis, University of Vienna, **2007**.

3. Results

3.1. Total Synthesis of the Antibiotic Kendomycin by Macrocyclization via Photo-Fries Rearrangement and Ring Closing Metathesis (RCM)

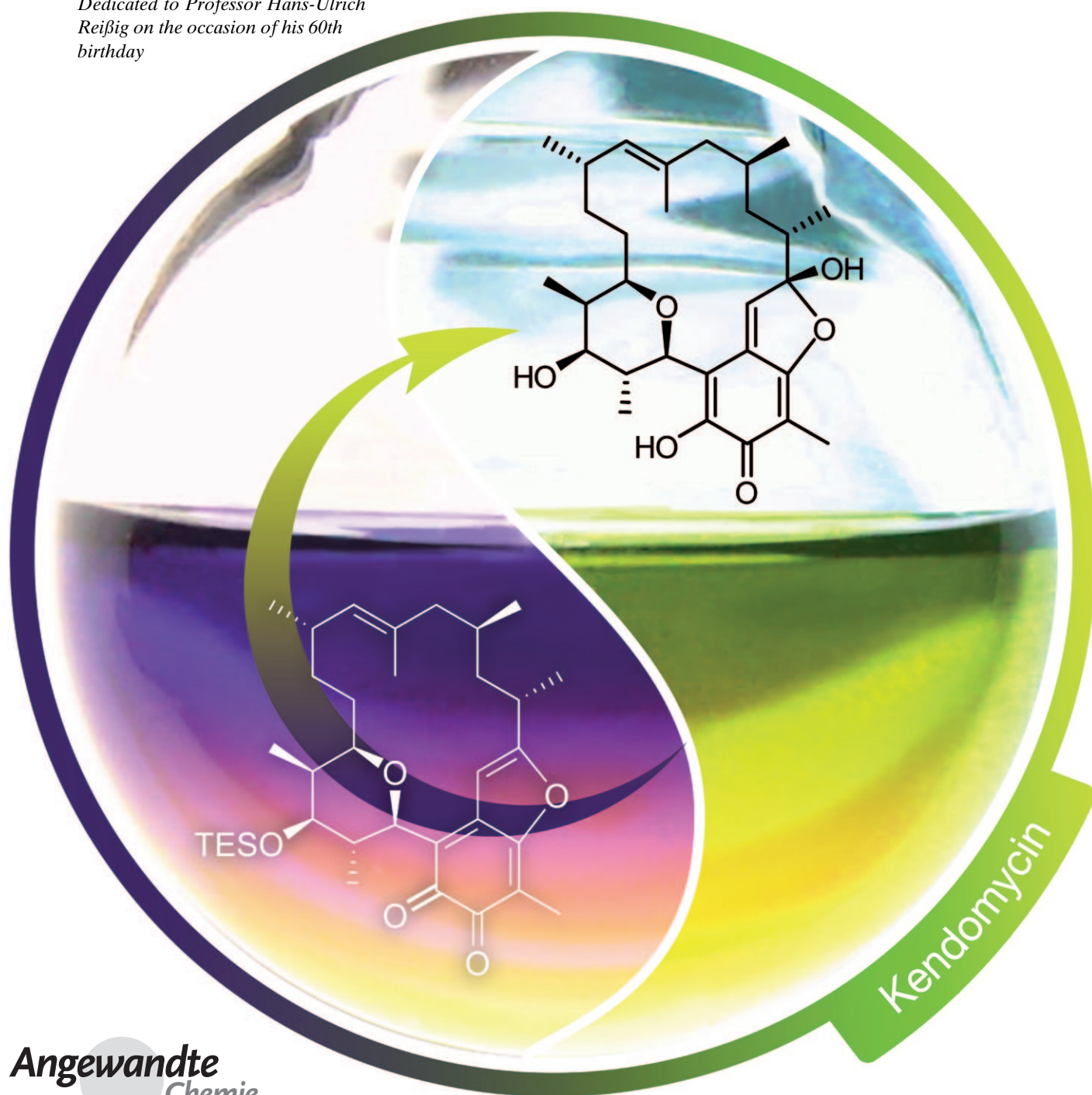
T. Magauer, H. J. Martin, J. Mulzer, *Angew. Chem., Int. Ed.* **2009**, 48, 6032–6036.

The supporting information was omitted in order to avoid redundancy. For detailed experimental procedures and NMR data see SI of the full paper (section 3.2).

Total Synthesis of the Antibiotic Kendomycin by Macrocyclization using Photo-Fries Rearrangement and Ring-Closing Metathesis**

Thomas Magauer, Harry J. Martin, and Johann Mulzer*

*Dedicated to Professor Hans-Ulrich
Reiig on the occasion of his 60th
birthday*



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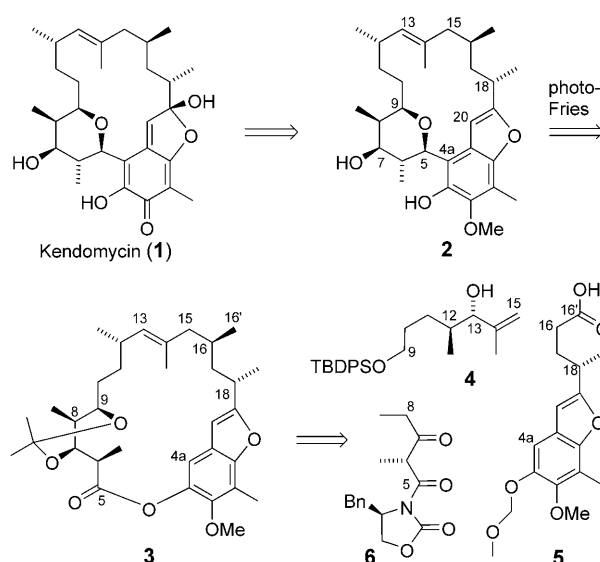
Angew. Chem. Int. Ed. 2009, 48, 6032–6036

Kendomycin [**1**, (–)-TAN2162], an ansamycin isolated from different *Streptomyces* species, has been shown in studies over the last decade to be a potent endothelin receptor antagonist and antiosteoporotic compound with remarkable antibacterial and cytostatic activity.^[1] The challenging structure and diverse pharmacological profile of kendomycin^[1,2] has motivated us^[3] and, sometime later a number of other groups,^[4–6] to carry out studies towards its synthesis. To date, three total syntheses^[4] and one formal synthesis^[5] have been reported, along with a number of fragment preparations.^[6] The main problem for all the approaches has been the formation of the strained macrocyclic *ansa*-ring. For example, macrocyclizations were performed using C-glycosidation,^[4a] Barbier-type organometallic addition,^[4d] Prins reaction,^[5] and Horner–Wadsworth–Emmons olefination.^[6c] Most strikingly, all attempts to achieve 13,14-macrocyclization by ring-closing metathesis (RCM)^[4b,6a,e] were plagued by low yields and formation of the undesired 13,14-*Z*-olefin. We tested alternative locations for RCM connections; however to our disappointment, both the 9,10- and the 19,20-positions proved to be unsuited.^[7] Nonetheless, we were still convinced that RCM should be a highly serviceable tool for ring closure.

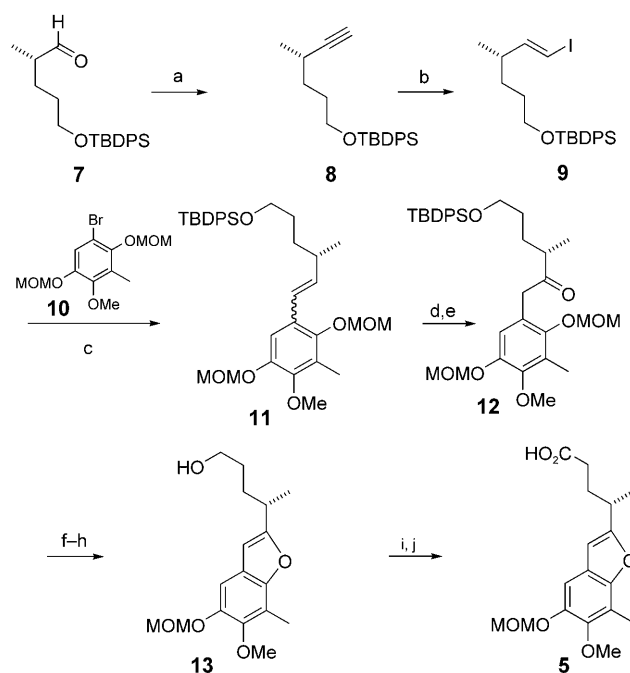
As we have demonstrated in previous studies,^[3,6e] hindered rotation around the C4a/C5 bond connecting the tetrahydropyran ring and the aromatic system makes ring closure difficult. Therefore, we decided to postpone tetrahydropyran formation until after macrocyclization, and consequently, we report herein two novel ring closures: the first by a photo-Fries ring contraction^[8] connecting C4a/C5, and the second by a RCM to form a 10,11-olefin. Both routes would lead to the known benzofuran intermediate **2**.^[4a]

The photo-Fries route (Scheme 1) centers around macro-lactone **3** as a key intermediate, which was assembled from building blocks **4**, **5**, and **6** by a Claisen–Ireland rearrangement (C15/C16 connection) and Evans aldolization (C8/C9-connection).

The synthesis of the benzofuran fragment **5** (Scheme 2) started with known aldehyde **7**,^[4a] which is easily available from citronellene (see the Supporting Information). A Colvin C₁ chain elongation^[9] furnished alkyne **8**, which was converted into vinyl iodide **9**. Negishi coupling^[10] with aryl bromide **10**^[6c] led to styrene **11**, which after epoxidation was subjected to palladium(0)-mediated rearrangement^[11] to ketone **12**. Acid-catalyzed formation of the furan ring concomitantly removed the 3-OMOM group, which was reinstalled. Desilylation delivered alcohol **13**, which was oxidized to carboxylic acid **5**.



Scheme 1. Retrosynthesis of **1**: Photo-Fries approach. TBDPS = *tert*-butyldiphenylsilyl, Bn = benzyl.



Scheme 2. Synthesis of carboxylic acid **5**. a) TMSCHN₂, *n*BuLi, THF, –78 °C to RT, 83%; b) [Cp₂ZrHCl], benzene, 50 °C; I₂, 0 °C, 76%; c) [Pd(PPh₃)₄], *t*BuLi, ZnCl₂, Et₂O/THF, 0 °C, 67%; d) DMDO, acetone, RT, 99% (d.r. 1.1:1); e) Pd(OAc)₂, PPh₃, *t*BuOH, reflux, 81% (2 steps); f) TfOH, toluene/EtOH (4:1), molecular sieves 4 Å, 80 °C, 5 min; g) MOMCl, NaH, DMF, 0 °C, 90% (2 steps); h) TBAF, THF, RT, 89%; i) IBX, DMSO, RT, 97%; j) NaClO₂/NaH₂PO₄, 2,3-dimethylbut-2-ene, *t*BuOH/H₂O, 99%. MOM = methoxymethyl, TMS = trimethylsilyl, Cp = cyclopentadienyl, DMDO = dimethyldioxirane, Tf = trifluoromethanesulfonyl, DMF = dimethylformamide, TBAF = tetra-*n*-butylammonium fluoride, THF = tetrahydrofuran, IBX = *o*-iodoxybenzoic acid, DMSO = dimethylsulfoxide.

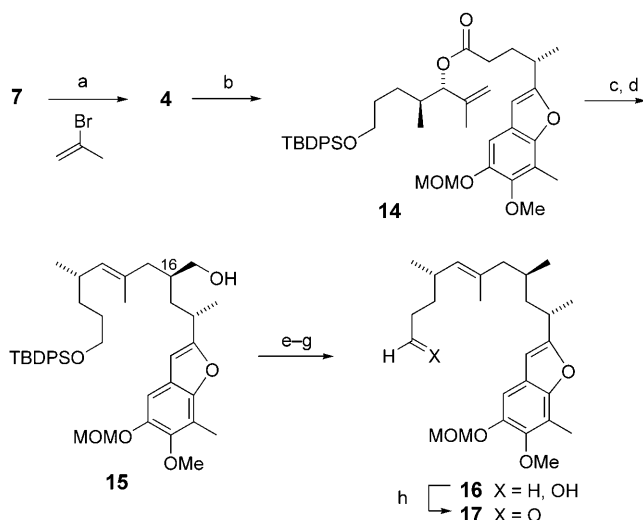
Allylic alcohol **4**,^[12] which was available from aldehyde **7** by Nozaki–Hiyama–Kishi addition^[13] of isopropenyl bromide

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[**] We thank Dr. Lothar Brecker and Susanne Felsing for NMR spectra, and Prof. A. Zeck for an authentic sample of **1**.

Supporting information for this article is available on the WWW under <http://dx.doi.org/10.1002/anie.200900522>.

(Scheme 3), was connected with carboxylic acid **5** to furnish ester **14** as the substrate of an Claisen–Ireland rearrangement, which, after reduction, led to primary alcohol **15** as an easily separable 4:1 diastereomeric mixture at C16.^[14] Reduction and desilylation gave alcohol **16**, which was oxidized to aldehyde **17**.

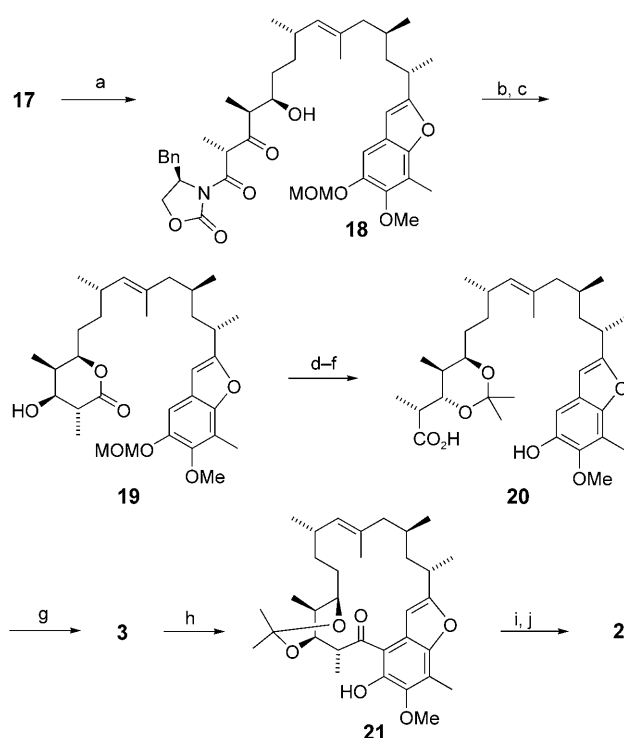


Scheme 3. Synthesis of aldehyde **17**. a) CrCl_2 (4 equiv), NiCl_2 (0.04 equiv), DMF, 0°C to RT, 86% (d.r. 1.4:1); b) EDCI, DMAP, **5**, CH_2Cl_2 , 85%; c) LHMDS, HMPA, TBSCl, -78°C to reflux; d) LiAlH_4 , Et_2O , 0°C , 84% (d.r. 4:1, 2 steps); e) MsCl , Et_3N , CH_2Cl_2 , 0°C ; f) LiAlH_4 , Et_2O , 0°C , 94% (2 steps); g) TBAF, THF, RT, 93%; h) IBX, DMSO, RT, 93%. EDCI = 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide, DMAP = 4-(dimethylamino)pyridine, HMDs = hexamethyldisilazane, HMPA = hexamethylphosphoramide, TBS = *tert*-butyldimethylsilyl, Ms = methanesulfonyl.

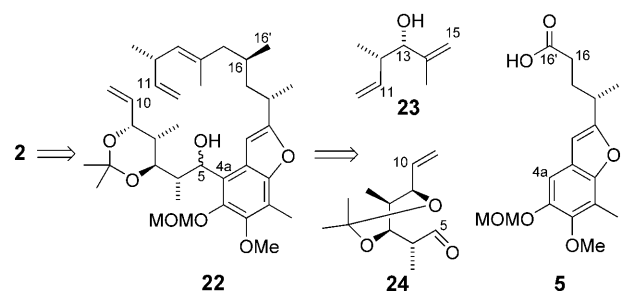
Aldol addition with the Evans ketoimide **6**^[15] furnished intermediate **18**, which has the full carbon skeleton of kendomycin. Reduction^[16] of the ketone and hydrolytic removal of the auxiliary led to lactone **19**, which was converted into *seco*-acid **20** by formation of the acetonide-protected methylester. Macrolactonization under modified Boden–Keck conditions^[17] gave monomer **3** in 55% yield, which underwent clean photo-Fries rearrangement to give ketone **21**. Reduction to the secondary alcohol, followed by removal of the acetonide and $\text{S}_{\text{N}}1$ cyclization, furnished key intermediate **2** (Scheme 4).

Our second route is outlined in Scheme 5. In contrast to earlier RCM attempts, RCM of triene **22** should not meet with major ring strain and thus proceed smoothly, as two monosubstituted olefins are connected. The tetrahydropyran ring is formed later, such that the above-mentioned atropisomerism cannot impede the macrocyclization. Triene **22** was to be constructed from building blocks **23**, **24**, and **5**.

Aldehyde **24** was obtained by Evans aldolization from ketoimide **6** and acrolein (Scheme 6) to give adduct **25** with good diastereoselectivity, which was converted into lactone **26** by reduction and removal of the auxiliary. For the conversion of **26** into aldehyde **24**, an analogous sequence was used as for the preparation of **20** from **19**.

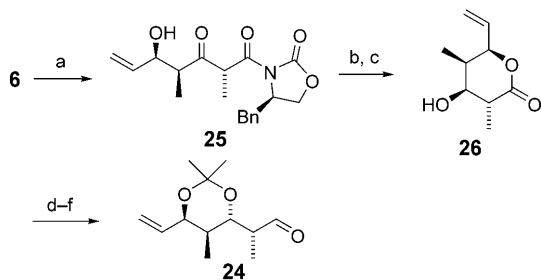


Scheme 4. Synthesis of benzofuran **2**. a) **6**, $\text{Sn}(\text{OTf})_2$, CH_2Cl_2 , Et_3N , -20°C , then -78°C , then **17**, 87% (d.r. 6:1); b) $\text{Me}_4\text{NBH}(\text{OAc})_3$, $\text{CH}_3\text{CN}/\text{AcOH}$ (2:1), -32°C to 0°C , 72% (d.r. 20:1); c) LiOH , H_2O_2 , THF/ H_2O (3:1), 96%; d) 3 M HCl, dioxane, 50°C ; e) $(\text{CH}_3)_2\text{C}(\text{OMe})_2$, CSA, RT, 85% (2 steps); f) LiOH , THF/ $\text{MeOH}/\text{H}_2\text{O}$ (2:1:1), 12 h, RT, 84%; g) EDCI, DMAP, DMAP-HCl, CHCl_3 , reflux, 20 h, 55%; h) $h\nu$ (254 nm), cyclohexane, 50 min, 75%; i) NaBH_4 , MeOH, RT, then 0.5 M HCl; j) TsOH, toluene, 60°C , 71% (2 steps). CSA = camphorsulfonic acid, Ts = 4-toluenesulfonyl.

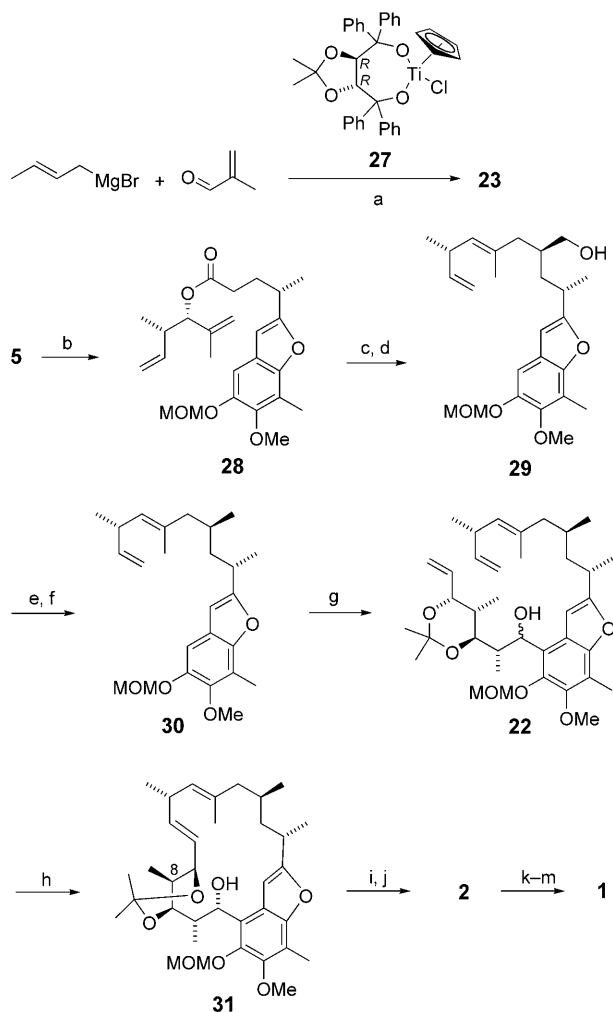


Scheme 5. Retrosynthesis of **2**: RCM route.

For the preparation of allylic alcohol **23**, a Duthaler–Hafner crotylation^[18] of methacrolein with titanate **27** was the method of choice (Scheme 7). Esterification of acid **5** with alcohol **23** paved the way for the Claisen–Ireland rearrangement, by which ester **28** was smoothly converted into the acid, and after reduction, into alcohol **29**. Deoxygenation furnished diene **30**. Subsequent *ortho*-directed metalation and addition of aldehyde **24** gave alcohol **22** as a mixture of diastereomers. RCM with second-generation Grubbs catalyst^[19] induced smooth ring closure to form the 10,11-*E*-olefin **31** exclusively. The extremely broad ^1H NMR signal of the 8- CH_3 group, which sharpens to a doublet on raising the temperature to



Scheme 6. Synthesis of aldehyde **24**. a) $\text{Sn}(\text{OTf})_2$, CH_2Cl_2 , Et_3N , -20°C , -78°C , then acrolein, 91% (d.r. 5:1); b) $\text{Me}_4\text{NBH}(\text{OAc})_3$, $\text{CH}_3\text{CN}/\text{AcOH}$ (2:1), -32°C to 0°C , 70% (d.r. 6:1); c) LiOH , H_2O_2 , $\text{THF}/\text{H}_2\text{O}$ (2:1), RT, 72%; d) $(\text{CH}_3)_2\text{C}(\text{OMe})_2$, CSA , RT, 91%; e) LiAlH_4 , Et_2O , 0°C , 96%; f) pyridine- SO_3 , Et_3N , $\text{CH}_2\text{Cl}_2/\text{DMSO}$, -5°C , 99%.



Scheme 7. RCM and synthesis of **1**. a) Reaction of but-2-enyl-MgBr with **27**, then methacrolein, Et_2O , -78°C , 52% (d.r. 50:1, 86% ee); b) DMAP, EDCl , CH_2Cl_2 , RT, then **23**, 81%; c) LHMDS (4 equiv), HMPA, THF, then **28** dissolved in TBSCl, -78°C to RT, then DMF, microwave irradiation (10 min, 180°C); d) LiAlH_4 , Et_2O , 0°C , 89% (d.r. 4:1, 2 steps); e) MsCl , CH_2Cl_2 , 0°C ; f) LiAlH_4 , Et_2O , 0°C , 89% (2 steps); g) $n\text{BuLi}$, TMEDA, THF, then **24**, -78°C to -30°C , 90% (d.r. 3.2:1); h) Grubbs II catalyst, 20 mol%, CH_2Cl_2 , reflux, 16 h, 62% (E only); i) $\text{N}_2(\text{COOK})_2$, AcOH , CH_2Cl_2 , 40 h, reflux, 60%; j) 3 M HCl, MeOH, RT, 96%; k) TESOTf , Et_3N , CH_2Cl_2 , 0°C , 82%; l) IBX, DMF, RT, 24 h; m) 0.1 M HF, MeCN, RT, 30% (2 steps). TMEDA=tetramethylethylenediamine, TES=triethylsilyl.

350 K (see Supporting Information), indicates that the conformational mobility in **31** is significantly restricted. Selective reduction of the double bond with diimide,^[20] followed by acid-catalyzed formation of the tetrahydropyran ring and removal of the OMOM group, led to intermediate **2**, which was oxidized to **1**.^[4a] The analytical data of the synthetic material were in full agreement with those of an authentic sample kindly provided by Professor Zeeck.

In conclusion, we have presented two novel approaches to the antibiotic kendomycin (RCM route: 23 linear steps, photo-Fries route: 29 linear steps). Apart from Claisen–Ireland rearrangements of unusual complexity, this work not only demonstrates the hitherto unrecognized capability of the photo-Fries ring contraction for the formation of macrocycles, but also reemphasizes the unparalleled potential of RCM for connecting monosubstituted olefin residues.

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Keywords: aldol reactions · antibiotics · benzofurans · Claisen–Ireland rearrangement · total synthesis

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3.2. Ring Closing Metathesis and Photo-Fries Reaction for the Construction of the Ansamycin Antibiotic Kendomycin. Development of a Protecting Group Free Oxidative Endgame

T. Magauer, H. J. Martin, J. Mulzer, *Chem. Eur. J.* **2009**, *Accepted*.

Ring Closing Metathesis and Photo-Fries Reaction for the Construction of the Ansamycin Antibiotic Kendomycin. Development of a Protecting Group Free Oxidative Endgame.

Thomas Magauer, Harry J. Martin, Johann Mulzer*^[a]

Abstract: Two convergent total syntheses of the *ansa*-polyketide (-)-Kendomycin **1** are described. The syntheses benefit from the use of readily available and cheap starting materials. Highly complex diastereoselective Claisen-Ireland rearrangements were used to introduce

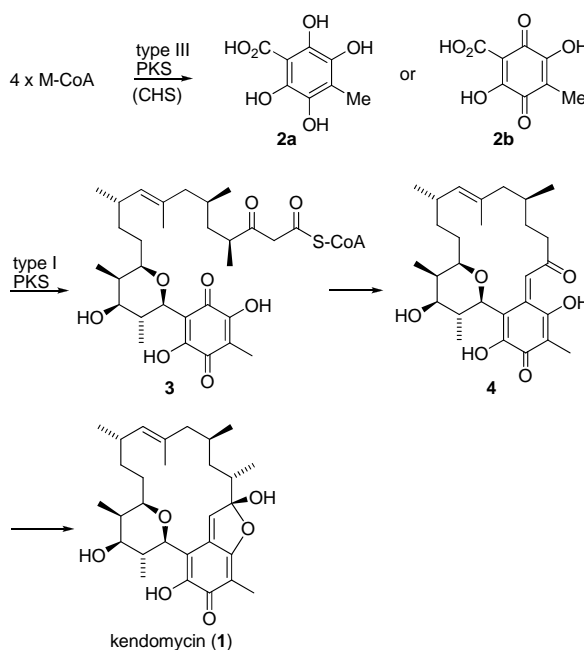
the (*E*)-double bond and the C16-Me group. The ring closure of the strained *ansa* macrocycle was achieved by RCM and a highly efficient combination of macrolactonization and photo-Fries reaction. A protecting group free endgame *via* an unstable *o*-quinone is presented. Additionally

some unsuccessful synthetic efforts towards the total synthesis of **1** are described.

Keywords: RCM • photo-Fries • Claisen-Ireland rearrangement • polyketides • *ansa*-compound

Introduction

Kendomycin [(-)-TAN 2162] (**1**) was first reported in 1996,^[1] and re-isolated in 2000 by Zeeck and Bode during their screening program for new metabolites from *Actinomycetes*.^[2] Biological testing revealed **1** to be a potent endothelin receptor antagonist and antiosteoporotic compound with remarkable antibacterial and cytostatic activity,^[2,3] most likely through proteasome inhibition.^[3a] Beside the diverse pharmacological qualities, which have attracted (bio)-chemists in the last years, Kendomycin discloses an unique molecular architecture with a fully carbogenic *ansa*-polyketide chain, nine stereogenic centers, a pentasubstituted tetrahydropyran ring and a remarkable *p*-quinone-methide chromophore. The biosynthesis (Scheme 1)^[2b,4] implies the formation of benzoic acid **2a** or the corresponding quinoid nucleus **2b** from malonate subunits under the mediation of chalcone synthase (CHS). This core unit is then loaded onto the type I polyketide synthase (PKS) to form keto acid **3** which undergoes cyclization to ketone **4** under decarboxylation. Ketalization leads to **1** eventually.



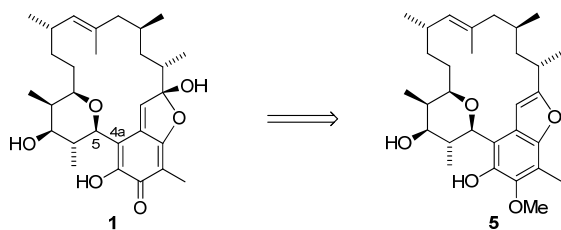
Scheme 1. Biosynthesis of kendomycin.

The challenging framework and the promising pharmacological profile of (**1**) has motivated us^[5] and sometime later, a number of other groups^[6-8] to carry out studies towards its synthesis. So far four total syntheses^[6] and one formal one^[7] have been reported, along with a number of fragment preparations.^[8] All these approaches loosely follow the biogenetic pathway by starting with an aromatic polyphenol subunit, attaching a polyketide chain and then aiming for cyclization. The main challenge has thus been the

formation of the strained macrocyclic *ansa*-ring and the late stage generation of the quinone and lactol units. So far, macrocyclizations have been performed *via* RCM^[6b], C-glycosidation,^[6a] Barbier-type organometal addition,^[6d] Prins reaction^[7] and Horner Wadsworth Emmons olefination.^[8e] In continuation of earlier reports^[5] we now want to disclose our recent efforts, which have culminated in two successful syntheses.^[6e]

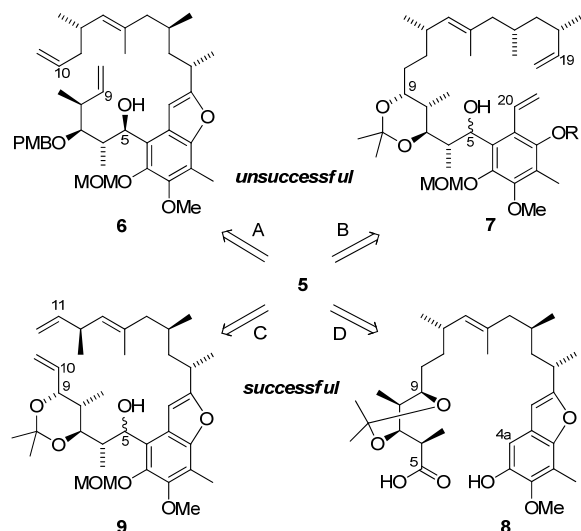
Results and Discussion

It is obvious that the formation of the quinone methide chromophore should be deferred to the end of the synthesis, *via* the oxidation of known^[6a] benzofuran **5** (Scheme 2). A further general consideration concerns the tetrahydropyran ring which preferably should be installed after the macrocyclization - mainly because of restricted rotation around the C4a-C5 bond^[5d] which might be disadvantageous for subsequent ring closures.



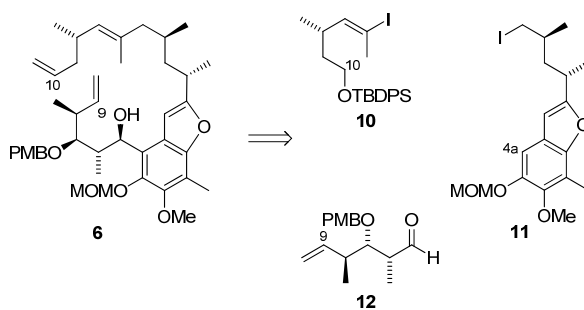
Scheme 2. Benzofuran precursor of kendomycin.

In this report we present four general approaches toward the synthesis of the common precursor **5** (Scheme 3). Three of them address the ring closing metathesis (RCM) at different sites as key steps. In the first approach (A), we intended to combine olefinic carbons C9 and C10 of compound **6** *via* RCM, followed by an addition of C5-OH to C9 for tetrahydropyran ring formation. In approaches B and C using compounds **7** and **9**, respectively as RCM precursors, the tetrahydropyran ring should be generated by diastereoselective S_N1-reaction of the C9-OH with an in situ generated benzylic cation at C5.^[9] The final approach (D) focuses on the macrolactonization of compound **8** followed by a photo-Fries reaction, and the tetrahydropyran should be formed by C5-carbonyl reduction and S_N1 cyclization. It should be noted at this point, that only approaches C and D have been successful, in contrast to route A where the RCM did not work and B, where the RCM precursor **7** could not be made at all.



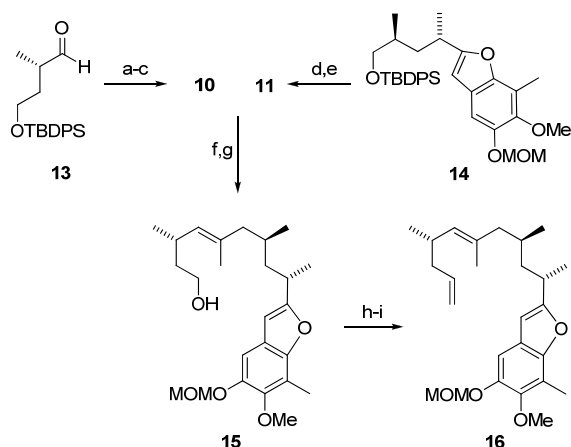
Scheme 3. Precursors for macrocyclizations..

RCM and trans-etherification (Route A): Retrosynthetically, the RCM precursor **6** was disconnected into vinyl iodide **10**, alkyl iodide **11** and aldehyde **12** (Scheme 4). The synthesis of the northern diene portion should be achieved by a Negishi cross coupling of iodides **10** and **11**, followed by chain elongation to the 10-olefin. *Ortho*-directed lithiation of C4a and addition to aldehyde **12** should set the stage for the envisaged RCM reaction.



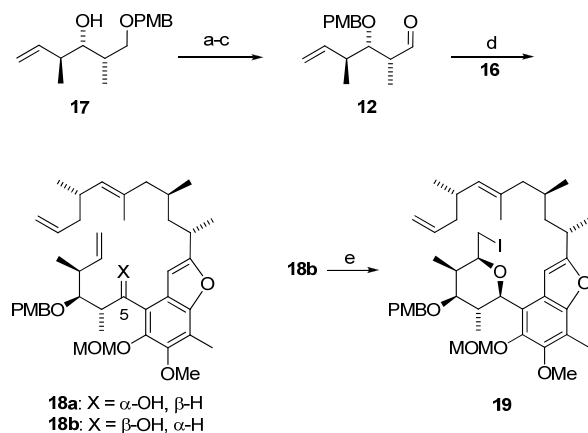
Scheme 4. Retrosynthetic disconnections for route A.

Vinyl iodide **10** was easily available from known aldehyde **13**.^[10] Colvin's one carbon chain elongation^[11] afforded the corresponding alkyne, which was alkylated with MeI and converted to **10** by hydrozirconation/iodination. Iodide **11** was prepared from known^[9] compound **14** via a two step standard procedure. Pd(0) assisted Negishi coupling^[12] of iodides **10** and **11**, followed by deprotection gave (*E*)-olefin **15** which was converted to 1,4-diene **16** *via* IBX-oxidation and Wittig methylenation (Scheme 5).



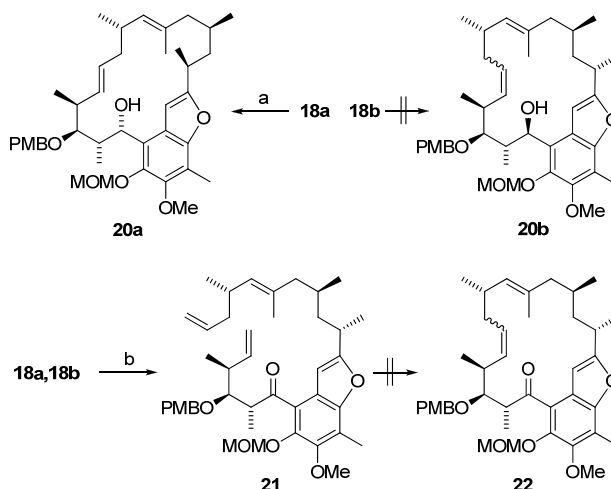
Scheme 5. Synthesis of compound **16**. a) TMSCHN₂, LDA, THF, -78 °C to r.t., 82%; b) BuLi, MeI, THF, -78 °C to r.t. 95%; c) Cp₂ZrClH, benzene, THF, I₂, 83%; d) TBAF, THF, 94%; e) I₂, PPh₃, CH₂Cl₂, 88%; f) **11**, *t*BuLi, ZnCl₂, Et₂O/THF, 5 mol% Pd(PPh₃)₄, -78 °C - r.t. add **10** in THF; g) TBAF, THF, 67% 2 steps; h) IBX, DMSO, r.t., 97%; i) MePPh₃Br, *t*BuOK, THF, 0 °C, 90%.

Aldehyde **12** was available from known alcohol **17**^[13] via 1,3 shift of the PMB protecting group and oxidation of the primary alcohol with IBX (Scheme 6). MOM-directed *ortho*-lithiation of **16** followed by nucleophilic addition to aldehyde **12** afforded benzylic alcohols **18a** and **18b** as a 1.5:1 diastereomeric mixture. The configuration at the benzylic carbon C5 was assigned by converting compound **18b** into cyclic iodoether **19**. 2D NMR experiments (NOESY) revealed that **19** and hence **18b** have the desired *R*-configuration at C5.



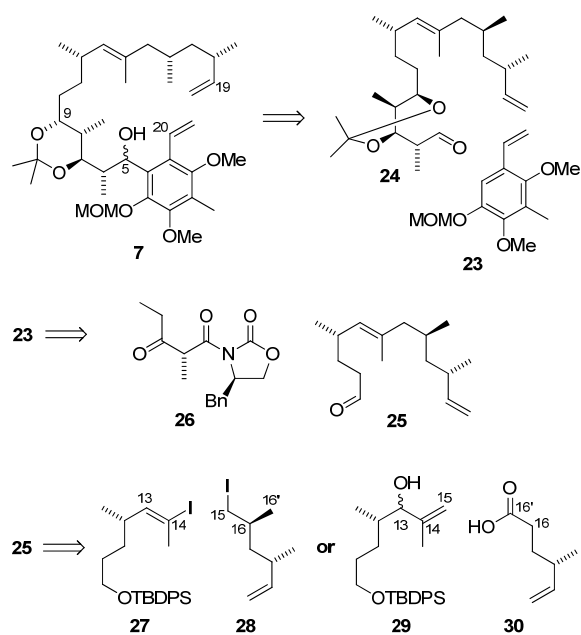
Scheme 6. a) DDQ, CH₂Cl₂, 3 Å MS, 0 °C, 74%; b) DIBAL, CH₂Cl₂, -78 °C to -10 °C, 93%; c) CH₂Cl₂, DMSO, (COCl)₂, NEt₃, -78 °C, 99%; d) **16**, *n*BuLi, TMEDA, THF, -40 °C then **12**, -78 °C to -25 °C, 75% (dr = 1.5:1); e) *t*butyl-4-methylpyridine, I₂, CH₂Cl₂, -78 °C to -10 °C, 50%.

Subjecting **18b** to Grubbs' II catalyst^[14] did not result in the desired cyclization to **20b**, but only decomposition of starting material was observed (Scheme 7). In contrast, **18a** underwent the cyclization and afforded macrocycle **20a** which was used for testing purposes. Unfortunately all attempts to form the tetrahydropyran by iodination, oxymercuration or selenocyclization failed. Additionally, as RCM of ketone **21** was not successful, we abandoned approach A at this point^[15] and turned to route B.



Scheme 7. a) Grubbs' II catalyst, 15 mol%, CH₂Cl₂, reflux, 16 h, 46%; b) IBX, DMSO, r.t., 96%.

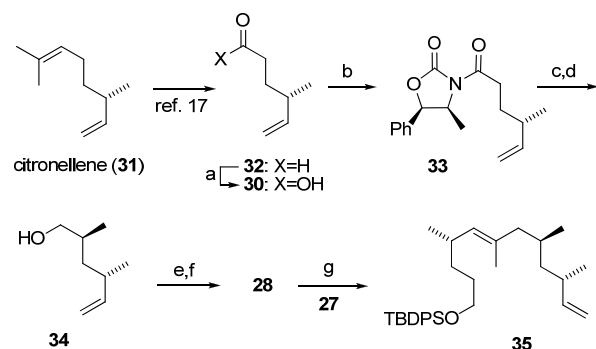
RCM reaction at C19/C20 (Route B): Installation of the 13,14-(*E*)-double bond via Negishi coupling and C4/5 connection via an *o*-lithiation aldehyde addition sequence have proven reliable and efficient. Additionally, we envisaged the Ireland-Claisen rearrangement^[16] as an appropriate tool for generating the 13,14-(*E*)-olefin along with the C-16 methyl group. Thus *seco* compound **7** should be available from styrene **23** and aldehyde **24**, which could be formed by an Evans aldol addition of aldehyde **25** and ketoimide **26** (Scheme 8). The installation of the C14/C15 double bond should then be achieved by either Negishi coupling of iodides **27**^[7] and **28** or by esterification of acid **30** with alcohol **29** followed by an Ireland-Claisen rearrangement.



Scheme 8. Retrosynthetic disconnections for route B.

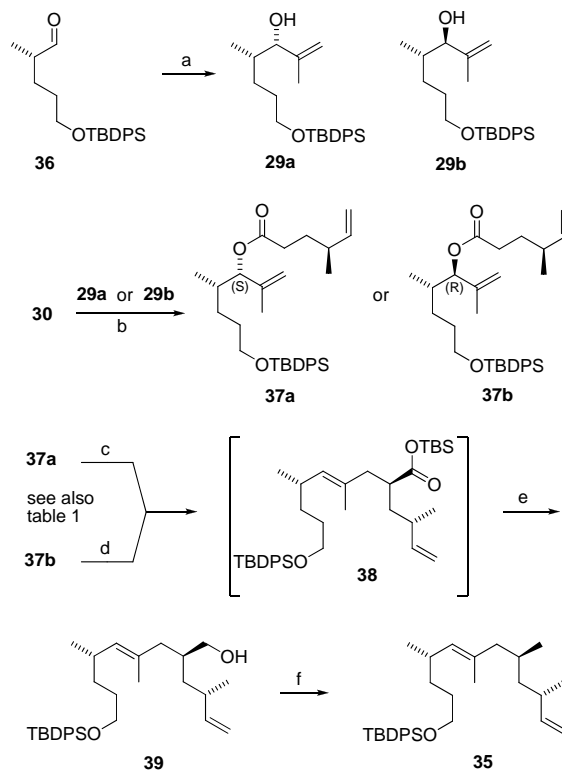
The synthesis started with known^[6a] aldehyde **32**, easily available from citronellene **31** in 2 steps.^[17] Pinnick oxidation^[18] to

the corresponding acid **30** followed by amidation afforded oxazolidinone **33** in good yield (Scheme 9). The second methyl group was introduced *via* diastereoselective alkylation with methyl iodide, and reductive removal of the auxiliary afforded primary alcohol **34**. Subsequent Finkelstein reaction delivered gram quantities of alkyl iodide **28** in excellent yield. Coupling of known vinyl iodide **27** with **28** smoothly afforded diene **35** as a key fragment.



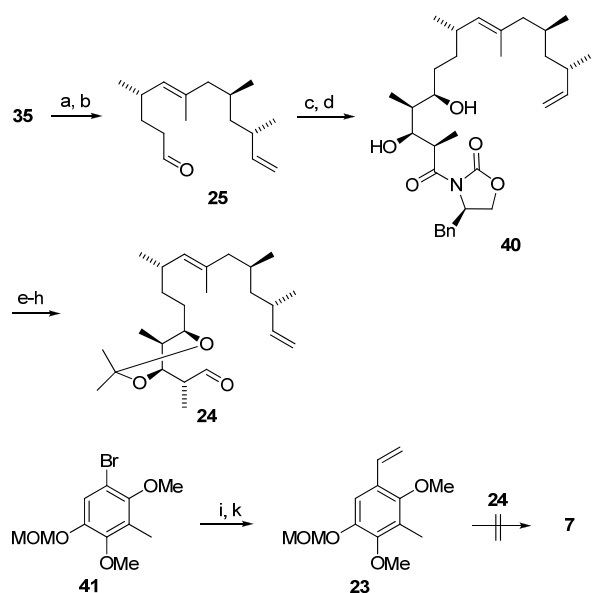
Scheme 9. a) NaClO_2 , NaH_2PO_4 , $t\text{BuOH}$, H_2O , 73% from **31**; b) DIC, DMAP, Evans' oxazolidinone, CH_2Cl_2 , 82%; c) LHMDs, MeI, THF, -78°C to r.t., 74% (dr = 10:1); d) LiAlH_4 , Et_2O , 0°C , 80%; e) MsCl , NEt_3 , CH_2Cl_2 , 0°C ; f) NaI , acetone, r.t., 86% from **34**; g) 2.2 equiv. $t\text{BuLi}$, ZnCl_2 , -78°C to 0°C , then **27**, $\text{Et}_2\text{O}/\text{THF}$, 5 mol% $\text{Pd}(\text{PPh}_3)_4$, 0°C , 95%.

In another approach (Scheme 10) for the synthesis of **35** we decided to use an Ireland-Claisen reaction. This should give access to the trisubstituted (*E*)-olefin and generate the stereocenter at C16 with the desired configuration. For this purpose, known aldehyde **36**^[6a] was treated with isopropenyl bromide in a Hiyama-Kishi reaction to give a 1.4:1 mixture of allylic alcohols **29**.^[19] The alcohols were separated and esterified with carboxylic acid **30** to afford compounds **37a** and **37b**, respectively. Treatment of **37a** with LDA in THF/HMPA afforded a (*Z*)-silyl ketene acetal, which was rearranged to the corresponding silyl ester **38** in good yield and acceptable diastereoselectivity (see also table 1).^[20] Reaction with potassium fluoride and subsequent reduction with LiAlH_4 furnished alcohol **39** which was reduced to give the C16-methyl group in **35**. Since the ester enolate geometry strongly depends on the solvents, treatment of **37b** with LDA in THF should give the corresponding (*Z*)-enolate, and thus, the rearrangement should likewise provide compound **39** after desilylation and reduction. Disappointingly, all attempts to rearrange the (*Z*)-enolate of **37b** proved to be low yielding.



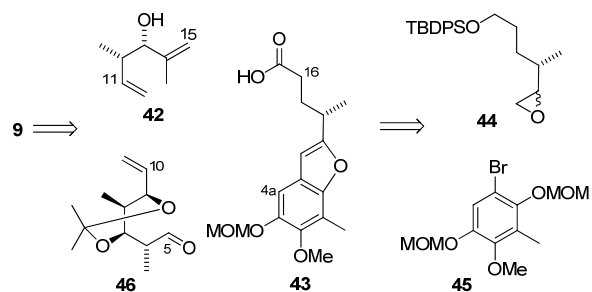
Scheme 10. a) CrCl_2 (4 equiv.), NiCl_2 (0.04 equiv.), DMF, 0°C to r.t., 86% (dr = 1.4:1); b) DIC, DMAP, CH_2Cl_2 , then **29a** or **29b**, 92%; c) LDA, THF-HMPA (23%) then TBSCl , -78°C to reflux; d) LDA, THF, then TBSCl -HMPA, -78°C to reflux; e) i. HMPA, KHCO_3 , KF, MeI, 0°C , ii. LiAlH_4 , Et_2O , 0°C , 63% from **37a** (dr = 5:1), 8% from **37b**; f) i. MsCl , NEt_3 , CH_2Cl_2 , 0°C , ii. LiAlH_4 , THF, 0°C to r.t., 90%.

Nevertheless, the rearrangement of **37a** had provided us with gram quantities of diene **35** and so we focused on the elongation sequence (Scheme 11). Deprotection and IBX oxidation furnished aldehyde **25** in 77% yield over two steps. Extended Evans aldol methodology^[21] followed by 1,3 reduction^[22] afforded stereotetrad (C6 to C9) **40** in good yield and high diastereoselectivity. Base induced hydrolysis to remove the auxiliary and treatment with camphorsulfonic acid in 2,2-dimethoxypropane and methanol afforded the methyl ester. Reduction with LiAlH_4 and oxidation of the resulting primary alcohol to aldehyde **24** paved the way for testing the final key steps. Thus, known aryl bromide **41**^[23] was formylated and then converted to styrene **23** *via* Wittig methylenation. To obtain the desired RCM precursor **7**, **23** and **24** had to be coupled as before, but unfortunately, addition of $n\text{BuLi}$ to **23** did not give the expected *ortho*-lithiated^[24] product but led to polymerization of the styrene unit. So, with a heavy heart after so much experimentation, we abandoned route B.



Scheme 11. a) TBAF, THF, r.t., 87%; b) IBX, DMSO, r.t., 95%; c) **26**, Sn(OTf)₂, CH₂Cl₂, NEt₃, -35 °C, then -78 °C, then add **25**, 76% (dr = 10:1); d) Me₄NBH(OAc)₃, CH₃CN:AcOH (1.9:1), -32 °C, 76% (dr = 20:1); e) LiOH, H₂O₂, THF/H₂O (3:1), 92%; f) 2,2-dimethoxypropane, CSA, 16 h, r.t., 90%; g) LiAlH₄, Et₂O, 0 °C, 99%; h) IBX, DMSO, r.t., 99%; i) *t*BuLi, DMF, 1N HCl, -78 °C to r.t., 83%; k) MePPh₃Br, *t*BuOK, THF, 0 °C, 98%.

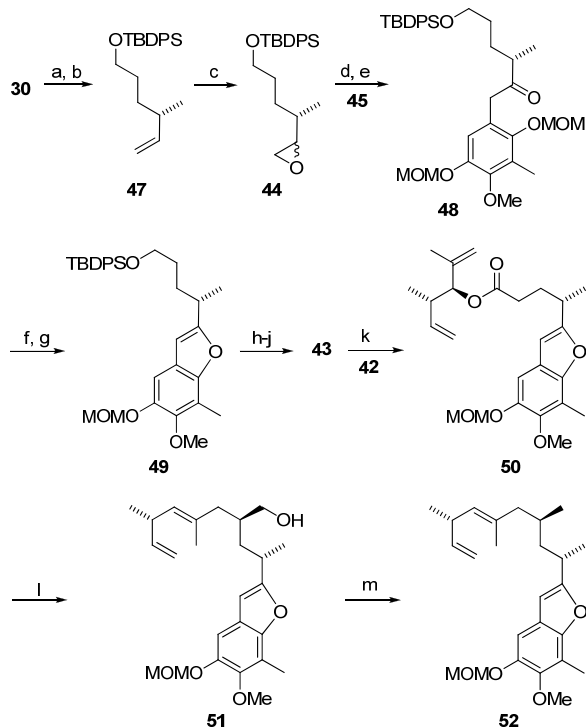
RCM reaction at C10/C11 (Route C): In our final RCM approach we aimed for the generation of a C10/C11 olefin which has subsequently to be reduced in presence of the 13,14-olefin. For the formation of the 13,14-tri-substituted double bond we wanted to reapply the Ireland-Claisen approach using the known allylic alcohol **42**^[25] and carboxylic acid **43** as simple precursors. Carboxylic acid **43** should be assembled from epoxide **44** and known aryl bromide **45**.^[8e] The missing tetrahydropyran side chain should be introduced in the usual way by *ortho*-lithiation of the C4a position and addition of aldehyde **46** (Scheme 12).



Scheme 12. Retrosynthetic disconnections for route C.

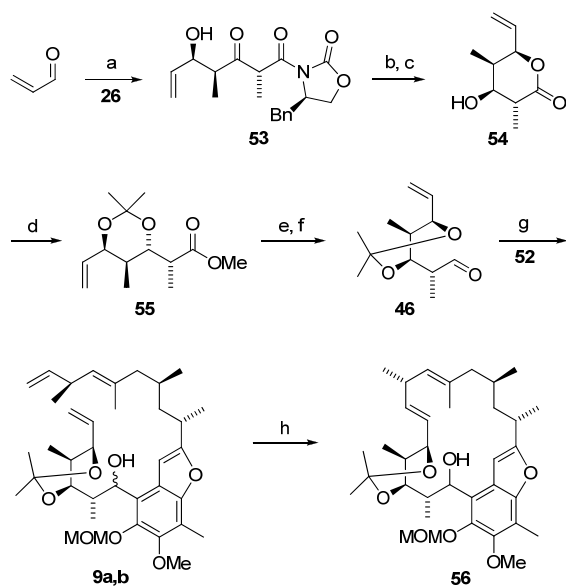
For the enantioselective preparation of allylic alcohol **42**, a Duthaler-Hafner crotylation^[26] of methacrolein proved to be the method of choice, as the asymmetric crotylation protocols by Roush^[27] or Brown^[28] lacked enantio- or diastereoselectivity in this case. The synthesis of acid **43** started from aldehyde **32** (available from citronellene **31** in two steps, see supporting information), which was reduced to the corresponding alcohol and converted into silyl ether **47** (Scheme 13) and then into epoxide **44**. Treatment of **44** with a cuprate reagent derived from bromide **45** gave the corresponding

alcohol as a mixture of diastereomers (ca. 1:1). Oxidation led to ketone **48**, which after treatment with triflic acid and reprotection with MOMCl furnished benzofuran **49** in good yield. Desilylation and two-step oxidation of the primary alcohol afforded carboxylic acid **43** which was esterified with alcohol **42** to provide the rearrangement precursor **50**. To our dismay, the Ireland-Claisen conditions we had used for Route B did not work out as expected. In our first tries, we had to struggle with moderate yields and very low diastereoselectivities. Fortunately, after a lot of optimization (see table 1), yield and diastereoselectivity were improved considerably. Subsequent reduction of the 16'-OH finished the synthesis of 1,3-diolefin **52**.



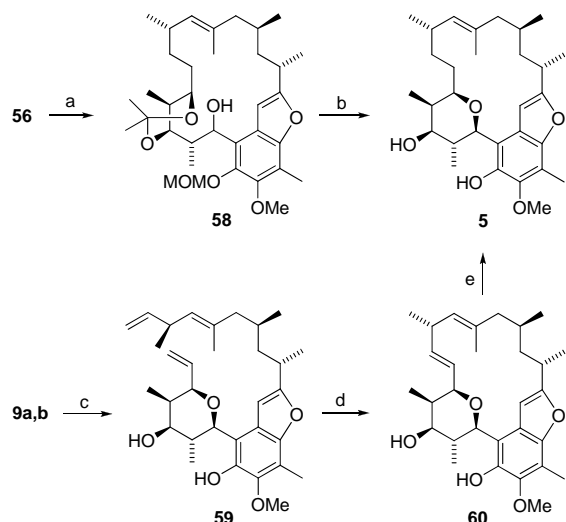
Scheme 13. a) LiAlH₄, Et₂O, 0 °C, 77% from citronellene **31**; b) TBDPSCl, imidazole, THF, r.t., 90%; c) mCPBA, CH₂Cl₂, 0 °C, 96%; d) **45**, Mg, THF, reflux 2h, -40 °C, CuI, then **44**, -40 °C to 0 °C, 4h, 87%; e) DMSO, (COCl)₂, NEt₃, CH₂Cl₂, -78 °C, 95%; f) TfOH, toluene, 80 °C; g) MOMCl, NaH, DMF, 95% from **47**; h) TBAF, THF, r.t., 89%; i) IBX, DMSO, r.t., 97%; j) NaClO₂, NaH₂PO₄, *t*BuOH, H₂O, 99 %; k) DMAP, EDCI-HCl, CH₂Cl₂, r.t., 86%; l) i. LHMDS (4 equiv.), HMPA, THF, then **50** dissolved in THF/TBSCl, -78 °C to r.t., then DMF, microwave, 15 min, 180 °C, ii. LiAlH₄, Et₂O, 0 °C, 89% (dr = 4:1); m) i. MsCl, CH₂Cl₂, 0 °C, ii. LiAlH₄, Et₂O, 0 °C, 89%.

Aldehyde **46** was obtained *via* Evans aldol addition of ketoimide **26** and acrolein (Scheme 14) to give adduct **53** in good yield and diastereoselectivity. Lactonization to **54** was performed *via* stereoselective carbonyl reduction and subsequent removal of the auxiliary. Treatment with camphorsulfonic acid in dimethoxypropane furnished ester **55** which was converted into aldehyde **46** by a reduction-oxidation sequence. *Ortho*-directed lithiation of **52** and addition of aldehyde **46** gave tri-olefin **9** as a 3.5:1 mixture of diastereomers **9a/9b**, which was separated by chromatography. RCM of the major diastereomer (**9a**) with Grubbs' second generation catalyst induced smooth ring closure to 10,11-(*E*)-olefin **56** exclusively.



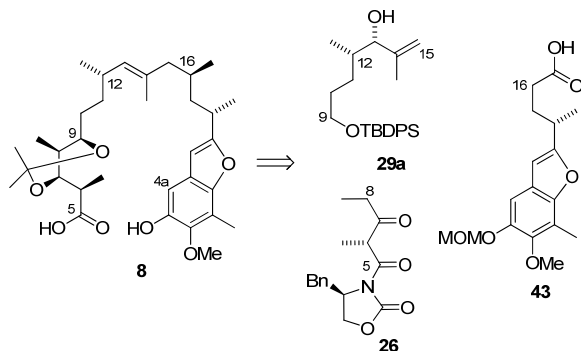
Scheme 14. a) $\text{Sn}(\text{OTf})_2$, CH_2Cl_2 , Et_3N , $-20\text{ }^\circ\text{C}$, $-78\text{ }^\circ\text{C}$, then acrolein, 91% (dr = 5:1); b) $\text{Me}_4\text{NBH}(\text{OAc})_3$, $\text{CH}_3\text{CN}/\text{AcOH}$ (2:1), $-32\text{ }^\circ\text{C}$ to $0\text{ }^\circ\text{C}$, 70% (dr = 6:1); c) LiOH , H_2O_2 , $\text{THF}/\text{H}_2\text{O}$ (2:1), r.t., 72%; d) $(\text{CH}_3)_2\text{C}(\text{OMe})_2$, CSA , r.t., 91%; e) LiAlH_4 , Et_2O , $0\text{ }^\circ\text{C}$, 96%; f) $\text{Pyridine}\cdot\text{SO}_3$, NEt_3 , $\text{CH}_2\text{Cl}_2/\text{DMSO}$, $-5\text{ }^\circ\text{C}$, 99%; g) $n\text{BuLi}$, TMEDA , THF , then **52**, $-78\text{ }^\circ\text{C}$ to $-30\text{ }^\circ\text{C}$, 90% (dr = 3.5:1); h) Grubbs' II catalyst, 20 mol%, CH_2Cl_2 , reflux, 16 h, 62% (*E*) only.

Site selective reduction of the 10,11-olefin with diimide,^[29] followed by acid induced formation of the tetrahydropyran ring and concomitant removal of the MOM-group led to key intermediate **5**. Since the minor diastereomer **9b** did not undergo the RCM reaction and the $\text{S}_{\text{N}}1$ tetrahydropyran formation is independent of the configuration at C5 we concluded that it might be advantageous to change the order of the cyclization reactions (Scheme 15). Treatment of the **9a,b**-mixture with HCl resulted in clean formation of tetrahydropyran **59**, which, not surprisingly showed the typical atropisomerism (1.5:1) of those compounds. Pleasingly, the subsequent RCM afforded the desired macrocycle in excellent yield and almost exclusively as the (*E*)-isomer **60** (15:1).^[30] The success of this RCM came totally unexpected, as we had anticipated major problems from the tetrahydropyran ring. Diolefin **59** was reduced with high site selectivity to compound **5** with diimide.



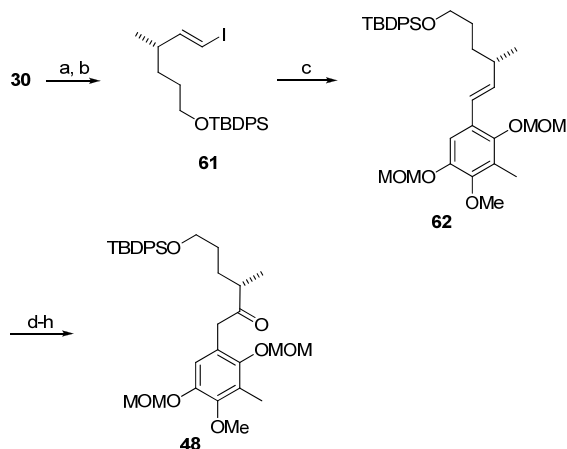
Scheme 15. Synthesis of benzofuran **5** via RCM. a) $\text{N}_2(\text{COOK})_2$, AcOH , CH_2Cl_2 , 40h, reflux, 58%; b) 3N HCl , MeOH , r.t., 96%; c) 3N HCl , MeOH , r.t., 71%; d) Grubbs' II catalyst, 20 mol%, CH_2Cl_2 , reflux, 16 h, 83% (*E:Z* = 15:1); e) $\text{N}_2(\text{COOK})_2$, AcOH , CH_2Cl_2 , 5h, reflux, 71%.

Macrolactonization and photo-Fries reaction to close the C4a/C5 bond (route D): This approach (Scheme 16) was centered around *seco* acid **8** as a key intermediate. The carbon skeleton should be assembled from the established building blocks **43** and **29a** which would give the (*E*)-13,14-olefinic unit via Claisen–Ireland rearrangement. Evans aldol addition of a C9-aldehyde with ketoimide **26** should be used for the C8–C5-chain elongation.



Scheme 16. Retrosynthesis for route D.

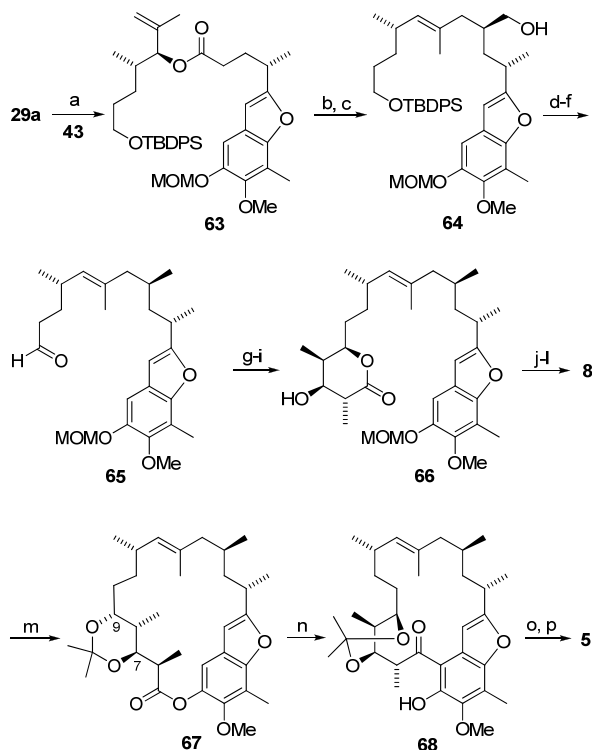
For ketone **48**, which serves as the precursor of acid **43**, we developed a new route (Scheme 17). Starting with aldehyde **30**, Colvin's chain elongation furnished the corresponding alkyne which was converted into vinyl iodide **61**. Negishi coupling with aryl bromide **45** furnished styrene **62**, which, after epoxidation was subjected to a $\text{Pd}(0)$ -mediated rearrangement^[31] to ketone **48**.



Scheme 17. Synthesis of compound **48**. a) TMSCHN_2 , $n\text{BuLi}$, THF , $-78\text{ }^\circ\text{C}$ to r.t., 83%; b) Cp_2ZrHCl , benzene, $50\text{ }^\circ\text{C}$; I_2 , $0\text{ }^\circ\text{C}$, 76%; c) **45**, $t\text{BuLi}$, ZnCl_2 , $\text{Et}_2\text{O}/\text{THF}$, $-78\text{ }^\circ\text{C}$ to $0\text{ }^\circ\text{C}$, $\text{Pd}(\text{PPh}_3)_4$, then add **61**, 67%; d) DMDO , acetone, r.t., 99%; e) $\text{Pd}(\text{OAc})_2$, PPh_3 , $t\text{BuOH}$, reflux, 81%.

Allylic alcohol **29a** was connected with acid **43** to furnish ester **63** as the substrate of an Ireland–Claisen rearrangement (Scheme 18). Treatment with excess LHMDS and reductive work-up led to

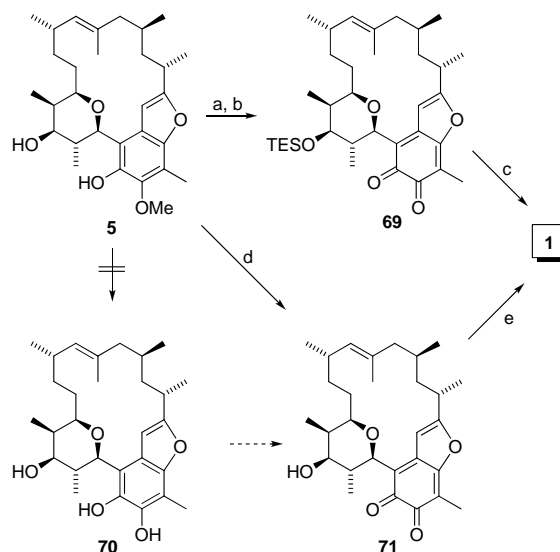
primary alcohol **64** as an easily separable 4:1 diastereomeric mixture. Subsequent reduction of the carboxyl to the methyl group followed by desilylation and oxidation gave aldehyde **65** which was subjected to an aldol addition with ketoimide **26**. Diastereoselective 1,3-reduction followed by acid catalyzed lactonization furnished lactone **66** which was converted into *seco* acid **8** via the 7,9-acetonide protected methyl ester. Macrolactonization of **8** under modified Boden-Keck conditions^[32] worked nicely to give 55% of monomer **67**, which underwent clean photo-Fries rearrangement to ketone **68**. Reduction of the ketone to the alcohol (diastereomeric mixture) followed by removal of the acetonide and S_N1-cyclization furnished key intermediate **5**.



Scheme 18. Synthesis of benzofuran **5** via photo-Fries rearrangement. a) EDCI, DMAP, **43**, CH₂Cl₂, 85%; b) LHMDS, HMPA, TBSCl, -78 °C to reflux; c) LiAlH₄, Et₂O, 0 °C, 84% from **63** (dr = 4:1); d) i. MsCl, Et₃N, CH₂Cl₂, 0 °C, ii. LiAlH₄, Et₂O, 0 °C, 94% (2 steps); e) TBAF, THF, r.t., 93%; f) IBX, DMSO, r.t., 93%; g) **26**, Sn(OTf)₂, CH₂Cl₂, Et₃N, -20 °C, then -78 °C, then **65**, 87% (dr = 6:1); h) Me₄NBH(OAc)₃, CH₃CN/AcOH (2:1), -32 °C to 0 °C, 72% (dr = 20:1); i) LiOH, H₂O₂, THF/H₂O (3:1), 96%; j) 3N HCl, dioxane, 50 °C; k) (CH₃)₂C(OMe)₂, CSA, r.t., 85% 2 steps; l) LiOH, THF/MeOH/H₂O (2:1:1), 12 h, r.t., 84%; m) EDCI, DMAP, DMAP-HCl, CHCl₃, reflux, 20 h, 55%; n) *hν*, 254 nm, cyclohexane, 50 min, 75%; o) NaBH₄, MeOH, r.t., then 0.5N HCl; p) TsOH, toluene, 60 °C, 71% from **68**.

Completion of the total synthesis: With two successful approaches for benzofuran intermediate **5** in hands, we focused on the crucial oxidative endgame (Scheme 19). Firstly, we reproduced Lee's endgame^[6a] by starting with protection of the C7-OH to give the corresponding TES ether which was then oxidized with IBX to provide the unstable yet isolable *o*-quinone **69**. On treatment of **69** with aqueous HF, the silyl group was removed and 1,6 conjugate addition of water occurred to furnish the target molecule (**1**). In an alternative approach we tried to avoid the OTES protecting group. For this purpose we envisaged a biomimetic pathway, by first converting **5** into catechol **70**, followed by oxidation to quinone **71** and spontaneous addition of water. Unfortunately we could not

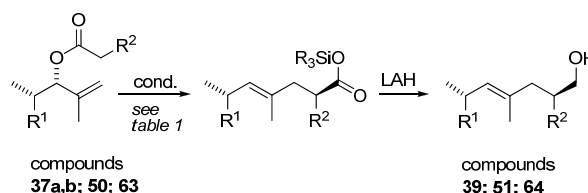
remove the phenolic methyl ether even under a variety of conditions. Still convinced that it should be possible to work out a protecting group free endgame we tried the direct oxidation of **5** with different oxidants, for instance Fremy's salt ((KSO₃)₂NO), CAN, Ag₂O, PIDA, NaIO₄ and IBX. These experiments all failed, but finally we discovered that DDQ in CH₂Cl₂/H₂O cleanly oxidized **5** to *o*-quinone **71**, which was immediately hydrolyzed to kendomycin (**1**) on treatment with diluted hydrochloric acid.



Scheme 19. Oxidation of **5**. a) TESOTf, Et₃N, CH₂Cl₂, 0 °C, 82%; b) IBX, DMF, r.t., 24 h c) 0.1 M HF, MeCN, r.t., 30 % (2 steps); d) DDQ, CH₂Cl₂/H₂O = 10:1, r.t., 52% e) aq. HCl (1%), MeCN, 50%.

Conclusion

In conclusion we have presented four synthetic approaches, two of which resulted in convergent total syntheses of kendomycin (**1**). For the stereoselective installation of the (*E*)-13,14-olefin we investigated the experimental conditions for three Ireland-Claisen reactions of unusual complexity, summarized in Scheme 20 and Table 1, respectively.



Scheme 20. Ireland-Claisen rearrangements.

Table 1. Reaction conditions for Ireland-Claisen rearrangements.

| compound | Base (equiv.) /SiR ₃ X (equiv.) ^[a] | reaction conditions ^[b] | product | yield ^[c] | dr ^[d] |
|------------|---|--|-----------|----------------------|-------------------|
| 37a | LDA (1.2) / TBSCl (1.1) | THF/HMPA, 2h | 39 | 63 % | 5:1 |
| 37b | LDA (1.2) / TBSCl (1.1) | THF, 2h | 39 | traces | n.d. |
| 50 | LDA (1.25) / TBSCl (1.1) | THF/HMPA, 3h | 51 | 20% | n.d. |
| 50 | LDA (1.25) / TBSOTf (1.1) | THF/HMPA, 15h | 51 | 35% | 5:1 |
| 50 | LDA (3.0) / TMSCl (3.0) | THF/HMPA, 14h r.t | 51 | 17% | n.d. |
| 50 | LDA (3.0) / TBSCl (10.0) | THF/HMPA/toluene ^[e] , 1h | 51 | 59% | 1.1:1 |
| 50 | LDA (3.2) / TBSCl (5.5) | THF, 2h | 51 | 19% | 4:1 |
| 50 | LDA (5.0) / TBSCl (7.0) | THF/HMPA, 3h | 51 | 46% | 10:1 |
| 50 | LHMDS (1.25) / TBSCl (1.2) | THF/HMPA, 3h ^[f] | 51 | traces | n.d. |
| 50 | LHMDS (4.0) / TBSCl (6.0) | THF/HMPA, 3h | 51 | 64 | 1:1 |
| 50 | LHMDS (4.0) / TBSCl (6.0) | THF/HMPA, 3h^[f] | 51 | 84% | 4:1 |
| 63 | LDA (1.25) / TBSCl (6.0) | THF/DMPU, 2h ^[f] | 64 | n.d. | n.d. |
| 63 | LDA (5.0) / TBSCl (5.5) | THF/DMPU, 2h | 64 | n.d. | n.d. |
| 63 | LHMDS (4.0) / TBSCl (6.0) | THF/HMPA, 3h ^[f] | 64 | 47% | 2:1 |
| 63 | LHMDS (5.0) / TBSCl (6.0) | THF/HMPA, 4h ^[f] | 64 | 63% | 2:1 |
| 63 | LHMDS (6.0) / TBSCl (7.0) | THF/HMPA, 2h ^[f] | 64 | 58% | 2:1 |
| 63 | LHMDS (4.0) / TBSCl (6.0) | THF/HMPA; DMF^{[f],[g]} | 64 | 89% | 4:1 |

[a] Enolization and silyl ketene acetal formation were performed at -78 °C. [b] The reactions were refluxed, unless otherwise stated. All rearrangement products were treated with LiAlH₄ after workup. [c] Yields were determined after reductive workup [d] The diastereomeric ratio (dr) was determined by ¹H-NMR. [e] Internal quench conditions. [f] The silyl ketene acetal was isolated before rearrangement. [g] The starting material was added as a solution in THF/TBSCl. The rearrangement was performed under microwave irradiation at 180 °C.

For the formation of the tetrahydropyran ring a remarkably efficient S_N1 cyclization was used either before or after the macrocyclization. Regarding the crucial issue of ring closure, our work not only demonstrates the so far unrecognized capability of the photo-Fries ring contraction for the formation of macrocycles, but also reemphasizes the unparalleled potential of RCM for connecting monosubstituted olefin residues. Additionally a protecting group free endgame for converting **5** into **1** was developed, which saves another synthetic step.

Experimental Section

All solvents were distilled prior to use, except THF, which was purchased from Acros Organics (99.85%, H₂O < 50 ppm) and used without further purification. Et₂O, toluene and benzene were distilled from sodium. CH₂Cl₂ and CHCl₃ were passed through an Al₂O₃-MgSO₄ column or distilled over P₂O₅. Acetone was distilled over P₂O₅. DMF, DMSO, NEt₃, *i*Pr₃NH, *i*Pr₃NEt, TMEDA, HMPA and 2,6-lutidine were distilled from CaH₂. TBSCl was dissolved in hexane or THF (3 M), treated with Et₃N (3%) and transferred *via* a syringe filter to the reaction mixture. CpZrHCl was prepared according to the Negishi procedure^[33]. Solvent degassing was achieved by repeated (at least 4 cycles) freeze-pump-thaw (FPT) cycles. All non-aqueous reactions were performed under an atmosphere of argon using oven-dried or flame-dried glassware and standard syringe/septa techniques. ¹H- and ¹³C-NMR spectra were measured in CDCl₃ on a Bruker Avance DRX-400 or DRX-600 at 400.13 MHz (100.61 MHz) or 600.13 MHz (150.90 MHz), respectively. Chemical shifts are given in ppm and were referenced to residual CHCl₃ (¹H, δ = 7.26 ppm, ¹³C, δ = 77.00 ppm) or toluene (¹H, δ = 7.09, 7.00, 6.98 ppm, ¹³C, δ = 137.9, 129.2, 128.3, 125.5, 20.4 ppm). Data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad), coupling constant in Hz, integration. Assignments of proton resonances were confirmed by correlated spectroscopy. IR spectra were recorded as thin films on a silicon plate on a Perkin-Elmer 1600 FT-IR spectrometer. Mass spectra were measured on a Micro mass, trio 200 Fisons Instruments. High-resolution mass spectra (HRMS) were performed with a Finnigan MAT 8230 with a resolution of 10000. Optical rotations were measured on a Perkin-Elmer 351 polarimeter at 20 °C (reported as follows: concentration (c in g/100mL), solvent). The reaction progress was monitored

on precoated TLC plates (Merck Kieselgel 60 F254). Spots were visualized under UV light (254nm) and/or were stained with ceric ammonium molybdate (CAM), *p*-anisaldehyde or potassium permanganate stain. Column chromatography was performed with Merck silica gel 60 (230-400 mesh). Analytical HPLC was performed on a Jasco System (PU-980 pump, UV 975 and RI 930) using a Nucleosil 50 column (5 μm, Ø 4 mm x 241 mm) at ambient temperature. Preparative HPLC was performed on a Dynamix Model SD-1 equipped with a Model UV-1 absorbance detector using a Supersphere (60 Å pore size, 4 μm particle size, Ø 25 mm x 250 mm) at ambient temperature. Yields refer to chromatographically purified compounds, unless otherwise stated.

S)-tert-Butyl(4-methylhex-5-enyloxy)diphenylsilane (47). β(-)-Citronellene (20.3 g, 147 mmol, 1.0 equiv.) and sodium acetate (12.6 g, 154 mmol, 1.05 equiv.) were dissolved in CH₂Cl₂ (490 mL) and cooled to -20 °C. *m*-CPBA (75 %, 35.4 g, 154 mmol, 1.05 equiv.) was added in small portions and stirring was continued for 1.5 h, allowing the suspension to warm to 0 °C. The reaction was quenched by careful addition of saturated aqueous NaHCO₃ (200 mL) and extracted with CH₂Cl₂ (3 x 70 mL). The combined organic fractions were washed with 1 N NaOH (100 mL), dried over MgSO₄ and concentrated *in vacuo*. The crude product was dissolved in Et₂O (245 mL), cooled to 0 °C and H₃IO₆ (50 g, 220 mmol, 1.5 equiv.) in THF (220 mL) was added within 45 min. Stirring was continued until TLC analysis showed complete consumption of the starting material. The mixture was diluted with Et₂O (500 mL), H₂O (300 mL) was added and the phases were separated. The organic layer was washed twice with brine, dried over MgSO₄ and filtered. This solution was recooled to 0 °C and LiAlH₄ (4 M in Et₂O, 44 mL, 176 mmol, 1.2 equiv.) was added *via* a dropping funnel over 2 h. The solution was slowly quenched with ethylacetate (10 mL), 1 N KHSO₄ (200 mL) was carefully added and the aqueous layer was extracted with Et₂O (3 x 100 mL). The organic fraction was dried over MgSO₄, filtered and evaporated to dryness. Purification of the residue by flash chromatography (pentane : diethylether = 5 : 1) afforded the alcohol as a colorless oil (13.0 g, 77% over 3 steps). ¹H-NMR (400MHz, CDCl₃): δ = 5.69 (ddd, *J* = 17.4, 10.2, 7.4 Hz, 1H), 4.99-4.90 (m, 1H), 3.63 (t, *J* = 6.3 Hz, 2 H), 2.20-2.07 (m, 1H), 1.62-1.51 (m, 2H), 1.40-1.32 (m, 2H), 1.31-1.25 (br, OH), 1.00 (d, *J* = 6.6 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ = 144.4, 112.8, 63.1, 37.6, 32.6, 30.5, 20.2; IR (film): 3331, 3077, 2935, 1640, 1455, 1419, 1374, 1058 cm⁻¹; HRMS (ESI) (*m/z*): [M-H₂O]⁺ calcd for C₇H₁₂, 96.0939, found, 96.0919; [α]_D²⁰ +18.8 (c 1.25, CH₂Cl₂).

Above prepared alcohol (3.30 g, 28.8 mmol, 1.0 equiv.) in DMF (29 mL) was cooled to 0 °C and imidazole (3.92 g, 57.6 mmol, 2 equiv.) was added. After 5 min TBDPSCl (7.4

mL, 28.8 mmol, 1.0 equiv.) was transferred to the solution *via* cannula and stirring was continued for 1 h at ambient temperature. The reaction mixture was diluted with Et₂O (200 mL), quenched with NH₄Cl (100 mL) and the phases were separated. The aqueous layer was extracted with Et₂O (3 x 50 mL), the organic layer was washed with brine, dried over MgSO₄ and concentrated *in vacuo*. Purification by flash chromatography (hexane : ethylacetate = 10 : 1) afforded compound **47** as a colorless oil (9.10 g, 90%). ¹H-NMR (400MHz, CDCl₃): δ = 7.71-7.64 (m, 4H), 7.45-7.33 (m, 6H), 5.68 (ddd, *J* = 17.3, 10.1, 7.3 Hz, 1H), 4.96-4.88 (m, 1H), 3.65 (t, *J* = 6.6 Hz, 2H), 2.16-2.04 (m, 1H), 1.62-1.52 (m, 2H), 1.39-1.32 (m, 2H), 1.05 (s, 9H), 0.98 (d, *J* = 6.6 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ = 144.7, 135.6, 134.2, 129.5, 127.6, 112.5, 64.1, 37.4, 32.7, 30.2, 26.9, 20.2, 19.2; IR (film): 2932, 1639, 1589, 1473, 1427, 1389, 1361, 1112 cm⁻¹; HRMS (ESI) (*m/z*): [M-*t*Bu]⁺ calcd for C₁₉H₂₅OSi, 295.1518, found, 295.1522; [α]_D²⁰ +6.7 (c 1.10, CH₂Cl₂).

(S)-tert-Butyl(4-(oxiran-2-yl)pentyl)oxy)diphenylsilane (44). Alkene **47** (5.91 g, 16.74 mmol, 1.0 equiv.) was dissolved in CH₂Cl₂ (57 mL) and cooled to 0 °C. *m*-CPBA (75%, 9.3 g, 40.17 mmol, 2.4 equiv.) was added in small portions and stirring was continued for 3 h. The reaction was filtered over Celite, quenched by the careful addition of saturated aqueous NaHCO₃ (70 mL) and extracted with CH₂Cl₂ (3 x 70 mL). The combined organic fractions were dried over MgSO₄ and concentrated *in vacuo*. Purification of the residue by flash chromatography (hexane : ethylacetate from 10 : 1 to 5 : 1) afforded epoxide **44** as a colorless oil (mixture of diastereomers) (5.91 g, 96%). ¹H-NMR (The asterisk denotes the minor diastereomer, 400MHz, CDCl₃): δ = 7.69-7.65 (m, 4H), 7.45-7.35 (m, 6H), 3.70-3.63 (m, 2H), 2.75-2.64 (m, 2H), 2.51-2.47 (m, 1H), 2.47-2.44* (m, 1H), 1.72-1.55 (m, 2H), 1.52-1.40 (m, 1H), 1.40-1.19 (m, 2H), 1.06 (s, 9H), 1.02 (d, *J* = 6.6 Hz, 3H), 0.92* (d, *J* = 6.8, 3H); ¹³C-NMR (The asterisk denotes the minor diastereomer, 100MHz, CDCl₃): δ = 135.6, 134.1*, 134.0, 129.6, 129.5*, 127.6, 64.1*, 64.0, 57.0, 56.9*, 46.9, 45.6*, 36.0, 35.8*, 30.7*, 30.1, 29.9*, 29.7, 26.9, 19.2, 17.1, 15.6*; IR (film): 3071, 3048, 2932, 1590, 1472, 1428, 1390, 1361, 1268, 1189, 1112 cm⁻¹; HRMS (ESI) (*m/z*): [M-*t*Bu]⁺ calcd for C₂₁H₂₅O₂Si, 311.1467, found, 311.1464.

(S)-6-(tert-Butyldiphenylsiloxy)-1-(4-methoxy-2,5-bis(methoxymethoxy)-3-methylphenyl)-3-methylhexan-2-one (48). Bromide **45** (11.3 g, 35.18 mmol, 3 equiv.) was dissolved in THF (50 mL). Mg (855 mg, 35.18 mmol, 3 equiv.), a crump of iodine and 2 drops of dibromoethane were added and the mixture was heated to reflux until the Mg has been completely consumed (1.5 h). The reaction was allowed to cool to room temperature and transferred to a solution of CuI (223 mg, 1.17 mmol, 0.1 equiv.) in THF (12 mL) at -50 °C. The resulting grey suspension was stirred for 30 min at -30 °C and then cooled to -45 °C. Epoxide **44** (4.3 g, 11.7 mmol, 1 equiv.) in THF (23 mL) was added dropwise and the temperature was raised to 0 °C within 4 h. The reaction was quenched by the addition of saturated aqueous NH₄Cl (100 mL) and the phases were separated. The aqueous layer was extracted with Et₂O (4 x 50 mL), the combined organic phases were dried over MgSO₄, filtered and concentrated. Purification by flash chromatography (hexane : ethylacetate from 10 : 1 to 3:1) gave 6.2 g (87%) of a diastereomeric mixture of the alcohols.

Oxalylchloride (1.72 mL, 20.30 mmol, 2 equiv.) was dissolved in CH₂Cl₂ (50 mL), cooled to -78 °C and DMSO (2.88 mL, 40.60 mmol, 4 equiv.) was added dropwise. After 40 min, above alcohol (6.2 g, 10.15 mmol, 1 equiv.) in CH₂Cl₂ (20 mL) was added *via* syringe and stirring was continued for additional 45 min. DIPEA (10.6 mL, 60.90 mmol, 6 equiv.) was added and the solution was warmed to 0 °C. The reaction was hydrolyzed with saturated aqueous NH₄Cl (100 mL), extracted with CH₂Cl₂ (3 x 50 mL), washed with brine, dried over MgSO₄ and filtered. Purification by column chromatography (hexane : ethylacetate from 5 : 1 to 3 : 1) afforded 5.8 g (95%) of ketone **48**. ¹H-NMR (400MHz, CDCl₃): δ = 7.68-7.63 (m, 4H), 7.45-7.34 (m, 6H), 6.74 (s, 1H), 5.14 (s, 2H), 4.83 (s, 2H), 3.80 (s, 3H), 3.73 (d, *J* = 3.8 Hz, 2H), 3.63 (t, *J* = 6.2 Hz, 2H), 3.51 (s, 3H), 3.48 (s, 3H), 2.68-2.58 (m, 1H), 2.20 (s, 3H), 1.80-1.70 (m, 1H), 1.56-1.47 (m, 2H), 1.48-1.39 (m, 1H), 1.09 (d, *J* = 6.8 Hz, 3H), 1.04 (s, 9H); ¹³C-NMR (100MHz, CDCl₃): δ = 211.8, 150.0, 148.0, 146.9, 135.5, 134.0, 129.5, 127.6, 125.8, 123.5, 116.2, 99.7, 95.5, 63.7, 60.4, 57.4, 56.2, 45.0, 43.3, 30.1, 29.1, 26.8, 19.2, 16.5, 10.4; IR (film): 2933, 1710, 1559, 1481, 1237, 1155, 1112, 967 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₃₅H₄₈O₇Si, 608.3169, found, 608.3186; [α]_D²⁰ +9.1 (c 0.95, CH₂Cl₂).

(S)-tert-Butyl(4-(6-methoxy-5-(methoxymethoxy)-7-methylbenzofuran-2-yl)pentyl)oxy)diphenylsilane (49). Ketone **48** (15.8 g, 25.95 mmol, 1.0 equiv.) and 15.8 g of molecular sieves (4Å) in 500 mL of toluene : EtOH = 4 : 1 were heated to 80 °C. After the addition of TfOH (689 μL, 7.79 mmol, 0.3 equiv.) stirring was continued at 80 °C for 5 min and then the mixture was rapidly cooled to 0 °C. The reaction was quenched by the addition of saturated NaHCO₃ (300 mL), filtered over Celite and the mixture was extracted with Et₂O (3 x 100 mL). The combined organic layers were dried over anhydrous MgSO₄ and the solvent was removed *in vacuo* affording crude furan (13 g, 100%), which was used without further purification in the next step. A small sample was purified by column chromatography (hexane : ethylacetate from 5 : 1 to 3 : 1) to obtain an analytically pure sample. ¹H-NMR (400MHz, CDCl₃): δ = 7.70-7.65 (m, 4H), 7.45-7.33 (m, 6H), 6.90 (s, 1H), 6.23 (d, *J* = 0.5 Hz, 1H), 5.52 (s, 1H), 3.84 (s, 3H), 3.69 (t, *J* = 6.3 Hz, 2H), 2.96-2.86 (m, 1H), 2.45 (s, 3H), 1.93-1.82 (m, 1H), 1.74-1.63 (m, 1H), 1.67-1.57 (m, 2H), 1.31 (d, *J* = 6.8 Hz, 3H), 1.07 (s, 9H); ¹³C-NMR (100MHz,

CDCl₃): δ = 164.0, 147.9, 145.1, 142.4, 135.6, 134.0, 129.5, 127.6, 124.1, 113.8, 102.0, 100.7, 63.8, 61.4, 33.3, 31.6, 30.0, 26.9, 19.2, 19.1, 9.3; IR (film): 3529, 2933, 2858, 1607, 1459, 1427, 1360, 1111, 864 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₃₁H₃₈O₄Si, 502.2539, found, 502.2537; [α]_D²⁰ +12.1 (c 2.45, CH₂Cl₂).

Crude furan (13 g, 25.95 mmol, 1.0 equiv.) in DMF (130 mL) was cooled to 0 °C. Then NaH (1.5 g, 38.85 mmol, 1.5 equiv.) was added in small portions, followed by the careful addition of neat MOMCl (2.75 mL, 36.26 mmol, 1.4 equiv.). The dark-brown solution was stirred for 1 h, diluted with Et₂O (200 mL) and quenched with saturated aqueous NH₄Cl (150 mL). The product was extracted with Et₂O : hexane = 1 : 1 (3 x 50 mL), washed with brine and dried over anhydrous MgSO₄. The solvent was evaporated and the pale yellow oil was purified by column chromatography (hexane : ethylacetate = 3 : 1) to furnish furan **49** (13.4 g, 95%). ¹H-NMR (400MHz, CDCl₃): δ = 7.68-7.62 (m, 4H), 7.44-7.31 (m, 6H), 7.07 (s, 1H), 6.22 (s, 1H), 5.21 (s, 2H), 3.84 (s, 3H), 3.67 (t, *J* = 6.2 Hz, 2H), 3.55 (s, 3H), 2.93-2.86 (m, 1H), 2.40 (s, 3H), 1.90-1.80 (m, 1H), 1.71-1.59 (m, 1H), 1.63-1.54 (m, 2H), 1.29 (d, *J* = 7.1 Hz, 3H), 1.04 (s, 9H); ¹³C-NMR (100MHz, CDCl₃): δ = 164.0, 149.2, 146.9, 145.5, 135.5, 134.0, 129.5, 127.6, 123.4, 115.3, 105.0, 100.8, 96.2, 63.8, 61.0, 56.1, 33.3, 31.6, 30.0, 26.8, 19.2, 19.1, 9.1; IR (film): 2932, 1684, 1653, 1559, 1473, 1427, 1260, 1153, 1112, 1044 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₃₃H₄₂O₅Si, 546.2802, found, 546.2792; [α]_D²⁰ +12.6 (c 1.40, CH₂Cl₂).

(S)-4-(6-Methoxy-5-(methoxymethoxy)-7-methylbenzofuran-2-yl)pentanoic acid (43). A solution of benzofuran **49** (7.63 g, 13.93 mmol, 1.0 equiv.) in THF (280 mL) was treated with TBAF (1 M in THF, 15.33 mL, 15.33 mmol, 1.1 equiv.) and stirred overnight at room temperature. Finally the reaction was quenched with NH₄Cl (150 mL) and the aqueous layer was extracted with Et₂O (3 x 100 mL). The combined organic extracts were dried over MgSO₄, filtered and evaporated to dryness. Purification by column chromatography using gradient elution (hexane : ethylacetate from 3 : 1 to 1 : 1) furnished the alcohol as a pale yellow oil (3.84 g, 89%). ¹H-NMR (400MHz, CDCl₃): δ = 7.07 (s, 1H), 6.27 (s, 1H), 5.21 (s, 2H), 3.84 (s, 3H), 3.64 (t, *J* = 6.3 Hz, 2H), 3.54 (s, 3H), 2.99-2.89 (m, 1H), 2.42 (s, 3H), 1.88-1.77 (m, 1H), 1.73-1.63 (m, 1H), 1.65-1.54 (m, 2H), 1.32 (d, *J* = 7.1 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ = 163.7, 149.2, 146.9, 145.6, 123.4, 115.3, 105.0, 101.0, 96.2, 62.9, 61.0, 56.1, 33.5, 31.6, 30.3, 19.1, 9.1; IR (film): 3854, 3676, 2935, 1653, 1559, 1457, 1153, 1043 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₁₇H₂₂O₅, 308.1624, found, 308.1620; [α]_D²⁰ +13.4 (c 1.60, CH₂Cl₂).

A 100 mL Schlenk flask was charged with above alcohol (3.57 g, 11.58 mmol, 1.0 equiv.) and DMSO (60 mL, 0.2 M). IBX (8.1 g, 28.94 mmol, 2.5 equiv.) was added over a period of 20 min and stirring was continued for 2 h at ambient temperature. The solution was diluted with Et₂O : hexane = 1 : 1 (100 mL) and H₂O (100 mL). The mixture was filtered over Celite, the layers were separated and the aqueous layer was extracted with Et₂O : hexane = 1 : 1 (3 x 50 mL). The combined organic layers were dried over anhydrous MgSO₄, filtered and concentrated *in vacuo*. The crude product was filtered over a plug of silica to give pure aldehyde as a pale orange oil (3.43 g, 97%). ¹H-NMR (400MHz, CDCl₃): δ = 9.74 (t, *J* = 1.4 Hz, 1H), 7.08 (s, 1H), 6.28 (s, 1H), 5.21 (s, 2H), 3.84 (s, 3H), 3.54 (s, 3H), 3.03-2.93 (m, 1H), 2.47 (dt, *J* = 7.5, 1.4 Hz, 2H), 2.41 (s, 3H), 2.10-1.93 (m, 2H), 1.34 (d, *J* = 7.1 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ = 202.0, 162.3, 149.3, 147.1, 145.8, 123.2, 115.4, 105.1, 101.6, 96.1, 61.0, 56.1, 41.6, 33.0, 27.7, 19.0, 9.1; IR (film): 2932, 1723, 1653, 1559, 1457, 1340, 1219, 1153, 1119, 1090, 1042 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₁₇H₂₂O₅, 306.1467, found, 306.1464; [α]_D²⁰ +17.2 (c 0.75, CH₂Cl₂).

Above aldehyde (3.43 g, 11.19 mmol, 1.0 equiv.) was dissolved in 75 mL *t*-BuOH (0.15 M), treated with 2-methyl-2-butene (1 mL / mmol, 11.2 mL), and cooled to 5 °C. NaClO₂ (18.9 g, 167.85 mmol, 15 equiv.) and 18.9 g NaH₂PO₄ were dissolved in H₂O (110 mL, 1.5 M), transferred to a 250 mL dropping funnel, and added over a period of 20 min. After 50 min at r.t. TLC analysis showed complete consumption and the reaction mixture was partitioned between CH₂Cl₂ (150 mL) and brine (100 mL). The aqueous layer was extracted with three portions of CH₂Cl₂ (50 mL) and the combined organic extracts were dried over MgSO₄. Evaporation of the solvent gave crude acid, which was purified by flash chromatography (hexane : ethylacetate from 3 : 1 to 1 : 1) to give acid **43** as an orange-viscous oil (3.60 g, 99%). ¹H-NMR (400MHz, CDCl₃): δ = 7.08 (s, 1H), 6.29 (s, 1H), 5.21 (s, 2H), 3.85 (s, 3H), 3.54 (s, 3H), 3.04-2.94 (m, 1H), 2.42-2.35 (m, 2H), 2.41 (s, 3H), 2.12-1.92 (m, 2H), 1.34 (d, *J* = 6.8 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ = 177.7, 162.4, 149.3, 147.0, 145.7, 123.2, 115.4, 105.1, 101.6, 96.2, 61.0, 56.1, 33.0, 31.4, 30.2, 19.0, 9.1; IR (film): 3629, 2933, 1707, 1653, 1607, 1559, 1457, 1420, 1261, 1153, 1117, 1043 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₁₇H₂₂O₆, 322.1416, found, 322.1421; [α]_D²⁰ +35.5 (c 0.65, CH₂Cl₂).

(S)-((3S,4S)-2,4-Dimethylhexa-1,5-dien-3-yl) 4-(6-methoxy-5-(methoxymethoxy)-7-methylbenzofuran-2-yl)pentanoate (50). A mixture of acid **43** (2.55 g, 7.91 mmol, 1.0 equiv.), alcohol **42** (1.20 g, 9.51 mmol, 1.2 equiv.), EDCl·HCl (1.97 g, 10.28 mmol, 1.3 equiv.) and DMAP (1.26 g, 10.31 mmol, 1.3 equiv.) in CH₂Cl₂ (30 mL) was stirred at room temperature for 1.5 h. The reaction was diluted with CH₂Cl₂ (100 mL), quenched with 1% HCl (20 mL) and washed with brine (2 x 50 mL). The organic layer was dried over MgSO₄, concentrated *in vacuo* and the residue was purified by flash chromatography (hexane : ethylacetate from 15 : 1 to 5 : 1) to give ester **50** (2.75 g, 81%).

¹H-NMR (400MHz, CDCl₃): δ = 7.07 (s, 1H), 6.27 (s, 1H), 5.70 (ddd, J = 17.2, 10.2, 8.0 Hz, 1H), 5.21 (s, 2H), 5.06-4.98 (m, 2H), 5.05 (d, J = 7.8 Hz, 1H), 4.96-4.91 (m, 2H), 3.84 (s, 3H), 3.54 (s, 3H), 3.02-2.91 (m, 1H), 2.52-2.43 (m, 1H), 2.41 (s, 3H), 2.36-2.30 (m, 2H), 2.10-1.99 (m, 1H), 1.99-1.89 (m, 1H), 1.72 (s, 3H), 1.32 (d, J = 7.1 Hz, 3H), 0.96 (d, J = 7.1 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ = 172.5, 162.7, 149.3, 147.0, 145.7, 141.8, 139.8, 123.3, 115.2, 114.4, 113.4, 105.1, 101.4, 96.2, 80.3, 61.0, 56.1, 40.0, 33.0, 32.1, 30.5, 18.9, 18.2, 16.6, 9.1; IR (film): 2967, 1734, 1700, 1684, 1653, 1559, 1457, 1152, 1117 cm⁻¹; HRMS (ESI) (m/z): [M]⁺ calcd for C₂₅H₃₄O₆Na, 453.2253, found, 453.2269; [α]_D²⁰ +26.8 (c 1.30, CH₂Cl₂).

(2S,6S,E)-2-((S)-2-(6-Methoxy-5-(methoxymethoxy)-7-methylbenzofuran-2-yl)propyl)-4,6-dimethylocta-4,7-dien-1-ol (51). LHMS (1 M in THF, 11.1 mL, 12.08 mmol, 4 equiv.) was diluted with THF (12 mL), cooled to -78 °C and freshly distilled HMPA (7.5 mL) was slowly added via cannula. After 5 min ester **50** (2.3 g, 3.28 mmol, 1.0 equiv.) in THF (2.1 mL, 0.5 mL rinse) was transferred to a freshly prepared TBSCl-solution (3 M in THF, 6.56 mL, 19.68 mmol, 6 equiv.) and added dropwise to the above LHMS / HMPA mixture. The reaction mixture was stirred for 40 min at -78 °C, allowed to warm to 0 °C over 15 min, stirred for additional 5 min at room temperature and partitioned between H₂O (100 mL) and Et₂O (3 x 70 mL). The combined organic fractions were washed with brine, dried over MgSO₄ and evaporated to dryness. The crude ketene silyl acetal was dissolved in DMF (12 mL) and heated under microwave irradiation at 180 °C for 15 min. The mixture was partitioned between H₂O (100 mL) and Et₂O (100 mL), extracted with Et₂O (3 x 50 mL), washed with brine (50 mL), dried over MgSO₄ and concentrated under reduced pressure. The crude ester was dissolved in Et₂O (30 mL), transferred to an ice-bath and LiAlH₄ (4 M in Et₂O, 1.51 mL, 6.04 mmol, 2 equiv.) was added carefully via cannula. After 30 min at room temperature TLC analysis showed complete consumption of the starting material and the reaction mixture was quenched at 0 °C by slow addition of ethylacetate, diluted with Et₂O (100 mL) and washed with 1% HCl (100 mL). The aqueous layer was extracted with Et₂O (3 x 50 mL), and the combined organic fractions were washed with brine (50 mL), dried over MgSO₄ and the solvent was removed *in vacuo*. Purification by column chromatography (hexane : ethylacetate from 10 : 1 to 5 : 1) afforded alcohol **51** as a colorless oil (1.11 g, 89%, dr = 4 : 1 as determined by ¹H-NMR). **51**: ¹H-NMR (400MHz, CDCl₃): δ = 7.07 (s, 1H), 6.28 (s, 1H), 5.74 (ddd, J = 17.1, 10.5, 6.4 Hz, 1H), 5.21 (s, 2H), 5.05 (d, J = 8.8 Hz, 1H), 4.95 (dt, J = 17.2, 1.6 Hz, 1H), 4.89 (dt, J = 10.2, 1.5 Hz, 1H), 3.84 (s, 3H), 3.54 (s, 3H), 3.49 (d, J = 5.1 Hz, 2H), 3.12-2.97 (m, 2H), 2.41 (s, 3H), 2.13-1.99 (m, 2H), 1.88-1.78 (m, 1H), 1.78-1.69 (m, 1H), 1.62 (d, J = 1.3 Hz, 3H), 1.49-1.41 (m, 1H), 1.42-1.36 (br, OH), 1.31 (d, J = 7.1 Hz, 3H), 1.05 (d, J = 6.8 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ = 163.9, 149.2, 146.9, 145.6, 142.9, 133.3, 130.7, 123.4, 115.3, 112.0, 105.0, 100.9, 96.2, 66.1, 61.0, 56.1, 42.8, 37.7, 36.4, 36.2, 31.5, 20.5, 20.1, 16.2, 9.1; IR (film): 3451, 2927, 1559, 1449, 1340, 1219, 1154, 1116, 1091, 1044 cm⁻¹; HRMS (ESI) (m/z): [M]⁺ calcd for C₂₅H₃₆O₅, 416.2563, found, 416.2569; [α]_D²⁰ +9.5 (c 1.95, CH₂Cl₂). **R-51**: ¹H-NMR (400MHz, CDCl₃): δ = 7.07 (s, 1H), 6.26 (s, 1H), 5.75 (ddd, J = 17.0, 10.4, 6.4 Hz, 1H), 5.21 (s, 2H), 5.04 (dd, J = 8.8, 1.0 Hz, 1H), 4.98 (dt, J = 17.2, 1.6 Hz, 1H), 4.91 (dt, J = 10.1, 1.5 Hz, 1H), 3.84 (s, 3H), 3.57-3.52 (m, 2H), 3.54 (s, 3H), 3.11-3.00 (m, 2H), 2.41 (s, 3H), 2.05 (dd, J = 14.4, 6.8 Hz, 2H), 1.97 (dd, J = 13.5, 6.4 Hz, 1H), 1.79-1.69 (m, 2H), 1.62-1.51 (m, 1H), 1.55 (d, J = 1.3 Hz, 3H), 1.30 (d, J = 6.8 Hz, 3H), 1.04 (d, J = 6.8 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ = 163.8, 149.2, 147.0, 145.6, 142.9, 133.2, 130.7, 123.4, 115.3, 112.0, 105.0, 101.0, 96.2, 65.7, 61.0, 56.1, 42.7, 36.9, 36.3, 36.0, 31.4, 20.6, 20.3, 16.1, 9.1; IR (film): 3451, 2928, 1606, 1451, 1340, 1219, 1154, 1117, 1090, 1044 cm⁻¹; HRMS (ESI) (m/z): [M]⁺ calcd for C₂₅H₃₆O₅, 416.2563, found, 416.2565; [α]_D²⁰ -18.4 (c 1.10, CH₂Cl₂).

6-Methoxy-5-(methoxymethoxy)-7-methyl-2-((2S,4S,8S,E)-4,6,8-trimethyldeca-6,9-dien-2-yl)benzofuran (52). Alcohol **51** (370 mg, 0.89 mmol, 1.0 equiv.) was dissolved in CH₂Cl₂ (5 mL), cooled to 0 °C and treated with Et₃N (150 μ L, 1.06 mmol, 1.2 equiv.). After 5 min MsCl (80 μ L, 1.06 mmol, 1.2 equiv.) was added and stirring was continued for 30 min. The solution was poured onto H₂O (20 mL), extracted with CH₂Cl₂ (3 x 10 mL), washed with brine and dried over MgSO₄. The solvent was evaporated under reduced pressure and the crude mesylate was immediately redissolved in Et₂O (9 mL). LiAlH₄ (4 M in Et₂O, 670 μ L, 2.67 mmol, 3 equiv.) was carefully added to the ice cooled solution and the cloudy mixture was allowed to warm to room temperature over 30 min. After 2 h the reaction mixture was quenched at 0 °C by slow addition of ethylacetate, diluted with Et₂O (40 mL) and washed with 1% HCl (10 mL). The aqueous layer was extracted with Et₂O (3 x 10 mL), and the combined organic fractions were washed with brine (10 mL), dried over MgSO₄ and the solvent was removed *in vacuo*. Purification by flash chromatography (hexane : ethylacetate = 5 : 1) afforded diol **52** as an oil (310 mg, 89%). ¹H-NMR (400MHz, CDCl₃): δ = 7.07 (s, 1H), 6.24 (s, 1H), 5.76 (ddd, J = 17.2, 10.4, 6.1 Hz, 1H), 5.21 (s, 2H), 4.99-4.94 (m, 1H), 4.96 (dt, J = 17.3, 1.7 Hz, 1H), 4.88 (dt, J = 10.4, 1.6 Hz, 1H), 3.84 (s, 3H), 3.54 (s, 3H), 3.11-2.95 (m, 2H), 2.42 (s, 3H), 2.07 (dd, J = 13.3, 5.2 Hz, 1H), 1.79 (dd, J = 12.6, 8.1 Hz, 1H), 1.74-1.64 (m, 1H), 1.59-1.44 (m, 2H), 1.57 (d, J = 1.3 Hz, 3H), 1.27 (d, J = 6.8 Hz, 3H), 1.05 (d, J = 6.8 Hz, 3H), 0.83 (d, J = 6.3 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ = 164.7, 149.1, 146.8, 145.8, 142.9, 133.3, 130.1, 123.5, 115.3, 111.7, 105.0, 100.4, 96.2, 61.0, 56.1, 48.0, 43.0, 36.3, 31.3, 28.3, 20.6, 19.5, 19.1, 16.1, 9.1; IR (film): 2926, 1684, 1653, 1559, 1507, 1458, 1153, 1117, 1044 cm⁻¹; HRMS (ESI) (m/z): [M]⁺ calcd for C₂₅H₃₆O₄, 400.2614, found, 400.2607; [α]_D²⁰ +3.0 (c 1.35, CH₂Cl₂).

2R,4S,5R)-1-((R)-4-Benzyl-2-oxooxazolidin-3-yl)-5-hydroxy-2,4-dimethylhept-6-ene-1,3-dione (53). A 250 mL Schlenk flask was charged with acid-free Sn(OTf)₂ (5.3 g, 12.71 mmol, 1.1 equiv.) and CH₂Cl₂ (42 mL, 0.3 M). The white suspension was treated at -20 °C with Et₃N (1.76 mL, 12.71 mmol, 1.1 equiv.) whereupon the mixture turned pale yellow. After 5 min β -keto imide **26** (3.34 g, 11.54 mmol, 1.0 equiv.) in CH₂Cl₂ (19 mL, 0.6 M) was added dropwise and the clear solution was stirred for 1 h at -20 °C. Freshly distilled acrolein (2.31 mL, 34.62 mmol, 3 equiv.) was dissolved in CH₂Cl₂ (35 mL, 1 M) and slowly added at -78 °C. After 30 min at -78 °C, the yellow-orange solution was poured onto a cooled (0 °C) and vigorously stirred mixture of CH₂Cl₂ : 1 M NaHSO₄ (150 mL, 1 : 1). After 20 min at room temperature the aqueous phase was extracted with CH₂Cl₂ (3 x 50 mL), the organic phase was washed with saturated aqueous NaHCO₃, dried over MgSO₄ and concentrated. Purification of the residue by gradient flash chromatography (hexane : ethylacetate from 3 : 1 to 1 : 1) yielded **53** as a viscous oil (3.64 g, 91%, dr = 5 : 1 as determined by HPLC and ¹H-NMR). ¹H-NMR (400MHz, CDCl₃): δ = 7.37-7.27 (m, 3H), 7.23-7.17 (m, 2H), 5.82 (ddd, J = 16.9, 10.7, 5.9 Hz, 1H), 5.31 (d, J = 17.2 Hz, 1H), 5.20 (d, J = 10.6 Hz, 1H), 4.87 (q, J = 7.2 Hz, 1H), 4.80-4.72 (m, 1H), 4.49-4.43 (br, 1H), 4.30-4.24 (m, 1H), 4.19 (dd, J = 9.1, 3.0 Hz, 1H), 3.30 (dd, J = 13.5, 3.2 Hz, 1H), 2.93-2.84 (m, 1H), 2.78 (dd, J = 13.5, 9.6 Hz, 1H), 2.46(d, J = 3.3 Hz, OH), 1.48 (d, J = 7.3 Hz, 3H), 1.24 (d, J = 7.1 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ = 210.8, 170.4, 153.3, 137.5, 135.0, 129.3, 129.0, 127.4, 116.3, 72.7, 66.5, 55.3, 52.0, 48.9, 38.0, 12.8, 10.9; IR (film): 3629, 3510, 2984, 1773, 1684, 1653, 1559, 1456, 1362, 1214, 1121 cm⁻¹; HRMS (ESI) (m/z): [M]⁺ calcd for C₁₉H₂₃O₅NNa, 368.1473, found, 368.1477; [α]_D²⁰ -115.3 (c 1.6, CH₂Cl₂).

(3R,4S,5R,6R)-4-Hydroxy-3,5-dimethyl-6-vinyltetrahydro-2H-pyran-2-one (54). Me₄NBH(OAc)₃ (2.85 g, 10.85 mmol, 5 equiv.) was dissolved in 360 mL MeCN : AcOH = 1.9 : 1, cooled to -32 °C and aldol product **53** (750 mg, 2.17 mmol, 1.0 equiv.) in MeCN (6 mL) was added dropwise. The reaction was stirred for 3 h at -32 °C, allowed to warm to 0 °C overnight, diluted with CH₂Cl₂ (100 mL) and quenched by the addition of saturated aqueous Rochelle's salt (150 mL). Saturated aqueous NaHCO₃ was carefully added to the vigorously stirred solution over 20 min. After 1 h no more gas evolution was observed and the two-phase mixture was partitioned between CH₂Cl₂ (400 mL) and H₂O (100 mL). The aqueous layer was extracted with CH₂Cl₂ (3 x 100 mL), and the combined organic fractions were dried over MgSO₄, filtered and concentrated *in vacuo*. Purification of the residue by flash chromatography (hexane : ethylacetate = 3 : 1) afforded the diol as a viscous oil (527 mg, 70%, dr \geq 6 : 1 as determined by ¹H-NMR). ¹H-NMR (600MHz, CDCl₃): δ = 7.36-7.31 (m, 2H), 7.30-7.27 (m, 1H), 7.22-7.18 (m, 2H), 5.97 (ddd, J = 16.9, 10.9, 5.8 Hz, 1H), 5.32 (d, J = 17.4 Hz, 1H), 5.21 (d, J = 10.6 Hz, 1H), 4.73-4.68 (m, 1H), 4.34-4.30 (br, 1H), 4.27-4.22 (m, 1H), 4.20 (dd, J = 8.7, 2.6 Hz, 1H), 3.98 (d, J = 9.8 Hz, 1H), 3.84 (dq, J = 7.1, 1.9 Hz, 1H), 3.80-3.76 (br, OH), 3.41-3.34 (br, OH), 3.24 (dd, J = 13.5, 3.4 Hz, 1H), 2.80 (dd, J = 13.5, 9.3 Hz, 1H), 2.03-1.96 (m, 1H), 1.28 (d, J = 7.2 Hz, 3H), 0.84 (d, J = 7.2 Hz, 3H); ¹³C-NMR (150MHz, CDCl₃): δ = 178.0, 152.8, 137.9, 134.9, 129.4, 129.0, 127.5, 115.7, 75.7, 73.4, 66.2, 55.0, 39.3, 39.2, 37.8, 12.1, 9.7; IR (film): 3448, 2976, 1780, 1700, 1559, 1456, 1388, 1211 cm⁻¹; HRMS (ESI) (m/z): [M]⁺ calcd for C₁₉H₂₅O₃NNa, 370.1630, found, 370.1644; [α]_D²⁰ -30.5 (c 0.80, CH₂Cl₂).

A solution of the diol (1.98 g, 5.71 mmol, 1.0 equiv.) in THF-H₂O (80 mL, 3 : 1) was treated at 0 °C with H₂O₂ (30% in H₂O, 2.3 mL). LiOH (383 mg, 9.14 mmol, 1.6 equiv.) was added and stirring was continued for 1 h at ambient temperature. The reaction was acidified by the addition of 1 N HCl (20 mL) and stirred for further 5 min. The biphasic mixture was diluted with H₂O (100 mL), extracted with Et₂O (3 x 70 mL), dried over MgSO₄ and concentrated under reduced pressure. Purification by flash chromatography (hexane : ethylacetate from 3 : 1 to 1 : 1) gave lactone **54** (700 mg, 72%). ¹H-NMR (400MHz, CDCl₃): δ = 5.85 (ddd, J = 17.2, 11.0, 5.7 Hz, 1H), 5.40 (d, J = 17.2 Hz, 1H), 5.3 (d, J = 10.6 Hz, 1H), 4.80-4.74 (m, 1H), 3.88 (dd, J = 9.6, 4.0 Hz, 1H), 2.57-2.48 (m, 1H), 2.27-2.19 (m, 1H), 2.09-2.01 (br, OH), 1.41 (d, J = 7.1 Hz, 3H), 0.97 (d, J = 7.1 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ = 173.2, 133.6, 117.4, 79.9, 73.7, 60.4, 40.1, 37.9, 21.0, 14.4, 14.2, 5.5; IR (film): 3446, 2977, 1718, 1700, 1653, 1559, 1507, 1458, 1213, 1094 cm⁻¹; HRMS (ESI) (m/z): [M-H₂O]⁺ calcd for C₉H₁₂O₃, 152.0837, found, 152.0844; [α]_D²⁰ +103.8 (c 0.60, CH₂Cl₂).

(R)-Methyl 2-((4S,5S,6R)-2,2,5-trimethyl-6-vinyl-1,3-dioxan-4-yl)propanoate (55). Lactone **54** (700 mg, 4.11 mmol, 1.0 equiv.) was dissolved in 2,2-dimethoxypropane (40 mL), treated with camphorsulfonic acid (96 mg, 0.41 mmol, 0.1 equiv.) and stirred overnight at room temperature. The solution was diluted with Et₂O (100 mL), neutralized with saturated aqueous NaHCO₃ (50 mL), extracted with Et₂O (3 x 50 mL), washed with brine (70 mL) and dried over MgSO₄. The solvent was removed *in vacuo* and the residue was purified by flash chromatography (hexane : ethylacetate = 10 : 1) to yield ester **55** (903 mg, 91%). ¹H-NMR (400MHz, CDCl₃): δ = 5.79 (ddd, J = 17.1, 10.7, 6.1 Hz, 1H), 5.25 (dt, J = 17.3, 1.7 Hz, 1H), 5.16 (dt, J = 10.5, 1.6 Hz, 1H), 4.39-4.35 (m, 1H), 3.71-3.66 (m, 1H), 3.69 (s, 3H), 2.59 (dq, J = 6.99, 4.99 Hz, 1H), 2.00-1.90 (m, 1H), 1.34 (s, 3H), 1.33 (s, 3H), 1.21 (d, J = 7.1 Hz, 3H), 0.83 (d, J = 7.1 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ = 174.8, 134.7, 115.7, 100.7, 75.1, 70.8, 51.7, 42.9, 37.4, 25.2, 23.7, 12.8, 11.4; IR (film): 2988, 1740, 1700, 1653, 1559, 1458, 1301, 1226, 1176, 1025, 1001 cm⁻¹; HRMS (ESI) (m/z): [M-Me]⁺ calcd for C₁₂H₁₉O₄, 227.1283, found, 227.1289; [α]_D²⁰ -20.3 (c 1.05, CH₂Cl₂).

(R)-2-((4S,5S,6R)-2,2,5-Trimethyl-6-vinyl-1,3-dioxan-4-yl)propanal (46). Ester **55** (900 mg, 3.71 mmol, 1.0 equiv.) in Et₂O (40 mL) was cooled to 0 °C and LiAlH₄ (4 M in Et₂O, 1.49 mL, 5.57 mmol, 1.5 equiv.) was carefully added *via* cannula. After 30 min at 0 °C TLC analysis showed complete consumption of the starting material and the reaction mixture was quenched by slow addition of ethylacetate, diluted with Et₂O (80 mL) and washed with 1% HCl (100 mL). The aqueous layer was extracted with Et₂O (3 x 50 mL), and the combined organic fractions were washed with brine (50 mL), dried over MgSO₄ and concentrated *in vacuo*. Purification by column chromatography (hexane : ethylacetate = 3 : 1) afforded the alcohol as a colorless oil (761 mg, 96%). ¹H-NMR (400MHz, CDCl₃): δ = 5.80 (ddd, *J* = 17.1, 10.1, 6.2 Hz, 1H), 5.26(dt, *J* = 17.3, 1.6 Hz, 1H), 5.16 (dt, *J* = 10.8, 1.6 Hz, 1H), 4.39-4.34 (m, 1H), 3.70-3.65 (m, 2H), 3.57 (dd, *J* = 8.1, 2.7 Hz, 1H), 2.36 (t, *J* = 5.4 Hz, OH), 2.01-1.92 (m, 1H), 1.90-1.81 (m, 1H), 1.38 (s, 3H), 1.35 (s, 3H), 0.99 (d, *J* = 7.0 Hz, 3H), 0.83 (d, *J* = 6.8 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ = 135.7, 115.7, 100.6, 76.8, 71.1, 67.1, 37.2, 36.5, 25.4, 23.8, 13.0, 10.6; IR (film): 3423, 2987, 1684, 1653, 1559, 1507, 1457, 1380, 1226, 1180, 1027 cm⁻¹; HRMS (ESI) (*m/z*): [M-Me]⁺ calcd for C₁₁H₁₉O₃, 199.1334, found, 199.1332; [α]_D²⁰ +11.8 (c 0.80, CH₂Cl₂).

A solution of the above prepared alcohol (23 mg, 0.107 mmol, 1.0 equiv.) in CH₂Cl₂ (0.5 mL, 0.2 M) was cooled to -5 °C. Et₃N (45 μL, 0.321 mmol, 3 equiv.) and subsequently SO₃ Pyr (51 mg, 0.321 mmol, 3 equiv.) in DMSO (0.5 mL, 0.6 M) were added dropwise. The mixture was stirred for 1.5 h at -5 °C and quenched with aqueous 1 M KHSO₄ solution (0.5 mL). The phases were partitioned between brine and Et₂O (1 : 1, 40 mL) and the aqueous layer was extracted with Et₂O (3 x 10 mL). The combined organic fractions were concentrated to 5 mL under reduced pressure, filtered over a plug of silica and excess solvent was removed *in vacuo* to afford aldehyde **46** (23 mg, 99%). ¹H-NMR (400MHz, CDCl₃): δ = 9.71 (d, *J* = 1.0 Hz, 1H), 5.80 (ddd, *J* = 17.1, 10.7, 6.2 Hz, 1H), 5.27(dt, *J* = 17.3, 1.7 Hz, 1H), 5.18 (dt, *J* = 10.6, 1.6 Hz, 1H), 4.40-4.35 (m, 1H), 3.84 (dd, *J* = 8.1, 3.3 Hz, 1H), 2.44 (ddq, *J* = 7.0, 3.2, 0.9 Hz, 1H), 2.04-1.94 (m, 1H), 1.36 (s, 3H), 1.33 (s, 3H), 1.17 (d, *J* = 7.0 Hz, 3H), 0.87 (d, *J* = 6.8 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ = 204.2, 135.4, 115.9, 100.8, 73.2, 70.9, 48.6, 36.7, 25.1, 23.8, 12.7, 7.8; IR (film): 2986, 1734, 1684, 1653, 1559, 1507, 1458, 1380, 1225 cm⁻¹; HRMS (ESI) (*m/z*): [M-Me]⁺ calcd for C₁₁H₁₇O₃, 197.1178, found, 197.1171; [α]_D²⁰ -39.0 (c 0.70, CH₂Cl₂).

(S)-1-(6-Methoxy-5-(methoxymethoxy)-7-methyl-2-((2S,4S,8S,E)-4,6,8-trimethyldeca-6,9-dien-2-yl)benzofuran-4-yl)-2-((4R,5S,6R)-2,2,5-trimethyl-6-vinyl-1,3-dioxan-4-yl)propan-1-ol (9a,b). To a solution of benzofuran **52** (100 mg, 0.250 mmol, 1.4 equiv.) in THF (0.6 mL) freshly distilled TMEDA (80 μL, 0.535 mmol, 3 equiv.) was added at ambient temperature. The solution was cooled to -78 °C and *n*-BuLi (156 μL, 0.250 mmol, 1.4 equiv.) was added dropwise. After 1.5 h at -30 °C the orange solution was recooled to -78 °C and aldehyde **46** (38 mg, 0.178 mmol, 1.0 equiv.) in THF (0.5 mL) was added *via* cannula. The reaction mixture was warmed to -25 °C over 2 h, diluted with Et₂O (40 mL) and finally quenched with saturated aqueous NH₄Cl solution (10 mL). The reaction mixture was extracted with diethyl ether (3 x 10 mL), dried over MgSO₄ and the solvent was removed *in vacuo*. The residue was purified by column chromatography (hexane : ethylacetate from 20 : 1 to 5 : 1) to furnish 97 mg (90%, dr = 4 : 1 as determined by ¹H-NMR) of alcohols **9a** and **9b**. Separation of the diastereomers for analytical purpose was done by HPLC, yielding diastereomer **9a** and **9b** as light orange, viscous oils. **9a**: ¹H-NMR (600MHz, CDCl₃): δ = 6.63 (s, 1H), 5.79-5.72 (m, 2H), 5.21 (dt, *J* = 17.0, 1.7 Hz, 1H), 5.17 (dd, *J* = 6.0, 4.9 Hz, 1H), 5.13 (dt, *J* = 10.6, 1.7 Hz, 1H), 5.10 (d, *J* = 5.7 Hz, 1H), 5.09 (d, *J* = 5.7 Hz, 1H), 4.98-4.95 (m, 1H), 4.95 (dt, *J* = 17.4, 1.7 Hz, 1H), 4.88 (dt, *J* = 10.2, 1.5 Hz, 1H), 4.33 (t, *J* = 5.7 Hz, 1H), 3.77 (s, 3H), 3.56 (s, 3H), 3.44 (d, *J* = 4.5 Hz, OH), 3.32 (d, *J* = 8.5, 1.3 Hz, 1H), 3.09-3.03 (m, 1H), 3.03-2.96 (m, 1H), 2.39 (s, 3H), 2.35-2.29 (m, 1H), 2.09 (dd, *J* = 12.8, 5.7 Hz, 1H), 1.98-1.91 (m, 1H), 1.78 (dd, *J* = 13.2, 7.9 Hz, 1H), 1.72-1.65 (m, 1H), 1.61-1.54 (m, 1H), 1.58 (d, *J* = 1.1 Hz, 3H), 1.50-1.44 (m, 1H), 1.33 (s, 3H), 1.26 (d, *J* = 6.8 Hz, 3H), 1.13 (d, *J* = 6.8 Hz, 3H), 1.11 (s, 3H), 1.05 (d, *J* = 6.8 Hz, 3H), 0.82 (d, *J* = 6.4 Hz, 3H), 0.72 (d, *J* = 6.8 Hz, 3H); ¹³C-NMR (150MHz, CDCl₃): δ = 164.0, 150.1, 147.0, 144.0, 143.3, 135.8, 133.3, 130.1, 125.1, 123.0, 115.5, 114.2, 111.7, 101.3, 100.5, 99.9, 76.8, 74.2, 71.7, 60.6, 57.4, 47.9, 43.0, 40.7, 37.1, 36.3, 31.2, 28.3, 25.3, 23.7, 20.6, 19.5, 19.1, 16.0, 12.4, 9.1, 8.8; IR (film): 3497, 2965, 2930, 1844, 1636, 1458, 1381, 1224, 1159, 1116, 1054 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₃₇H₅₆O₇Na, 635.3924, found, 635.3919; [α]_D²⁰ -3.9 (c 0.95, CH₂Cl₂). **9b**: ¹H-NMR (600MHz, CDCl₃): δ = 6.51(s, 1H), 5.85 (ddd, *J* = 17.1, 10.7, 6.3 Hz, 1H), 5.76 (ddd, *J* = 17.2, 10.4, 6.0 Hz, 1H), 5.27 (dt, *J* = 17.4, 1.7 Hz, 1H), 5.17 (dd, *J* = 10.8, 1.6 Hz, 1H), 5.13-5.08 (m, 3H), 4.98-4.95 (m, 1H), 4.96 (dt, *J* = 17.4, 1.7 Hz, 1H), 4.58 (d, *J* = 10.2, 1.7 Hz, 1H), 4.40 (t, *J* = 5.5 Hz, 1H), 3.97 (dd, *J* = 7.9, 1.5 Hz, 1H), 3.79 (s, 3H), 3.60 (s, 3H), 3.24-3.10 (br, OH), 3.10-3.03 (m, 1H), 3.03-2.97 (m, 1H), 2.40 (s, 3H), 2.35-2.29 (m, 1H), 2.07 (dd, *J* = 13.0, 5.9 Hz, 1H), 2.03-1.97 (m, 1H), 1.79 (dd, *J* = 12.8, 8.3 Hz, 1H), 1.73-1.67 (m, 1H), 1.58-1.54 (m, 1H), 1.57 (d, *J* = 1.5 Hz, 3H), 1.53-1.49 (m, 1H), 1.44 (s, 3H), 1.39 (s, 3H), 1.28 (d, *J* = 6.8 Hz, 3H), 1.05 (d, *J* = 6.8 Hz, 3H), 0.87 (d, *J* = 6.8 Hz, 3H), 0.84 (d, *J* = 6.4 Hz, 3H), 0.72 (d, *J* = 6.8 Hz, 3H); ¹³C-NMR (150MHz, CDCl₃): δ = 164.2, 150.1, 147.5, 144.4, 143.3, 136.1, 133.3, 130.1, 125.5, 123.0, 115.6, 114.4, 111.7, 100.6, 100.4, 100.2, 73.5, 71.4, 70.7, 60.7, 57.7, 48.0, 42.8, 41.0, 36.8, 36.3, 31.2, 28.2, 25.6, 24.1, 20.6, 19.4, 18.9, 16.0, 12.9, 10.7, 9.1; IR (film): 3469, 2965, 2930, 1457, 1380, 1340, 1226, 1160, 1116, 1023 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₃₇H₅₆O₇Na, 635.3924, found, 635.3915; [α]_D²⁰ -13.1 (c 0.90, CH₂Cl₂).

Macrocycle 56. Compound **9a** (80 mg, 0.131 mmol, 1.0 equiv.) was dissolved in degassed CH₂Cl₂ (130 mL) and heated to reflux. Grubbs' II catalyst (22 mg, 0.026 mmol, 0.2 equiv.) in degassed CH₂Cl₂ (15 mL) was added *via* syringe pump within 16 h. After completion of the addition the mixture was stirred for another 30 min. The temperature was lowered to room temperature and air was bubbled through the solution to destroy excess catalyst. The solvent was evaporated and purification by column chromatography (hexane : ethylacetate from 10 : 1 to 5 : 1) afforded 47 mg (62%) of macrocycle **56** (rotamers) as a white foam. ¹H-NMR (600MHz, CDCl₃): δ = 6.42 (s, 1H), 5.78 (dd, *J* = 15.5, 4.2 Hz, 1H), 5.31 (dd, *J* = 15.5, 9.1 Hz, 1H), 5.10 (dd, *J* = 9.3, 4.7 Hz, 1H), 5.09 (d, *J* = 5.7 Hz, 1H), 5.03-4.95 (br, 1H), 4.82 (d, *J* = 8.7 Hz, 1H), 4.16 (dd, *J* = 8.7, 6.0 Hz, 1H), 3.78 (s, 3H), 3.58 (s, 3H), 3.54 (dd, *J* = 9.6, 3.6 Hz, 1H), 3.11-3.04 (m, 1H), 3.04-2.98 (m, 1H), 2.46-2.37 (m, 1H), 2.42 (s, 3H), 2.21 (d, *J* = 14.7 Hz, 1H), 2.11-2.03 (br, 1H), 2.02-1.92 (br, 1H), 1.72-1.58 (br, 2H), 1.51 (s, 3H), 1.48-1.41 (m, 1H), 1.29-1.18 (m, 12H), 1.01-0.95 (m, 6H), 0.72-0.53 (br, 3H); ¹H-NMR (400MHz, C₇D₈, 350 K): δ = 6.44 (s, 1H), 5.65 (ddd, *J* = 15.5, 4.9 Hz, 1H), 5.44 (ddd, *J* = 15.5, 8.2, 1.7 Hz, 1H), 5.32 (d, *J* = 8.2 Hz, 1H), 5.03 (d, *J* = 5.6 Hz, 1H), 4.99 (d, *J* = 5.6 Hz, 1H), 4.88 (d, *J* = 8.8 Hz, 1H), 4.23 (dd, *J* = 8.0, 6.1 Hz, 1H), 3.69 (dd, *J* = 8.8, 4.5 Hz, 1H), 3.66 (s, 3H), 3.33 (s, 3H), 2.99-2.89 (m, 2H), 2.72-2.63 (br, OH), 2.59-2.49 (m, 1H), 2.40 (s, 3H), 2.16-2.07 (m, 2H), 1.94 (ddd, *J* = 13.7, 8.9, 5.4 Hz, 1H), 1.80-1.70 (m, 1H), 1.70-1.60 (m, 1H), 1.52-1.41 (m, 1H), 1.50 (s, 3H), 1.47 (d, *J* = 6.6 Hz, 3H), 1.34 (s, 3H), 1.26 (d, *J* = 6.9 Hz, 3H), 0.99 (d, *J* = 6.4 Hz, 3H), 0.94 (d, *J* = 6.8 Hz, 3H), 0.90 (s, 3H), 0.70 (d, *J* = 7.0 Hz, 3H); ¹³C-NMR (100MHz, C₇D₈, 350 K): δ = 163.6, 149.0, 146.6, 139.5, 137.9, 133.4, 128.6, 127.9, 127.2, 122.8, 114.9, 102.5, 100.9, 99.1, 75.2, 71.9, 60.6, 57.5, 45.4, 43.4, 40.4, 35.4, 34.8, 32.6, 30.0, 29.2, 26.1, 21.4, 21.2, 21.0, 19.3, 17.0, 13.2, 11.8, 9.5; IR (film): 3440, 2960, 1683, 1652, 1557, 1455, 1378, 1163, 1113 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₃₅H₅₂O₇Na, 607.3612, found, 607.3616; [α]_D²⁰ +53.9 (c 1.20, CH₂Cl₂).

4-(2R,3R,4S,5R,6R)-4-Hydroxy-3,5-dimethyl-6-vinyltetrahydro-2H-pyran-2-yl)-6-methoxy-7-methyl-2-((2S,4S,8S,E)-4,6,8-trimethyldeca-6,9-dien-2-yl)benzofuran-5-ol (59). A mixture of **9a,b** (200 mg, 0.326 mmol) was dissolved in MeOH (7 mL) and treated with 3 drops of 3 N HCl. The reaction mixture was stirred at room temperature overnight, diluted with H₂O (50 mL) and extracted with CH₂Cl₂ (4 x 20 mL). The organic extracts were dried over MgSO₄, concentrated *in vacuo* and purified by flash chromatography (hexane : ethylacetate from 5 : 1 to 3 : 1) to afford tetrahydropyran **59** (120 mg, 72%, 1.5 : 1 rotamers) as a white foam. ¹H-NMR (Rotamers, 600MHz, CDCl₃): δ = 7.67 (br, 0.5OH)+5.60 (br, 0.5OH), 6.55(br, 0.5H)+6.18(br, 0.5H), 5.88-5.79 (m, 1H), 5.76 (ddd, *J* = 17.4, 10.2, 6.0 Hz, 1H), 5.27 (d, *J* = 16.6 Hz, 1H), 5.17 (br, 1H), 4.98-4.93 (m, 2H), 4.90-4.87 (m, 1H), 4.75 (br, 0.5H)+4.49 (br, 0.5H), 4.23 (dd, *J* = 4.0, 1.7 Hz, 1H), 3.84 (br, 3H), 3.70 (br, 1H), 3.10-3.02 (m, 1H), 3.02-2.96 (m, 1H), 2.42 (s, 3H), 2.25-1.95 (br, 1H), 2.16-2.11 (br, 1H), 2.08 (dd, *J* = 13.0, 5.5 Hz, 1H), 1.78 (dd, *J* = 12.3, 8.5 Hz, 1H), 1.73-1.65 (m, 1H+OH), 1.63-1.55 (m, 1H), 1.57 (s, 3H), 1.51-1.45 (m, 1H), 1.27 (d, *J* = 6.8 Hz, 3H), 1.09 (br, 3H), 1.05 (d, *J* = 6.8 Hz, 3H), 0.83 (d, *J* = 6.4 Hz, 3H), 0.82 (d, *J* = 6.4 Hz, 3H); ¹³C-NMR (The asterisk denotes signals not apparent in the ¹³C-spectrum, 150MHz, CDCl₃): δ = 163.7, 146.9*, 143.3, 136.4*, 133.3, 130.1, 115.2*, 114.3*, 111.7, 98.9*, 82.8*, 80.2*, 76.8*, 61.3*, 48.1*, 47.9, 42.9, 39.1, 37.7*, 36.3, 31.3, 28.3, 20.6, 19.5, 19.2, 16.0, 13.6*, 9.2, 6.6*; IR (film): 3391, 2964, 2927, 1636, 1604, 1455, 1405, 1384, 1284, 1114, 1048 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₃₂H₄₆O₅, 510.3345, found, 510.3331; [α]_D²⁰ +87.2 (c 1.75, CH₂Cl₂).

Diolefin 60. Compound **59** (50 mg, 0.098 mmol, 1.0 equiv.) was dissolved in degassed CH₂Cl₂ (100 mL) and heated to reflux (45 °C outside temperature). Grubbs' II catalyst (16.6 mg, 0.020 mmol, 0.2 equiv.) in degassed CH₂Cl₂ (13 mL) was added *via* syringe pump within 16 h. After completion of the addition the mixture was stirred for another 30 min. The temperature was lowered to room temperature and air was bubbled through the solution to destroy excess catalyst. The solvent was evaporated and purification by column chromatography (hexane : ethylacetate from 5 : 1 to 3 : 1) afforded 39 mg (83%, 15 : 1 = *E* : *Z*) of macrocycle **60**. The mixture was used in the next step without further purification. ¹H-NMR (400MHz, CDCl₃): δ = 6.66 (s, 1H), 5.59 (ddd, *J* = 15.6, 8.7, 2.0 Hz, 1H), 5.54 (s, OH), 5.33 (dd, *J* = 15.5, 2.1 Hz, 1H), 4.92 (d, *J* = 9.1 Hz, 1H), 4.74 (d, *J* = 10.1 Hz, 1H), 4.27 (dd, *J* = 4.4, 2.1 Hz, 1H), 3.81 (s, 3H), 3.76-3.69 (m, 1H), 3.17-3.04 (m, 1H), 3.04-2.93 (m, 1H), 2.44 (s, 3H), 2.12-2.04 (m, 1H), 1.95 (dd, *J* = 12.8, 4.9 Hz, 2H), 1.87-1.79 (m, 1H), 1.79-1.71 (m, 1H), 1.69 (s, 3H), 1.66-1.45 (m, 2H), 1.55 (br, OH), 1.33 (d, *J* = 7.1 Hz, 3H), 1.04 (d, *J* = 7.1 Hz, 3H), 1.00 (d, *J* = 6.8 Hz, 3H), 0.97 (d, *J* = 6.8 Hz, 3H) 0.95 (d, *J* = 6.8 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ = 162.0, 148.3, 141.8, 136.8, 133.0, 130.2, 125.2, 122.4, 115.5, 112.9, 104.6, 76.7, 75.7, 61.4, 44.4, 43.4, 38.9, 38.4, 35.8, 31.0, 28.8, 22.1, 21.3, 19.2, 18.1, 12.8, 9.4, 7.0; IR (film): 3450, 2967, 1683, 1653, 1456, 1404, 1380, 1321, 1109 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₃₀H₄₂O₅, 482.3032, found, 482.3023;

Tetrahydropyran 5. To a vigorously stirred refluxing solution of **60** (37 mg, 0.076 mmol, 1.0 equiv.) and AcOH (11 μL, 0.192 mmol, 2.5 equiv.) in CH₂Cl₂ (7 mL) was added dipotassium azodicarboxylate (89 mg, 0.460 mmol, 6 equiv.) over a period of 6 h. The mixture was cooled to room temperature, filtered over Celite and concentrated *in vacuo*. The crude product was purified by HPLC (hexane : ethylacetate = 4 : 1) to give **5** as a white foam (28 mg, 76%). All analytical data matched with those reported by Lee^[6a] and Rychnovsky.^[7] ¹H-NMR (600MHz, CDCl₃): δ = 6.55 (s, 1H), 5.53 (s, 1H), 4.60 (d, *J* = 9.5 Hz, 1H), 4.54 (d, *J* = 10.2 Hz, 1H), 3.83 (s, 3H), 3.67-3.62 (m, 1H), 3.44 (ddd, *J* = 11.0, 2.3, 1.1 Hz, 1H), 3.11-3.04 (m, 1H), 2.47-2.41 (m, 1H), 2.45 (s, 3H),

2.26-2.19 (m, 1H), 1.93-1.88 (m, 1H), 1.84-1.77 (m, 1H), 1.62 (s, 3H), 1.61-1.54 (m, 1H), 1.53-1.49 (br, OH), 1.48-1.41 (m, 2H), 1.38 (d, $J=6.8$ Hz, 3H), 1.35-1.18 (m, 5H), 1.04 (d, $J=7.0$ Hz, 3H), 0.90 (d, $J=6.6$ Hz, 3H), 0.81 (d, $J=6.4$ Hz, 3H), 0.76 (d, $J=6.4$ Hz, 3H); ^{13}C -NMR (150MHz, CDCl_3): $\delta = 159.7, 148.2, 141.6, 141.5, 131.5, 129.0, 122.1, 115.7, 112.5, 104.7, 77.8, 77.3$ (2xCH), 61.4, 43.8, 41.8, 39.6, 38.6, 33.7, 32.5, 31.5, 31.1, 27.5, 21.8, 21.0, 19.6, 18.7, 12.8, 9.4, 6.6; IR (film): 3463, 2924, 2854, 1457, 1375, 1325, 1109, 1001 cm^{-1} ; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{30}\text{H}_{40}\text{O}_3\text{Na}$, 507.3086, found, 507.3082; $[\alpha]_{\text{D}}^{20} +19.4$ (c 0.17, CHCl_3)

O-quinone 71. Macrocycle **5** (6 mg, 0.012 mmol, 1 equiv.) was dissolved in 1 mL $\text{CH}_2\text{Cl}_2 / \text{H}_2\text{O}$ (10 : 1) and treated with DDQ (4.2 mg, 0.019 mmol, 1.5 equiv.) at room temperature. The color of the solution turned dark purple within 15 min, whereas TLC analysis showed complete consumption of the starting material. The mixture was directly loaded onto a silica column and eluted (hexane : ethylacetate from 3 : 1 to 2 : 1), to collect purple-blue fractions. The solvent was carefully evaporated to afford labile o-quinone **71** (3 mg, 52%) as a violet-blue compound. ^1H -NMR (400MHz, CDCl_3): $\delta = 6.11$ (s, 1H), 4.72 (d, $J=9.8$ Hz, 1H), 4.17 (d, $J=10.0$ Hz, 1H), 3.55-3.48 (m, 1H), 3.32-3.26 (m, 1H), 2.97-2.87 (m, 1H), 2.34-2.27 (m, 1H), 2.27-2.21 (m, 1H), 1.90 (s, 3H), 1.88-1.80 (m, 1H), 1.81-1.73 (m, 1H), 1.68-1.54 (m, 3H), 1.62 (s, 3H), 1.47-1.37 (m, 1H), 1.44-1.31 (m, 1H), 1.36-1.24 (m, 2H), 1.19-1.09 (m, 1H), 1.25 (d, $J=6.8$ Hz, 3H), 0.91 (d, $J=6.1$ Hz, 3H), 0.90 (d, $J=6.8$ Hz, 3H), 0.89 (d, $J=6.5$ Hz, 3H), 0.80 (d, $J=6.5$ Hz, 3H); ^{13}C -NMR (100MHz, CDCl_3): $\delta = 177.2, 173.7, 164.3, 147.4, 131.3, 129.1, 125.4, 113.7, 105.3, 78.3, 76.4, 75.8, 42.1, 41.7, 39.4, 38.2, 33.8, 32.5, 32.1, 29.7, 27.7, 21.8, 21.0, 19.6, 17.9, 17.0, 13.0, 8.2, 6.4$; IR (film): 3625, 2924, 2359, 1732, 1699, 1652, 1584, 1455, 1377, 1326, 1094 cm^{-1} ; HRMS (ESI) (m/z): $[\text{M}+\text{MeCN}+\text{Na}]^+$ calcd for $\text{C}_{31}\text{H}_{43}\text{O}_3\text{NNa}$, 532.3039, found, 532.3058.

Kendomycin 1. O-quinone **71** (2 mg, 0.0043 mmol) was dissolved in MeCN (2 mL) and treated with one drop of 1% HCl. The initial blue solution turned yellow within 15 min and the reaction mixture was partitioned between ethylacetate (50 mL) and brine (15 mL). The aqueous phase was extracted with ethylacetate (3 x 10 mL), the organic layer was dried over MgSO_4 and concentrated *in vacuo*. Purification by flash chromatography (hexane : ethylacetate from 3 : 1 to 2 : 1) gave kendomycin **1** (1 mg, 50%) as a yellow solid. m.p. 226-227 °C (authentic sample: 235-236 °C); $[\alpha]_{\text{D}}^{20} -76.4$ (c 0.11, MeOH), (lit. $[\alpha]_{\text{D}}^{20} -80$ (c 2.71, MeOH)^[2], $[\alpha]_{\text{D}}^{20} -79.3$ (c 0.135, MeOH)^[2b-d], $[\alpha]_{\text{D}}^{20} -82.4$ (c 0.514, MeOH)^[2a]), ^1H -NMR (600MHz, CD_3COCD_3): $\delta = 8.10$ (s, 1H), 7.19 (s, 1H), 6.54 (s, 1H), 4.64 (d, $J=10.0$ Hz, 1H), 4.36 (d, $J=10.3$ Hz, 1H), 3.95 (d, $J=4.5$ Hz, 1H), 3.56 (m, 1H), 3.53 (ddd, $J=11.0, 2.3, 1.1$ Hz, 1H), 2.42 (m, 1H), 2.36 (m, 1H), 2.12 (br d, $J=17.0$ Hz, 1H), 1.96 (m, 1H), 1.88 (m, 1H), 1.84 (s, 3H), 1.71 (m, 1H), 1.67 (m, 1H), 1.64 (m, 1H), 1.61 (s, 3H), 1.57 (m, 1H (10- H^b)^[3a]), 1.45 (ddd, $J=12.9, 11.4, 2.9$ Hz, 1H), 1.33 (m, 2H (11- H_2)), 1.25 (m, 10- H^b), 0.95 (d, $J=7.0$ Hz, 3H), 0.94 (d, $J=6.5$ Hz, 3H), 0.89 (d, $J=6.5$ Hz, 3H), 0.87 (d, $J=6.5$ Hz, 3H), 0.71 (d, $J=7.0$ Hz, 3H); ^{13}C -NMR (150MHz, CD_3COCD_3): $\delta = 182.1, 168.6, 146.8, 141.3, 132.1, 130.2, 129.9, 119.1, 111.0, 104.2, 78.7, 77.8, 76.2, 46.1, 41.4, 40.8, 39.8, 38.1, 35.9, 33.6, 33.5, 26.5, 22.7, 19.9, 19.7, 13.3, 12.7, 7.6, 7.2$; IR (film): 3322, 2926, 1670, 1614, 1585, 1329, 1098 cm^{-1} ; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{29}\text{H}_{42}\text{O}_6$, 486.2981, found, 486.2975.

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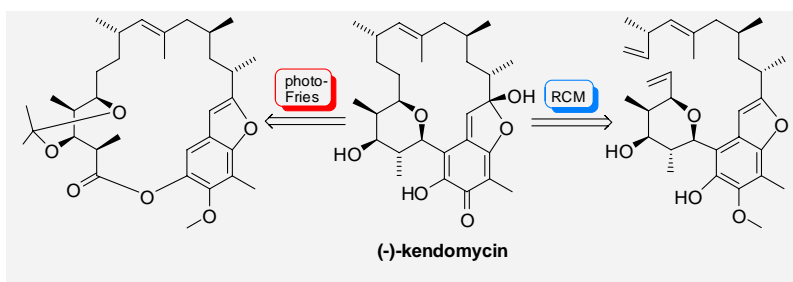
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*Thomas Magauer, Harry J. Martin,
and Johann Mulzer**
Page – Page

**Ring Closing Metathesis and
Photo-Fries Reaction for the
Construction of the Ansamycin
Antibiotic Kendomycin.
Development of a Protecting
Group Free Oxidative Endgame.**



The so far underestimated photo-Fries reaction serves as an efficient tool for the total synthesis of (-)-kendomycin. A powerful RCM is the basis of the second total synthesis.

The installation of the lactol unit was performed by a chemoselective oxidation-hydrolysis sequence, thus avoiding additional protecting groups.

**Ring Closing Metathesis and Photo-Fries Reaction
for the Construction of the Ansamycin Antibiotic
Kendomycin. Development of a Protecting Group
Free Oxidative Endgame.**

SUPPORTING INFORMATION

Thomas Magauer, Harry J. Martin and Johann Mulzer*

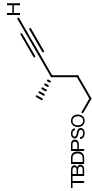
Institute of Organic Chemistry, University of Vienna
Währingerstraße 38
1090 Vienna
Austria

General Methods

All solvents were distilled prior to use, except THF, which was purchased from Acros Organics (99.85%, H₂O < 50 ppm) and used without further purification. Et₂O, toluene and benzene were distilled from sodium. CH₂Cl₂ and CHCl₃ were passed through an Al₂O₃-MgSO₄ column or distilled over P₂O₅. Acetone was distilled over P₂O₅. DMF, DMSO, NEt₃, *i*Pr₃NH, *i*Pr₂NEt, TMEDA, HMPA and 2,6-lutidine were distilled from CaH₂. TBSCl was dissolved in hexane or THF (3 M), treated with Et₃N (3%) and transferred *via* a syringe filter to the reaction mixture. CpZrHCl was prepared according to the Negishi procedure¹. Solvent degassing was achieved by repeated (at least 4 cycles) freeze-pump-thaw (FPT) cycles. All non-aqueous reactions were performed under an atmosphere of argon using oven-dried or flame-dried glassware and standard syringe/septa techniques. ¹H- and ¹³C-NMR spectra were measured in CDCl₃ on a Bruker Avance DRX-400 or DRX-600 at 400.13 MHz (100.61 MHz) or 600.13 MHz (150.90 MHz), respectively. Chemical shifts are given in ppm and were referenced to residual CHCl₃ (¹H, δ = 7.26 ppm, ¹³C, δ = 77.00 ppm) or toluene (¹H, δ = 7.09, 7.00, 6.98 ppm, ¹³C, δ = 137.9, 129.2, 128.3, 125.5, 20.4 ppm). Data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad), coupling constant in Hz, integration. Assignments of proton resonances were confirmed by correlated spectroscopy. IR spectra were recorded as thin films on a silicon plate on a Perkin-Elmer 1600 FT-IR spectrometer. Mass spectra were measured on a Micro mass, trio 200 Fisons Instruments. High-resolution mass spectra (HRMS) were performed with a Finnigan MAT 8230 with a resolution of 10000. Optical rotations were measured on a Perkin-Elmer 351 polarimeter at 20 °C (reported as follows: concentration (c in g/100mL), solvent). The reaction progress was monitored on precoated TLC plates (Merck Kieselgel 60 F254). Spots were visualized under UV light (254nm) and/or were stained with ceric ammonium molybdate (CAM), *p*-anisaldehyde or potassium permanganate stain. Column chromatography was performed with Merck silica gel 60 (230-400 mesh). Analytical HPLC was performed on a Jasco System (PU-980 pump, UV 975 and RI 930) using a Nucleosil 50 column (5 µm, Ø 4 mm x 241 mm) at ambient temperature. Preparative HPLC was performed on a Dynamix Model SD-1 equipped with a Model UV-1 absorbance detector using a Supersphere (60 Å pore size, 4 µm

¹ Huang, Z.; Negishi, E. *Org. Lett.* **2006**, *8*, 3675.

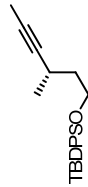
particle size, \varnothing 25 mm x 250 mm) at ambient temperature. Yields refer to chromatographically purified compounds, unless otherwise stated.



(S)-*tert*-Butyl(3-methylpent-4-ynoxy)diphenylsilane (72).

DIPA (1.23 mL, 8.81 mmol, 1.15 equiv.) was dissolved in THF (7 mL), cooled to -78°C and *n*-BuLi (2.5 M in hexane, 3.52 mL, 8.81 mmol, 1.15 equiv.) was added. The resulting solution was stirred for 10 min at 0°C , recooled to -78°C and subsequently treated with TMSCHN₂ (4.4 mL, 8.81 mmol, 1.15 equiv.). The temperature was kept for 0.5 h, after which aldehyde **13** (2.61 g, 7.66 mmol, 1 equiv.) in THF (1 M) was added *via* syringe. Stirring was continued for 1 h at -78°C , and 2 h at room temperature. The reaction was quenched by the addition of NH₄Cl (20 mL) and extracted with Et₂O (3 x 10 mL). The combined organic phases were washed with brine, dried over MgSO₄ and the solvent was removed under reduced pressure. The crude product was purified by column chromatography (hexane : ethylacetate = 10 : 1) affording 2.10 g (82%) of pure **72**.

¹H-NMR (400MHz, CDCl₃): δ 7.70–7.63 (m, 6H), 7.45–7.34 (m, 6H), 3.87–3.73 (m, 2H), 2.79–2.68 (m, 1H), 1.99 (d, *J* = 2.5 Hz, 1H), 1.72–1.66 (m, 2H), 1.19 (d, *J* = 7.1 Hz, 3H), 1.05 (s, 9H); ¹³C-NMR (100MHz, CDCl₃): δ 135.6, 129.5, 127.6, 68.3, 69.5, 39.5, 26.8, 22.2, 20.9, 19.2; HRMS (ESI) (*m/z*): [M+H]⁺ calcd for C₃₈H₃₉OSi, 279.1205, found, 279.1201; IR (film): 2932, 1700, 1653, 1472, 1428, 1112 cm⁻¹; [α]_D²⁰ +4.5 (c 1.11, CH₂Cl₂).

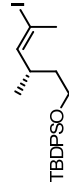


(S)-tert-Butyl(3-methylhex-4-ynloxy)diphenylsilane (73).

A solution of alkyne **72** (2.10 g, 6.23 mmol, 1 equiv.) in THF (20 mL) was cooled to -78°C and treated with *n*-BuLi (2.5 M in hexane, 5.49 mL, 13.72 mmol, 2.2 equiv.). After 20 min MeI (1.01 mL, 16.22 mmol, 2.6 equiv.) was added and the temperature was raised to room temperature over night. The solution was treated with NH_4Cl (20 mL), extracted with Et_2O (3 \times 10 mL), dried

over MgSO_4 , filtered and the solvent was evaporated. Purification by flash chromatography (hexane : ethylacetate = 50 : 1) yielded 2.08 g (95%) **73**.

$^1\text{H-NMR}$ (400MHz, CDCl_3): δ 7.71–7.64 (m, 4H), 7.45–7.34 (m, 6H), 3.86–3.72 (m, 2H), 1.75 (d, J = 2.8 Hz, 3H), 1.72–1.57 (m, 2H), 1.14 (d, J = 6.8 Hz, 3H), 1.05 (s, 9H); $^{13}\text{C-NMR}$ (100MHz, CDCl_3): δ 135.6, 134.1, 134.0, 129.5, 127.6, 127.5, 83.5, 61.9, 40.0, 26.8, 22.4, 21.4, 19.2, 3.5; HRMS (ESI) (m/z): $[\text{M-tBu}]^+$ calcd for $\text{C}_{19}\text{H}_{22}\text{OSi}$, 293.1362, found, 293.1360; IR (film): 3071, 2932, 2363, 1700, 1653, 1428, 1112 cm^{-1} ; $[\alpha]_D^{20}$ +5.9 (c 0.88, CH_2Cl_2).

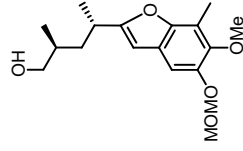


(5E)-tert-Butyl(5-iodo-3-methylhex-4-enyloxy)diphenylsilane (10).

A 250 mL Schlenk flask was charged with Cp_2ZrCl_2 (5.53 g, 18.92 mmol, 2.8 equiv.) and THF (50 mL). The solution was cooled to 0 °C and DIBAL-H (1 M in heptane, 18.93 mL, 18.93 mmol, 2.8 equiv.) was slowly added *via* cannula. The white precipitate was stirred for 1.5 h at room temperature, after which the supernatant was carefully removed. Alkyne **73** (2.37 g, 6.76 mmol, 1 equiv.) in benzene (15 mL) was added and the resulting suspension was heated to 40 °C for 3.5 h. The mixture was cooled to 0 °C and I_2 (4.80 g, 18.92 mmol, 2.8 equiv.) in benzene (40 mL) was slowly transferred to the solution.

The reaction was quenched after 5 min by the addition of 1 M $\text{Na}_2\text{S}_2\text{O}_3$ solution. Stirring was continued until both layers became colorless. The aqueous phase was extracted with Et_2O (4 x 20 mL), the combined organic layers were washed with brine, dried over MgSO_4 , filtered and evaporated to dryness. Subsequent flash chromatography (hexane : ethylacetate = 50 : 1) yielded 2.67 g (83%) of vinyl iodide **10**.

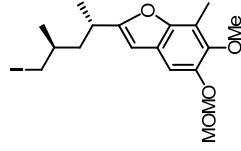
$^1\text{H-NMR}$ (400MHz, CDCl_3): δ 7.71–7.65 (m, 4H), 7.48–7.37 (m, 6H), 5.95 (dq, J = 9.2, 1.2 Hz, 1H), 3.66 (dd, J = 6.8, 5.6 Hz, 2H), 2.79–2.67 (m, 1H), 2.40 (d, J = 1.5 Hz, 3H), 1.63–1.52 (m, 1H), 1.51–1.41 (m, 1H), 1.07 (s, 9H), 0.97 (d, J = 6.8 Hz, 3H); $^{13}\text{C-NMR}$ (100MHz, CDCl_3): δ 146.8, 135.5, 134.0, 129.6, 127.6, 93.1, 61.6, 39.5, 32.0, 27.8, 26.9, 20.3, 19.2; HRMS (ESI) (m/z): $[\text{M-tBu}]^+$ calcd for $\text{C}_{19}\text{H}_{22}\text{IOSi}$, 421.0485, found, 421.0470; IR (film): 3071, 2958, 1634, 1589, 1471, 1428, 1389, 1360 cm^{-1} ; $[\alpha]_D^{20}$ +19.3 (c 0.81, CH_2Cl_2).



(2S,4S)-4-(6-Methoxy-5-(methoxymethoxy)-7-methylbenzofuran-2-yl)-2-methylpentan-1-ol (74).

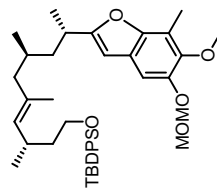
Compound **14** (4.96 g, 8.85 mmol, 1 equiv.) was dissolved in THF (180 mL) and TBAF (1 M in THF, 10.62 mL, 10.62 mmol, 1.2 equiv.) was added at ambient temperature. The mixture was stirred overnight, finally quenched with 200 mL of H_2O and the aqueous layer was extracted with Et_2O (3 x 70 mL). The combined organic phases were dried over MgSO_4 , filtered and the solvent was evaporated. Purification by column chromatography (hexane : ethylacetate from 5 : 1 to 3 : 1) furnished 2.68 g (94%) of **74**.

$^1\text{H-NMR}$ (400MHz, CDCl_3): δ 7.07 (s, 1H), 6.26 (s, 1H), 5.20 (s, 2H), 3.83 (s, 3H), 3.54 (s, 3H), 3.56–3.47 (m, 2H), 3.03 (m, 1H), 2.41 (s, 3H), 1.79–1.69 (m, 1H), 1.67–1.59 (m, 2H), 1.30 (d, J = 6.8 Hz, 3H), 0.94 (d, J = 6.5 Hz, 3H); $^{13}\text{C-NMR}$ (100MHz, CDCl_3): δ 164.2, 123.4, 115.3, 105.0, 100.7, 96.2, 68.2, 61.0, 56.1, 39.0, 33.4, 31.2, 19.3, 16.7, 9.1; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{18}\text{H}_{26}\text{O}_5$, 322.1780, found, 322.1786; IR (film): 3745, 3676, 2932, 1684, 1653, 1560, 1457, 1043 cm^{-1} ; $[\alpha]_D^{20}$ +0.9 (c 0.78, CH_2Cl_2).



2-((2*S*,4*S*)-5-iodo-4-methylpentan-2-yl)-6-methoxy-5-(methoxymethoxy)-7-methylbenzofuran (11).

To a solution of alcohol **74** (700 mg, 2.17 mmol, 1 equiv.) in CH₂Cl₂ (14 mL) was added imidazole (192 mg, 2.82 mmol, 1.3 equiv.) and triphenylphosphine (740 mg, 2.82 mmol, 1.3 equiv.) at 0 °C. After 5 min iodine (716 mg, 2.82 mmol, 1.3 equiv.) was added in three portions. Stirring was continued for 1 h at room temperature. The reaction was diluted with CH₂Cl₂, quenched with NH₄Cl (60 mL) and extracted with CH₂Cl₂ (3 x 20 mL). The organic layer was washed with brine, dried over MgSO₄, filtered and the solvent was evaporated. The crude residue was purified by flash chromatography (hexane : ethylacetate = 15 : 1) to obtain 822 mg (88%) of pure iodine **11**. ¹H-NMR (400MHz, CDCl₃): δ 7.08 (s, 1H), 6.28 (s, 1H), 5.21 (s, 2H), 3.84 (s, 3H), 3.54 (s, 3H), 3.27 (d, *J* = 4.8 Hz, 2H), 2.97 (m, 1H), 2.43 (s, 3H), 1.73–1.56 (m, 2H), 1.50–1.40 (m, 1H), 1.32 (d, *J* = 6.9 Hz, 3H), 0.97 (d, *J* = 6.6 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 163.3, 149.2, 146.9, 145.7, 123.3, 115.3, 105.0, 100.9, 96.1, 60.9, 56.1, 42.5, 31.9, 31.2, 27.9, 19.6, 17.9, 9.1; HRMS (EI, 70eV, 70 °C) [M]⁺ calcd for C₁₈H₂₃O₄, 432.0798, found, 432.0802; IR (film): 2962, 1605, 1455, 1422, 1338, 1311, 1260, 1218, 1152 cm⁻¹; [α]_D²⁰ +8.4 (c 0.65, CH₂Cl₂).

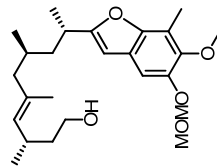


tert-Butyl((3*S*,7*S*,9*S*)-9-(6-methoxy-5-(methoxymethoxy)-7-methylbenzofuran-2-yl)-3,5,7-trimethyldec-4-en-1-yl)dimethylsilane (75).

To flame-dried ZnCl₂ (126 mg, 0.925 mmol, 1.5 equiv.) was added alkyl iodide **11** (400 mg, 0.925 mmol, 1.5 equiv.) in degassed Et₂O (5 mL). The suspension was cooled to -78 °C and *t*-BuLi (1.7 M in pentane, 1.63 mL, 2.776 mmol, 4.5 equiv.) was rapidly added. The pale yellow solution was stirred for 5 min at -78 °C and 1.5 h at 0 °C. Then Pd(PPh₃)₄ (36 mg, 0.033 mmol, 0.05 equiv.) and vinyl iodide **10** (295 mg, 0.617 mmol, 1 equiv.) in degassed THF (3 mL) were added at 0 °C. The yellow mixture was stirred for 2.5 h at 0 °C and allowed to warm to room temperature

overnight. The reaction was diluted with Et₂O (10 mL) and quenched by the addition of NH₄Cl (20 mL). The phases were separated, extracted with Et₂O (4 x 10 mL), washed with brine, dried over MgSO₄, filtered and evaporated to dryness. The crude product (405 mg, 100%) was directly used in the next step. A small amount was purified by HPLC (hexane : ethylacetate = 20 : 1) to furnish pure **75**.

¹H-NMR (400MHz, CDCl₃): δ 7.69–7.62 (m, 4H), 7.43–7.31 (m, 6H), 7.08 (s, 1H), 6.24 (s, 1H), 5.21 (s, 2H), 4.84 (d, *J* = 9.3 Hz, 1H), 3.84 (s, 3H), 3.66–3.60 (m, 2H), 3.55 (s, 3H), 3.05–2.94 (m, 1H), 2.65–2.53 (m, 1H), 2.43 (s, 3H), 2.04 (dd, *J* = 12.5, 5.1 Hz, 1H), 1.72 (dd, *J* = 11.5, 8.5 Hz, 1H), 1.68–1.56 (m, 2H), 1.54 (s, 3H), 1.53–1.38 (m, 3H), 1.27 (d, *J* = 6.9 Hz, 3H), 1.04 (s, 9H), 0.92 (d, *J* = 6.6 Hz, 3H), 0.75 (d, *J* = 6.1 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 164.8, 149.2, 146.9, 145.5, 135.6, 135.5, 134.2, 137.7, 1322.6, 129.4, 127.5, 123.5, 115.3, 105.0, 100.4, 96.2, 62.3, 61.0, 56.1, 48.0, 43.0, 40.5, 31.3, 28.8, 28.2, 26.9, 21.3, 19.5, 19.2, 16.0, 9.11; HRMS (ESI) [m/z]: [M]⁺ calcd for C₄₁H₅₆O₅Si, 656.3897, found, 656.3877; IR (film): 2928, 1589, 1455, 1428, 1339, 1218, 1153, 1112, 1044 cm⁻¹; [α]_D²⁰ +3.3 (c 1.32, CH₂Cl₂).

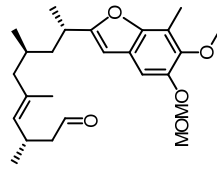


(3*S*,7*S*,9*S*)-9-(6-methoxy-5-(methoxymethoxy)-7-methylbenzofuran-2-yl)-3,5,7-trimethyldec-4-en-1-ol (15).

Crude product **75** (405 mg, 0.616 mmol, 1 equiv.) was dissolved in THF (20 mL) and TBAF was added at room temperature. The solution was stirred for 24 h and quenched by the addition of H₂O (20 mL). The layers were separated, extracted with Et₂O (3 x 15 mL), dried over MgSO₄ and concentrated. Purification by flash column chromatography (hexane : ethylacetate = 3 : 1) afforded 173 mg (67% over 2 steps) of primary alcohol **15**.

¹H-NMR (400MHz, CDCl₃): δ 7.08 (s, 1H), 6.25 (s, 1H), 5.21 (s, 2H), 4.91 (d, *J* = 9.3 Hz, 1H), 3.84 (s, 3H), 3.67–3.56 (m, 2H), 3.54 (s, 3H), 3.06–2.95 (m, 2H), 2.59–2.46 (m, 1H), 2.42 (s, 3H), 2.07

(dd, $J = 12.6, 5.3$ Hz, 1H), 1.77 (dd, $J = 12.6, 8.3$ Hz, 1H), 1.73–1.59 (m, 2H), 1.58 (d, $J = 1.3$ Hz, 3H), 1.57–1.41 (m, 3H), 1.27 (d, $J = 6.8$ Hz, 3H), 0.96 (d, $J = 6.8$ Hz, 3H), 0.83 (d, $J = 6.3$ Hz, 3H); $^{13}\text{C-NMR}$ (100MHz, CDCl_3): δ 164.7, 149.1, 146.9, 145.5, 133.0, 132.4, 123.5, 115.2, 105.0, 100.4, 96.2, 61.6, 61.0, 56.1, 48.0, 43.0, 40.5, 31.3, 29.3, 28.2, 21.4, 19.5, 19.1, 16.0, 9.1; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{25}\text{H}_{38}\text{O}_5$, 418.2719, found, 418.2726; IR (film): 3442, 2926, 1700, 1653, 1604, 1452, 1340, 1219, 1153, 1117, 1090, 1045 cm^{-1} ; $[\alpha]_D^{20} +14.5$ (c 1.80, CH_2Cl_2).



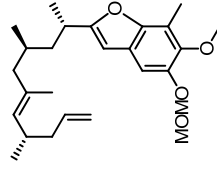
(3S,7S,9S,E)-9-(6-Methoxy-5-(methoxymethoxy)-7-methylbenzofuran-2-yl)-3,5,7-trimethyldec-4-enal (76).

A 50 mL flask was charged with alcohol **15** (560 mg, 1.34 mmol, 1 equiv.) in DMSO (7 mL). After the addition of IBX (1.39 g, 4.96 mmol, 3.6 equiv.) the solution was stirred for 30 min at ambient temperature.

The solution was diluted with Et_2O : hexane = 1 : 1 (15 mL) and 15 mL of H_2O to precipitate remaining IBX. The mixture was filtered over Celite, the phases were separated and the aqueous layer was extracted with Et_2O : hexane = 1 : 1 (3 x 10 mL). The combined organic layers were dried over anhydrous MgSO_4 , filtered over a plug of silica and evaporated to dryness. Crude aldehyde **76** (541 mg, 97%) was used in the next step without further purification.

$^1\text{H-NMR}$ (400MHz, CDCl_3): δ 9.69 (t, $J = 2.4$ Hz, 1H), 7.08 (s, 1H), 6.24 (s, 1H), 5.21 (s, 2H), 4.95 (d, $J = 9.7$ Hz, 1H), 3.84 (s, 3H), 3.54 (s, 3H), 3.04–2.91 (m, 2H), 2.42 (s, 3H), 2.37–2.32 (m, 2H), 2.06 (dd, $J = 12.5, 5.4$ Hz, 1H), 1.75 (dd, $J = 12.5, 8.5$ Hz, 1H), 1.71–1.62 (m, 1H), 1.60 (d, $J = 1.3$ Hz, 3H), 1.59–1.52 (m, 1H), 1.50–1.42 (m, 1H), 1.27 (d, $J = 7.1$ Hz, 3H), 1.03 (d, $J = 6.8$ Hz, 3H), 0.79 (d, $J = 6.31$ Hz, 3H); $^{13}\text{C-NMR}$ (100MHz, CDCl_3): δ 202.7, 164.6, 149.2, 146.9, 133.9, 130.6, 123.5, 115.3, 105.0, 100.4, 96.2, 61.0, 56.1, 51.2, 47.8, 43.0, 31.3, 28.2, 27.9, 21.2, 19.5, 19.2,

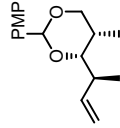
16.2, 9.1; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{25}\text{H}_{38}\text{O}_5$, 416.2563, found, 416.2556; IR (film): 2959, 1725, 1653, 1605, 1455, 1419, 1379, 1219, 1153 cm^{-1} ; $[\alpha]_D^{20} +36.9$ (c 1.05, CH_2Cl_2).



6-Methoxy-5-(methoxymethoxy)-7-methyl-2-((2S,4S,8S,E)-4,6,8-trimethylundeca-6,10-dien-2-yl)benzofuran (16).

MePPH_3Br (189 mg, 0.52 mmol, 2 equiv.) was suspended in THF (2 mL) and placed on an ice bath. Subsequently $t\text{-BuOK}$ (59 mg, 0.52 mmol, 2 equiv.) was added and the yellow suspension was stirred for 1.5 h at room temperature. The orange solution was cooled to 0 °C and aldehyde **76** (110 mg, 0.26 mmol, 1 equiv.) in THF (0.8 mL) was added dropwise. After 1 h the reaction was quenched by the addition of saturated aqueous NH_4Cl (20 mL) and the aqueous phase was extracted with Et_2O : hexane = 1 : 1 (3 x 15 mL). The combined organic layers were dried over MgSO_4 , filtered and concentrated under reduced pressure. Purification by flash chromatography (hexane : ethylacetate = 3 : 1) gave 99 mg (90%) of diene **16**.

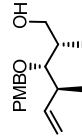
$^1\text{H-NMR}$ (400MHz, CDCl_3): δ 7.09 (s, 1H), 6.25 (s, 1H), 5.77 (ddt, $J = 17.1, 10.2, 7.0$ Hz, 1H), 5.22 (s, 3H), 5.03–4.91 (m, 2H), 3.85 (s, 3H), 3.55 (s, 3H), 3.07–2.96 (m, 1H), 2.50–2.40 (m, 1H), 2.43 (s, 3H), 2.08 (dd, $J = 12.6, 5.3$ Hz, 1H), 2.05–1.99 (m, 2H), 1.77 (dd, $J = 12.6, 8.3$ Hz, 1H), 1.74–1.64 (m, 1H), 1.64–1.44 (m, 2H), 1.57 (d, $J = 1.3$ Hz, 3H), 1.28 (d, $J = 7.1$ Hz, 3H), 0.95 (d, $J = 6.6$ Hz, 3H), 0.84 (d, $J = 6.3$ Hz, 3H); $^{13}\text{C-NMR}$ (100MHz, CDCl_3): δ 164.8, 149.2, 146.9, 145.5, 137.6, 132.5, 132.3, 123.5, 115.3, 105.0, 100.4, 96.2, 61.0, 56.1, 48.0, 43.0, 42.0, 32.5, 31.3, 28.2, 20.8, 19.5, 19.2, 16.2, 9.1; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{28}\text{H}_{38}\text{O}_4$, 414.2770, found, 414.2761; IR (film): 2958, 1700, 1653, 1606, 1452, 1379, 1219, 1153, 1044 cm^{-1} ; $[\alpha]_D^{20} +16.8$ (c 2.50, CH_2Cl_2).

**(4S,5S)-4-((S)-But-3-en-2-yl)-2-(4-methoxyphenyl)-5-methyl-1,3-dioxane (77).**

Alcohol **17** (950 mg, 3.59 mmol, 1 equiv.) and 1 g of powdered molecular sieve 3Å in CH₂Cl₂ (32 mL) was cooled to 0 °C and DDQ (978 mg, 4.3 mmol, 1.2 equiv.) was added in three portions over a period of 5 min. The resulting green-black solution was stirred for 1.5 h, allowing the reaction to warm to ambient temperature.

After the addition of Et₂O (80 mL), the reaction was quenched with saturated Na₂S₂O₃ (30 mL) and NaHCO₃ (30 mL). The aqueous layer was extracted with Et₂O (3 x 20 mL), washed with brine and dried over MgSO₄, filtrated and evaporated to dryness. Purification by flash chromatography (hexane : ethylacetate = 10 : 1) afforded 700 mg (74%, 85% BORSM) **77** as a colorless oil.

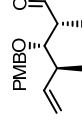
¹H-NMR (400MHz, CDCl₃): δ 7.41 (d, *J* = 8.7 Hz, 2H), 6.87 (d, *J* = 8.7 Hz, 2H), 5.98 (ddd, *J* = 17.2, 10.5, 6.6 Hz, 1H), 5.43 (s, 1H), 5.10–5.00 (m, 2H), 4.04 (dd, *J* = 4.0, 3.0 Hz, 2H), 3.79 (s, 3H), 3.56 (dd, *J* = 9.9, 2.2 Hz, 1H), 2.44–2.34 (m, 1H), 1.72–1.64 (m, 1H), 1.19 (d, *J* = 7.1, 3H), 0.97 (d, *J* = 7.1, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 159.7, 141.6, 131.6, 130.8, 127.2, 113.8, 113.5, 101.4, 83.6, 55.3, 45.1, 38.5, 30.1, 17.2, 14.3, 10.9; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₁₆H₂₂O₃, 262.1569, found, 262.1557; IR (film): 3077, 2966, 2838, 1677, 1615, 1517, 1463, 1395, 1302, 1248 cm⁻¹; [α]_D²⁰ -30.5 (c 1.98, CH₂Cl₂).

**(2S,3S,4S)-3-(4-Methoxybenzyloxy)-2,4-dimethylhex-5-en-1-ol (78).**

Acetal **77** (690 mg, 2.63 mmol, 1 equiv.) in CH₂Cl₂ (7 mL) was cooled to -78 °C. DIBAL-H (1.5 M in toluene 5.3 mL, 8.00 mmol, 3 equiv.) was added *via* a dropping funnel over 15 min. The reaction mixture was warmed to -10 °C within 5 h, diluted with CH₂Cl₂ (7 mL) and finally quenched with Na-K-tartrate (15 mL). The resulting suspension was stirred until precise phase separation was achieved (2 h). The mixture was extracted with CH₂Cl₂ (3 x 5 mL), washed with brine (15 mL),

dried over MgSO₄ and concentrated. Flash chromatography (hexane : ethylacetate = 3 : 1) of the residual oil yielded 650 mg (93%) of **78**.

¹H-NMR (400MHz, CDCl₃): δ 7.27 (d, *J* = 8.1 Hz, 2H), 6.87 (d, *J* = 8.1 Hz, 2H), 5.96 (ddd, *J* = 18.1, 9.3, 8.2, 1H), 5.13–5.01 (m, 2H), 4.58 (d, *J* = 10.7, 1H), 4.44 (d, *J* = 10.7, 1H), 3.79 (s, 3H), 3.64–3.51 (m, 2H), 3.38 (dd, *J* = 3.8 Hz, 1H), 2.52 (m, 1H), 1.99–1.89 (m, 1H), 1.75 (t, *J* = 5.1 Hz, 1H), 1.03 (d, *J* = 6.8 Hz, 3H), 0.94 (d, *J* = 7.1 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 159.2, 141.8, 130.9, 129.4, 114.5, 113.8, 83.9, 73.7, 66.2, 55.3, 40.8, 37.6, 17.4, 11.2; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₁₆H₂₄O₃, 264.1725, found, 264.1719; IR (film): 3417, 3074, 2964, 1613, 1586, 1514, 1462, 1301, 1248, 1173 cm⁻¹; [α]_D²⁰ +3.6 (c 1.96, CH₂Cl₂).

**(2R,3S,4S)-3-(4-Methoxybenzyloxy)-2,4-dimethylhex-5-enal (12).****Method A:**

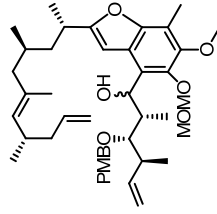
To alcohol **78** (112 mg, 0.42 mmol, 1 equiv.) in CH₂Cl₂ (2 mL) NaHCO₃ (125 mg, 1.48 mmol, 3.5 equiv.) and Dess-Martin periodinate (467 mg, 1.10 mmol, 2.6 equiv.) were added. The reaction mixture was stirred for 0.5 h at 0 °C, followed by 2 h at room temperature. Then a mixture of saturated aqueous Na₂S₂O₃ (5 mL) and saturated NaHCO₃ (5 mL) was added and stirred until the solution was clear. The mixture was extracted with Et₂O (3 x 10 mL), dried over MgSO₄, filtered and evaporated to dryness, obtaining 98 mg (89%) of a colorless oil, which was directly used in the next step.

Method B:

A 25 ml Schlenk flask was charged with oxalychloride (295 μL, 3.48 mmol, 2 equiv.) and CH₂Cl₂ (3.4 mL) and the resulting solution was cooled to -78 °C. Then DMSO (494 μL, 6.96 mmol, 4 equiv.) was added dropwise (gas outlet!). After the reaction was stirred for 1 h, alcohol **78** (0.3 M in CH₂Cl₂, 5.8 mL, 1.74 mmol, 1 equiv.) was slowly added and stirring was continued for additional 45 min. After the addition of DIPEA (1.45 mL, 10.44 mmol, 6 equiv.) a white slurry was formed and the reaction was warmed to 0 °C, hydrolyzed with saturated aqueous NH₄Cl (15 mL), extracted with CH₂Cl₂ (3 x 20), washed with brine, dried over MgSO₄ and filtered. The

solvent was carefully removed under reduced pressure and the residue was rapidly filtered over a small pad of silica to give 455 mg (99%) of **12**.

¹H-NMR (600MHz, CDCl₃): δ 9.62 (d, *J* = 1.4 Hz, 1H), 7.13 (d, *J* = 8.5 Hz, 2 H), 6.76 (d, *J* = 8.8 Hz, 2H), 5.76 (ddd, *J* = 17.1, 10.4, 8.2 Hz, 1H), 4.99–4.95 (m, 2H), 4.38 (d, *J* = 10.8 Hz, 1H), 4.33 (d, *J* = 10.8 Hz, 1H), 3.70 (s, 3H), 3.61 (dd, *J* = 5.7, 4.5 Hz, 1H), 2.49 (ddq, *J* = 7.2, 2.3, 1.1 Hz, 1H), 2.43–2.37 (m, 1H), 1.07 (d, *J* = 7.2 Hz, 3H), 0.96 (d, *J* = 6.8 Hz, 3H); ¹³C-NMR (150MHz, CDCl₃): δ 204.4, 159.2, 140.3, 130.3, 129.3, 115.7, 113.7, 81.6, 73.3, 55.2, 49.2, 41.1, 17.1, 8.9; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₁₆H₂₂O₃, 262.1569, found, 262.1560; IR (film): 2971, 1722, 1613, 1586, 1514, 1462, 1345, 1302, 1248, 1174 cm⁻¹; [α]_D²⁰ -20.9 (c 2.31, CH₂Cl₂).



(1*S*,2*S*,3*S*,4*S*)-1-(6-methoxy-5-(methoxymethoxy)-7-methyl-2-((2*S*,4*S*,8*S*)-4,6,8-

trimethylundeca-6,10-dien-2-yl)benzofuran-4-yl)-3-(4-methoxybenzyloxy)-2,4-dimethylhex-5-

en-1-ol (**18a**) and (1*R*,2*S*,3*S*,4*S*)-1-(6-methoxy-5-(methoxymethoxy)-7-methyl-2-((2*S*,4*S*,8*S*)-

4,6,8-trimethylundeca-6,10-dien-2-yl)benzofuran-4-yl)-3-(4-methoxybenzyloxy)-2,4-

dimethylhex-5-en-1-ol (**18b**).

Method A:

Diene **16** (170 mg, 0.410 mmol, 1 equiv.) was dissolved in THF (1.5 mL, 0.3 M). After the addition of TMEDA (148 μL, 0.98 mmol, 2.4 equiv.) the solution was cooled to -40 °C and *n*-BuLi (2.5 M in hexane, 197 μL, 0.49 mmol, 1.2 equiv.) was slowly added *via* cannula. The temperature was raised to -30 °C and stirring was continued at this temperature for 1 h. The reaction mixture was cooled to -78 °C and aldehyde **12** in THF (0.5 mL) was added *via* cannula. Stirring was continued for 2 h allowing the reaction to warm to -25 °C.

The reaction was taken up in Et₂O and quenched by the addition of saturated NH₄Cl. Extraction was done by Et₂O (3 x 10 mL). The combined organic layers were dried over MgSO₄, filtered and evaporated under reduced pressure. Purification by HPLC (hexane : ethylacetate = 20 : 1) afforded 127 mg of pure **18a** and 81 mg of pure **18b**, (208 mg, 75%, **18a** : **18b** = 1.5 : 1).

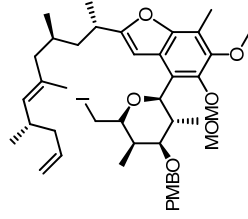
Method B:

NaBH₄ (125 mg, 3.33 mmol, 15 equiv.) was added to dry MeOH (2 mL) at 0 °C. Ketone **21** (150 mg, 0.22 mmol, 1 equiv.) in MeOH (0.5 mL) was added *via* syringe. The reaction was allowed to warm to ambient temperature and stirred for 6 h. TLC analysis showed complete consumption of the starting material and the reaction was taken up in Et₂O (40 mL). HCl (0.5 N, 10 mL) was added and stirring was continued for 10 min. The mixture was neutralized by adding saturated NaHCO₃ and the phases were separated. The aqueous layer was extracted with Et₂O (3 x 10 mL), the organic phases were dried over MgSO₄ and concentrated. Purification by column chromatography (hexane : ethylacetate = 20 : 1) afforded 92 mg (61%, **18a** : **18b** = 1 : 3).

18b: ¹H-NMR (400MHz, CDCl₃): δ 7.38 (d, *J* = 8.7 Hz, 2H), 6.89 (d, *J* = 8.7 Hz, 2H), 6.34 (br, 1H), 6.04 (ddd, *J* = 17.5, 10.0, 7.6 Hz, 1H), 5.75 (ddt, *J* = 17.1, 10.1, 7.0 Hz, 1H), 5.18–4.84 (m, 8H), 4.73 (d, *J* = 11.0 Hz, 1H), 4.67 (d, *J* = 11.0 Hz, 1H), 3.91 (dd, *J* = 8.5, 1.6 Hz, 1H), 3.80 (s, 3H), 3.79 (s, 3H), 3.52 (s, 3H), 3.04–2.93 (m, 1H), 2.60–2.50 (m, 1H), 2.50–2.49 (m, 1H), 2.39 (s, 3H), 2.07 (dd, *J* = 12.5, 4.8 Hz, 1H), 2.03–1.97 (m, 2H), 1.72 (dd, *J* = 12.5, 8.7 Hz, 1H), 1.69–1.61 (m, 1H), 1.61–1.42 (m, 4H), 1.53 (d, *J* = 1.2 Hz, 3H), 1.26 (d, *J* = 6.8 Hz, 3H), 1.05 (d, *J* = 6.8 Hz, 3H), 0.93 (d, *J* = 6.6 Hz, 3H), 0.82 (d, *J* = 6.1 Hz, 3H), 0.64 (d, *J* = 6.8 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 164.3, 159.0, 147.5, 144.5, 142.7, 137.6, 132.5, 132.3, 131.7, 129.2, 115.3, 114.4, 114.0, 113.7, 100.1, 100.0, 82.5, 74.1, 60.7, 57.6, 55.2, 47.9, 43.1, 42.0, 41.3, 40.9, 32.5, 31.3, 28.2, 20.8, 19.3, 19.0, 17.1, 16.1, 10.3, 9.1; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₄₂H₆₀O₇, 676.4339, found, 676.4341; IR (film): 3490, 2959, 1640, 1613, 1455, 1393, 1301, 1247, 1113 cm⁻¹; [α]_D²⁰ +5.7 (c 1.62, CH₂Cl₂).

18a: ¹H-NMR (400MHz, CDCl₃): δ 7.29 (d, *J* = 8.6 Hz, 2H), 6.87 (d, *J* = 8.8 Hz, 2H), 6.56 (s, 1H), 5.85 (ddd, *J* = 18.2, 9.3, 8.2 Hz, 1H), 5.75 (ddt, *J* = 17.2, 10.1, 7.1 Hz, 1H), 5.27 (dd, *J* = 6.3, 4.6 Hz, 1H), 5.10 (s, 2H), 5.10–4.89 (m, 5H), 4.51 (d, *J* = 10.5 Hz, 1H), 4.28 (d, *J* = 10.5 Hz, 1H), 3.80 (s, 3H), 3.78 (s, 3H), 3.56 (s, 3H), 3.42 (d, *J* = 4.29 Hz, 1H), 3.22 (dd, *J* = 7.7, 2.7 Hz, 1H), 3.01–2.90

(m, 1H), 2.59–2.48 (m, 1H), 2.48–2.37 (m, 2H), 2.40 (s, 3H), 2.07–1.97 (m, 3H), 1.74 (dd, $J = 12.7$, 8.3 Hz, 1H), 1.69–1.59 (m, 1H), 1.55 (s, 3H), 1.54–1.39 (m, 2H), 1.23 (d, $J = 6.8$ Hz, 3H), 1.13 (d, $J = 7.1$ Hz, 3H), 0.97 (d, $J = 6.8$ Hz, 3H), 0.93 (d, $J = 6.8$ Hz, 3H), 0.79 (d, $J = 6.3$ Hz, 3H); ^{13}C -NMR (100MHz, CDCl_3): δ 164.2, 159.0, 150.2, 147.5, 144.0, 142.0, 137.6, 132.5, 132.3, 131.1, 129.2, 125.5, 122.6, 115.3, 114.3, 113.7, 101.1, 99.9, 85.4, 73.3, 60.7, 57.6, 55.3, 48.0, 42.9, 42.0, 41.4, 41.1, 32.5, 31.2, 28.2, 20.8, 19.3, 18.9, 16.8, 26.1, 9.3, 9.1; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{42}\text{H}_{60}\text{O}_7$, 676.4339, found, 676.4344; IR (film): 3490, 2959, 1640, 1613, 1514, 1455, 1393, 1301, 1247, 1113 cm^{-1} ; $[\alpha]_D^{20}$ +41.3 (c 1.27, CH_2Cl_2).

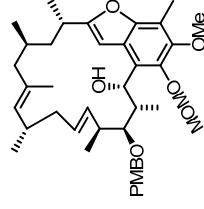


4-((2*R*,3*R*,4*R*,5*R*,6*R*)-(4-methoxybenzyloxy)-3,5-dimethyltetrahydro-2H-pyran-2-yl)-6-methoxy-5-(methoxymethoxy)-2-((3*S*,7*S*,*E*)-3,5,7-trimethyldeca-5,9-dienyl)benzofuran (19).

A solution of **18b** (5 mg, 0.007 mmol, 1 equiv.) in CH_2Cl_2 (1 mL) was cooled to -78°C . 2,6-Di-*tert*-butyl-4-methylpyridine (2 mg, 0.012 mmol, 1.6 equiv.) was added, followed by I_2 (3 mg, 0.011 mmol, 1.5 equiv.). The temperature was raised to -10°C over 4 h and the reaction mixture was diluted with Et_2O (30 mL) and hydrolyzed with NaHCO_3 / $\text{Na}_2\text{S}_2\text{O}_3$ (10 mL). The phases were separated and the aqueous layer was extracted with Et_2O (3 x 10 mL). The organic phase was dried over MgSO_4 , filtered and concentrated under reduced pressure. Flash chromatography (hexane : ethylacetate = 5 : 1) gave 3 mg (50%) of **19**.

^1H -NMR (600MHz, CDCl_3): δ 7.31 (d, $J = 8.6$ Hz, 2H), 6.90 (d, $J = 8.6$ Hz, 2H), 0.89 (d, $J = 7.2$ Hz, 3H), 0.82 (d, $J = 6.4$ Hz, 3H), 0.72 (d, $J = 6.4$ Hz, 3H); ^{13}C -NMR (150MHz, CDCl_3): δ 164.2, 159.3, 150.6, 147.3, 144.2, 137.6, 132.5, 132.3, 130.3, 129.6, 121.7, 121.6, 115.3, 113.8, 100.6, 83.5, 79.5, 78.8, 70.0, 60.8, 57.9, 55.3, 48.1, 42.9, 42.7, 42.0, 35.4, 33.4, 32.5, 31.6, 31.1, 28.1, 22.6,

20.8, 19.4, 18.8, 16.2, 14.1, 13.1, 9.1, 5.4, 5.2; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{43}\text{H}_{59}\text{O}_7$, 802.3305, found, 802.3301; IR (film): 2924, 1640, 1613, 1513, 1455, 1372, 1247, 1112 cm^{-1} ; $[\alpha]_D^{20}$ +16.7 (c 0.30, CH_2Cl_2).



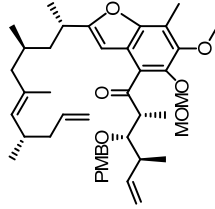
Macrocycle 20a.

Compound **18a** (57 mg, 0.083 mmol, 1 equiv.) was dissolved in degassed CH_2Cl_2 (30 mL, 3 mM) in a 250 mL Schlenk flask. The solution was heated to reflux (45°C) and Grubbs' II catalyst (11 mg, 0.012 mmol, 0.15 equiv.) was added in degassed CH_2Cl_2 (14 mL, 1 mM) over 16 h via syringe pump.

After completion of addition the mixture was stirred for another 2 h. The temperature was lowered to room temperature and air was bubbled through the solution in order to destroy excess catalyst. The solvent was evaporated and purification by column chromatography (hexane : ethylacetate = 10 : 1) afforded 25 mg (46%) of macrocycle **20a**.

^1H -NMR (600MHz, CDCl_3): δ 7.28 (d, $J = 8.6$ Hz, 2H), 6.85 (d, $J = 8.6$ Hz, 2H), 6.43 (s, 1H), 5.45 (dd, $J = 15.5$, 8.1 Hz, 1H), 5.34–5.27 (m, 1H), 5.19–5.15 (br, 1H), 5.15 (s, 2H), 4.90 (d, $J = 9.5$ Hz, 1H), 4.45 (d, $J = 10.8$ Hz, 1H), 4.33 (d, $J = 10.8$ Hz, 1H), 3.80 (s, 3H), 3.76 (s, 3H), 3.60 (s, 3H), 3.54–3.49 (br, 1H), 3.29 (dd, $J = 4.72$ Hz, 1H), 3.05–2.96 (m, 1H), 2.55–2.46 (br, 1H), 2.41 (s, 3H), 2.31–2.42 (m, 1H), 2.16–2.10 (m, 1H), 2.09–2.03 (m, 1H), 1.98 (dd, $J = 14.7$, 4.3 Hz, 1H), 1.88 (dd, $J = 14.7$, 6.3 Hz, 1H), 1.86–1.81 (m, 1H), 1.73–1.63 (m, 2H), 1.62 (s, 3H), 1.58–1.49 (m, 1H), 1.28 (d, $J = 6.9$ Hz, 3H), 1.15 (d, $J = 6.9$ Hz, 3H), 0.95 (d, $J = 6.8$ Hz, 3H), 0.88 (d, $J = 6.5$ Hz, 3H), 0.87 (d, $J = 6.4$ Hz, 3H); ^{13}C -NMR (150MHz, CDCl_3): δ 163.8, 158.9, 150.2, 147.5, 144.8, 133.8, 132.2, 131.5, 130.9, 129.3, 127.8, 124.6, 121.6, 114.7, 113.6, 101.2, 100.3, 83.6, 74.4, 73.0, 60.7, 57.8, 55.2, 45.8, 42.5, 42.0, 41.3, 40.4, 32.0, 31.6, 29.4, 20.8, 20.3, 19.5, 19.1, 19.0, 10.9,

9.1; HRMS (ESI) (m/z): [M]⁺ calcd for C₄₀H₅₆O₇, 648.4026, found, 648.4051; IR (film): 3676, 2926, 1700, 1653, 1612, 1514, 1456, 1301, 1248, 1160, 1113 cm⁻¹; [α]_D²⁰ +0.6 (c 0.85, CH₂Cl₂).



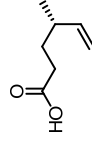
(2R,3S,4S)-1-(6-Methoxy-5-(methoxymethoxy)-7-methyl-2-((2S,4S,8S,E)-4,6,8-trimethylundeca-6,10-dien-2-yl)benzofuran-4-yl)-3-(4-methoxybenzoyloxy)-2,4-dimethylhex-5-en-1-one (21).

A mixture of alcohols **18a** and **18b** (160 mg, 0.23 mmol, 1 equiv.) was dissolved in DMSO (1 mL), IBX (193 mg, 0.69 mmol, 2.9 equiv.) was slowly added and the resulting clear solution was stirred for 50 min.

Dilution with Et₂O : hexane = 1 : 1 (50 mL) and H₂O (15 mL) caused precipitation of remaining IBX, which was removed by filtration over a small plug of Celite. The aqueous phase was extracted with Et₂O (3 x 10 mL), the combined organic phases were dried over MgSO₄ and concentrated *in vacuo*. Purification by column chromatography (hexane : ethylacetate from 20 : 1 to 10 : 1) afforded 152 mg (96%) of pure ketone **21**.

¹H-NMR (400MHz, CDCl₃): δ 7.18 (d, *J* = 8.6 Hz, 2H), 6.82 (d, *J* = 8.6 Hz, 2H), 6.52 (s, 1H), 5.82 (ddd, *J* = 17.6, 10.1, 6.9 Hz, 1H), 5.77–5.68 (m, 1H), 5.13 (d, *J* = 5.3 Hz, 1H), 5.06 (d, *J* = 5.3 Hz, 1H), 5.02–4.84 (m, 4H), 4.76 (d, *J* = 17.2 Hz, 1H), 4.32 (d, *J* = 10.6 Hz, 1H), 4.28 (d, *J* = 10.6 Hz, 1H), 3.96–3.88 (m, 1H), 3.84 (s, 3H), 3.78 (s, 3H), 3.63 (dd, *J* = 5.5, 5.0 Hz, 1H), 3.41 (s, 3H), 3.02–2.91 (m, 1H), 2.45 (s, 3H), 2.44–2.33 (m, 2H), 2.05–1.96 (m, 3H), 1.72 (dd, *J* = 12.8, 8.5 Hz, 1H), 1.68–1.59 (m, 1H), 1.53 (s, 3H), 1.51–1.44 (m, 2H), 1.31 (d, *J* = 6.8 Hz, 3H), 1.23 (d, *J* = 6.8 Hz, 3H), 1.07 (d, *J* = 7.1 Hz, 3H), 0.92 (d, *J* = 6.8 Hz, 3H), 0.77 (d, *J* = 6.3 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 205.9, 166.6, 158.9, 150.1, 147.7, 144.9, 141.2, 137.6, 132.5, 132.2, 131.0, 128.9, 123.8, 123.1, 115.3, 114.4, 113.6, 101.0, 100.4, 84.2, 73.8, 61.0, 58.0, 55.2, 49.4, 48.0, 42.8,

42.1, 42.0, 32.5, 31.3, 29.7, 28.2, 20.8, 19.2, 18.9, 17.6, 16.1, 10.8, 9.5; HRMS (ESI) (m/z): [M+Na]⁺ calcd for C₄₂H₅₈O₇, 697.4080, found, 697.4089; IR (film): 2956, 1673, 1588, 1513, 1454, 1413, 1384, 1337, 1302, 1247, 1161, 1032 cm⁻¹; [α]_D²⁰ +42.0 (c 1.64, CH₂Cl₂).



(S)-4-Methylhex-5-enoic acid (30).

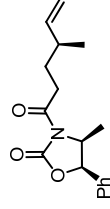
β(-)-Citronellene (3.8 g, 27.48 mmol, 1.0 equiv.) and sodium acetate (2.4 g, 28.85 mmol, 1.05 equiv.) were dissolved in CH₂Cl₂ (80 mL) and cooled to -20 °C. *m*-CPBA (75%, 6.64 g, 28.85 mmol, 1.05 equiv.) was added in small portions and stirring was continued for 1.5 h, allowing the suspension to warm to 0 °C. The reaction was quenched by careful addition of saturated aqueous NaHCO₃ (50 mL) and extracted with CH₂Cl₂ (3 x 50 mL). The combined organic fractions were washed with 1 N NaOH (10 mL), dried over MgSO₄ and concentrated *in vacuo*.

The crude epoxide was dissolved in Et₂O (70 mL), cooled to 0 °C and H₃IO₆ (6.26 g, 27.48 mmol, 1.0 equiv.) in THF (30 mL) was added within 25 min. Stirring was continued until TLC analysis showed complete consumption of the starting material. The mixture was diluted with Et₂O (500 mL), H₂O (300 mL) was added and the phases were separated. The organic layer was washed twice with brine, dried over MgSO₄ and evaporated to dryness.

The crude aldehyde was dissolved in 180 mL *t*-BuOH (0.15 M), treated with 2-methyl-2-butene (1 mL / mmol, 27 mL), and cooled to 10 °C. NaClO₂ (46.5 g, 412.20 mmol, 15 equiv.) and 46.5 g NaH₂PO₄ were dissolved in H₂O (270 mL, 1.5 M), transferred to a 500 mL dropping funnel, and added over a period of 30 min. After 1.5 h at 0 °C TLC analysis showed complete consumption and the reaction mixture was diluted with Et₂O (400 mL). The organic layer was extracted with 1N NaOH (3 x 70 mL), washed with Et₂O (3 x 30 mL) and acidified with 1 N HCl. The aqueous phase was extracted with CH₂Cl₂ (4 x 50 mL), the combined organic extracts were washed with brine (60 mL) and dried over MgSO₄. Evaporation of the solvent gave pure acid **30** (2.56 g, 73% 3 steps).

¹H-NMR (400MHz, CDCl₃): δ 5.64 (ddd, *J* = 17.4, 9.9, 7.7 Hz, 1H), 5.02–5.94 (m, 2H), 2.35 (dt, *J* = 8.7, 6.5 Hz, 2 H), 2.21–2.12 (m, 1H), 1.73–1.55 (m, 2H), 1.02 (d, *J* = 6.8 Hz, 3H); ¹³C-NMR

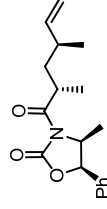
(100MHz, CDCl₃): δ 180.1, 143.2, 113.8, 37.4, 31.9, 31.1, 20.1; IR (film): 3853-2932, 2966, 1711, 1641, 1456, 1420, 1376, 1280, 1217, 1110 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₇H₁₂O₂, 128.0837, found, 128.0839; [α]_D²⁰+11.5 (c 1.00, CH₂Cl₂).



(4S,5R)-4-Methyl-3-((S)-4-methylhex-5-en-1-yl)-5-phenyloxazolidin-2-one (33).

A mixture of acid **30** (200 mg, 1.56 mmol, 1.0 equiv.), (4S, 5R)-4-methyl-5-phenyloxazolidin-2-one (304 mg, 1.72 mmol, 1.1 equiv.), DIC (151 μL, 1.72 mmol, 1.1 equiv.) and DMAP (190 mg, 1.56 mmol, 1.0 equiv.) in CH₂Cl₂ (8 mL) was stirred at ambient temperature overnight. The reaction was diluted with EE (100 mL), quenched with 1% HCl (2 mL) and washed with brine (20 mL). The organic layer was dried over MgSO₄, concentrated and the residue was purified by flash chromatography (hexane : ethylacetate = 5 : 1) to furnish **33** (367 mg, 82%) as a white solid.

¹H-NMR (400MHz, CDCl₃): δ 7.45-7.34 (m, 3H), 7.33-7.28 (m, 2H), 5.69 (ddd, *J* = 17.9, 10.6, 7.5 Hz, 1H), 5.65 (d, *J* = 7.3 Hz, 1H), 5.05-4.94 (m, 2H), 4.80-4.71 (m, 1H), 3.04-2.85 (m, 2H), 2.28-2.16 (m, 1H), 1.74-1.65 (m, 2H), 1.05 (d, *J* = 6.8 Hz, 3H), 0.89 (d, *J* = 6.6 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 173.2, 153.0, 143.6, 133.4, 128.7, 125.6, 113.6, 78.9, 54.7, 37.4, 33.5, 30.8, 20.2, 14.5; IR (film): 3070, 2962, 1783, 1699, 1640, 1347, 1197, 1122, 1090, 1067, 1040 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₁₇H₂₄O₃N, 287.1521, found, 287.1519; [α]_D²⁰-27.6 (c 0.95, CH₂Cl₂).

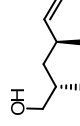


(4S,5R)-3-((2S,4S)-2,4-Dimethylhex-5-en-1-yl)-4-methyl-5-phenyloxazolidin-2-one (79).

A solution of compound **33** (60 mg, 0.208 mmol, 1.0 equiv.) in THF (1.4 mL) was cooled to -78 °C and treated with LHMDS (1 M in THF, 229 μL, 0.229 mmol, 1.1 equiv.). After 1 h at -78 °C, MeI

(40 μL, 0.626 mmol, 3 equiv.) was added and the solution was allowed to warm to room temperature overnight. The solution was quenched by the addition of saturated aqueous NH₄Cl (0.2 mL), diluted with CH₂Cl₂ (50 mL) and washed with brine (10 mL). The organic extracts were dried over MgSO₄, concentrated *in vacuo* and purified by flash chromatography (hexane : ethylacetate from 10 : 1 to 5 : 1) to afford 46 mg (74%, dr = 10:1 as determined by ¹H-NMR) of **79** as an oil.

¹H-NMR (400MHz, CDCl₃): δ 7.44-7.34 (m, 3H), 7.33-7.28 (m, 2H), 5.69 (ddd, *J* = 17.9, 9.7, 8.0 Hz, 1H), 5.63 (d, *J* = 7.3 Hz, 1H), 4.98-4.91 (m, 2H), 4.79-4.71 (m, 1H), 3.83-3.73 (m, 1H), 2.23-2.12 (m, 1H), 1.84 (ddd, *J* = 13.6, 8.7, 6.0 Hz, 1H), 1.37 (ddd, *J* = 13.8, 8.3, 5.6 Hz, 1H), 1.19 (d, *J* = 6.8 Hz, 3H), 1.02 (d, *J* = 6.8 Hz, 3H), 0.88 (d, *J* = 6.6 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 176.9, 152.5, 144.1, 133.4, 128.7, 125.6, 113.1, 78.8, 54.9, 40.0, 36.1, 35.7, 20.3, 17.8, 14.4; IR (film): 2967, 1783, 1701, 1456, 1343, 1247, 1196, 1121, 1068 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₁₈H₂₃O₃N, 301.1678, found, 301.1680; [α]_D²⁰+5.7 (c 1.20, CH₂Cl₂).



(2S,4S)-2,4-Dimethylhex-5-en-1-ol (34).

Oxazolidinone **79** (70 mg, 0.232 mmol, 1.0 equiv.) in THF (2.5 mL) was treated with LAH (18 mg, 0.487 mmol, 2.1 equiv.) at 0 °C. The reaction mixture was quenched after 1 h by slow addition of ethylacetate, diluted with Et₂O (50 mL) and washed with 1% HCl (10 mL). The aqueous layer was extracted with Et₂O (3 x 10 mL), and the combined organic fractions were washed with brine (20 mL), dried over MgSO₄ and concentrated *in vacuo*. Purification by column chromatography (pentane : diethylether = 3 : 1) afforded alcohol **34** as a colorless oil (23 mg, 80%).

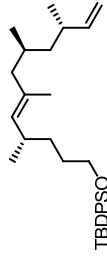
¹H-NMR (400MHz, CDCl₃): δ 5.71 (ddd, *J* = 17.4, 10.1, 7.5 Hz, 1H), 5.01-4.89 (m, 2H), 3.69-3.57-3.50 (m, 1H), 3.45-3.38 (m, 1H), 2.30-2.19 (m, 1H), 1.76-1.66 (m, 1H), 1.25 (t, *J* = 5.4 Hz, OH), 1.32 (ddd, *J* = 13.8, 7.0, 6.6 Hz, 1H), 1.19-1.11 (m, 1H), 0.98 (d, *J* = 6.6 Hz, 3H), 0.93 (d, *J* = 6.8 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 145.0, 112.4, 68.2, 40.2, 35.2, 33.2, 20.1, 16.9; IR (film): 3356,

2959, 2925, 2361, 1458 cm^{-1} ; HRMS (ESI) (m/z): $[M]^+$ calcd for $\text{C}_8\text{H}_{16}\text{O}$, 128.1201, found, 128.1203; $[\alpha]_D^{20} +1.1$ (c 1.00, CH_2Cl_2).



(3S,5S)-6-iodo-3,5-dimethylhex-1-ene (28).

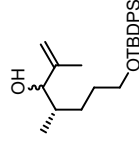
Alcohol **34** (4.50 g, 35.09 mmol, 1.0 equiv.) was dissolved in CH_2Cl_2 (115 mL), cooled to 0 °C and treated with Et_3N (7.29 mL, 52.62 mmol, 1.5 equiv.). After 5 min MsCl (8.3 mL, 49.13 mmol, 1.4 equiv.) was added and stirring was continued for 30 min. The solution was poured onto H_2O (100 mL), extracted with CH_2Cl_2 (3 x 50 mL), washed with brine and dried over MgSO_4 . The solvent was evaporated under reduced pressure and the crude mesylate was immediately dissolved in acetone (170 mL). NaI (36.8 g, 245 mmol, 7 equiv.) was added and the solution was refluxed for 4 h. The mixture was cooled to room temperature, filtered over Celite and saturated aqueous $\text{Na}_2\text{S}_2\text{O}_3$ (40 mL) was added. The solution was diluted with H_2O (200 mL) and the aqueous layer was extracted with Et_2O (3 x 50 mL). The combined organic fractions were dried over MgSO_4 , filtered and evaporated to dryness. Purification of the crude product by column chromatography (pentane : diethylether = 10 : 1) yielded 7.2 g (86%) of pure iodide **28**. ^1H -NMR (400MHz, CDCl_3): δ 5.66 (ddd, J = 17.5, 10.0, 7.6 Hz, 1H), 5.01–4.91 (m, 2H), 3.25 (dd, J = 9.7, 4.2 Hz, 1H), 3.18 (dd, J = 9.9, 5.8 Hz, 1H), 2.24–2.12 (m, 1H), 1.50–1.41 (m, 1H), 1.35–1.27 (m, 1H), 1.24–1.16 (m, 1H), 0.99 (d, J = 6.6 Hz, 3H), 0.96 (d, J = 6.6 Hz, 3H); ^{13}C -NMR (100MHz, CDCl_3): δ 144.2, 113.0, 43.3, 35.4, 31.9, 21.1, 20.5, 18.2; IR (film): 2959, 2925, 2361, 1456, 1377, 1192, 1120 cm^{-1} ; HRMS (ESI) (m/z): $[M]^+$ calcd for $\text{C}_8\text{H}_{15}\text{I}$, 238.0218, found, 238.0223; $[\alpha]_D^{20} +10.1$ (c 1.20, CH_2Cl_2).



tert-Butyldiphenyl((4S,8S,10S,F)-4,6,8,10-tetramethyldodeca-5,11-dienyloxy)silane (35).

A mixture of flame-dried ZnCl_2 (51 mg, 0.38 mmol, 1.87 equiv.) and alkyl iodide **28** (90 mg, 0.38 mmol, 1.87 equiv.) in degassed Et_2O (3 mL) was cooled to –78 °C and $t\text{-BuLi}$ (1.7 M in pentane, 443 μL , 0.75 mmol, 3.74 equiv.) was added *via* syringe in one portion. The solution was placed on an ice-bath and stirred for 1 h. To this white suspension, vinyl iodide **27** (99 mg, 0.201 mmol, 1.0 equiv.) and $\text{Pd}(\text{PPh}_3)_4$ (12 mg, 0.010 mmol, 0.05 equiv.) in degassed THF (2 mL) were added dropwise *via* cannula. The suspension cleared within 2 h at 0 °C, whereas TLC analysis showed complete consumption of the vinyl iodide. The mixture was treated with saturated aqueous NH_4Cl , extracted with Et_2O and the organic layer was washed with brine. Purification by column chromatography (hexane : ethylacetate = 50 : 1) gave alkene **35** (90 mg, 95%) as an oil.

^1H -NMR (400MHz, CDCl_3): δ 7.70–7.64 (m, 4H), 7.44–7.34 (m, 6H), 5.70 (ddd, J = 17.3, 10.0, 7.5 Hz, 1H), 4.99–4.87 (m, 2H), 4.85 (d, J = 9.3 Hz, 1H), 3.63 (t, J = 6.4 Hz, 2H), 2.36–2.26 (m, 1H), 2.27–2.19 (m, 1H), 2.06–1.97 (m, 1H), 1.68–1.60 (m, 2H), 1.60–1.46 (m, 2H), 1.51 (d, J = 1.1 Hz, 3H), 1.42–1.32 (m, 1H), 1.28–1.19 (m, 1H), 1.18–1.09 (m, 2H), 0.95 (d, J = 6.6 Hz, 3H), 0.90 (d, J = 6.6 Hz, 3H), 0.78 (d, J = 6.3 Hz, 3H); ^{13}C -NMR (100MHz, CDCl_3): δ 145.4, 135.6, 134.2, 133.0, 132.4, 129.4, 127.5, 112.0, 64.2, 47.8, 44.2, 35.3, 33.9, 32.1, 30.7, 28.1, 26.9, 21.4, 20.1, 19.7, 19.2, 16.0; IR (film): 2957, 2861, 1456, 1428, 138, 1112 cm^{-1} ; HRMS (ESI) (m/z): $[\text{M}^+\text{tBu}]^+$ calcd for $\text{C}_{28}\text{H}_{39}\text{OSi}$, 419.2770, found, 419.2772; $[\alpha]_D^{20} +8.2$ (c 1.30, CH_2Cl_2).



(4S)-7-(tert-Butyldiphenylsilyloxy)-2,4-dimethylhept-1-en-3-ol (29a) and (3R,4S)-7-(tert-Butyldiphenylsilyloxy)-2,4-dimethylhept-1-en-3-ol (29)

Method A:

A Schlenk flask was charged with anhydrous CrCl_2 (108 mg, 0.879 mmol, 4 equiv.) and NiCl_2 (1 mg, 0.009 mmol, 0.04 equiv.) and cooled to 0 °C. To this mixture degassed DMF (3.3 mL) was added and stirring was continued for 10 min at 0 °C. The dark-green solution was warmed to ambient temperature and aldehyde **36** (78 mg, 0.219 mmol, 1.0 equiv.) in 0.5 mL of degassed DMF, followed by 2-bromopropene (38 μL , 0.439 mmol, 2 equiv.) was added. The reaction

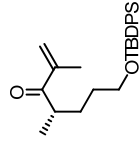
mixture was stirred for 3 h until TLC analysis revealed full consumption of the starting material. Finally the reaction was quenched by the addition of diethylether (5 ml) and water (5 mL). The aqueous phase was extracted with Et₂O (3 x 10 mL). The combined organic fractions were dried over MgSO₄ and the solvent was removed *in vacuo*. Purification of the residue by HPLC (hexane : ethylacetate = 10 : 1) afforded 40 mg of alcohol **29a** and 34 mg of alcohol **29b** (74 mg, dr = 1.4 : 1, 86% for both diastereomers).

Method B:

Ketone **80** (250 mg, 0.63 mmol, 1 equiv.) in MeOH (10 mL) was treated with CeCl₃ (710 mg, 1.90 mmol, 3 equiv.) and NaBH₄ (70 mg, 1.90 mmol, 3 equiv.) at 0 °C. The reaction was quenched after 5 min by the addition of 1% HCl (1 mL), diluted with H₂O (50 mL) and extracted with Et₂O (4 x 20 mL). The combined organic fractions were washed with brine (20 mL), dried over MgSO₄ and concentrated. The diastereomeric alcohols (250 mg, **29a** : **29b** = 2 : 1, 99% for both diastereomers) were separated as described above.

29a: ¹H-NMR (400MHz, CDCl₃): δ 7.70-7.65 (m, 4H), 7.45-7.35 (m, 6H), 4.92-4.85 (m, 2H), 3.76 (dd, *J* = 7.3, 3.3 Hz, 1 H), 3.67 (t, *J* = 6.3 Hz, 2H), 1.72-1.66 (m, 1H), 1.71-1.66 (m, 1H), 1.70 (s, 3H), 1.68-1.56 (m, 1H), 1.56-1.44 (m, 1H), 1.18-1.09 (m, 1H), 1.05 (s, 9H), 0.82 (d, *J* = 6.8 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 146.6, 135.6, 134.1, 129.5, 127.6, 112.3, 80.8, 64.3, 35.6, 30.1, 27.7, 26.9, 19.2, 17.5, 16.2; IR (film): 3446, 3070, 2930, 2858, 1652, 1558, 1472, 1428, 1387, 1112, 900 cm⁻¹; HRMS (ESI) (*m/z*): [M-tBu]⁺ calcd for C₂₁H₂₇O₂Si, 339.1780, found, 339.1778; [α]_D²⁰ -5.0 (c 1.2, CH₂Cl₂).

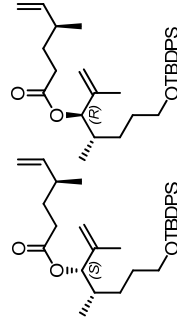
29b: ¹H-NMR (400MHz, CDCl₃): δ 7.69-7.64 (m, 4H), 7.45-7.35 (m, 6H), 4.95-4.86 (m, 2H), 3.86-3.81 (br, 1H), 3.65 (t, *J* = 6.3 Hz, 2H), 1.68(s, 3H), 1.67-1.52 (m, 2H), 1.64-1.57 (m, 1H), 1.51-1.42 (m, 1H), 1.23-1.10 (m, 1H), 1.05 (s, 9H), 0.88 (d, *J* = 6.6 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 146.6, 135.6, 134.1, 129.5, 127.6, 112.3, 80.8, 64.3, 35.6, 30.1, 27.7, 26.9, 19.2, 17.5, 16.2; IR (film): 3442, 3071, 2932, 2858, 1653, 1559, 1472, 1428, 1389, 1361, 1112, 900 cm⁻¹; HRMS (ESI) (*m/z*): [M-tBu]⁺ calcd for C₂₁H₂₇O₂Si, 339.1780, found, 339.1778; [α]_D²⁰ -6.6 (c 1.2, CH₂Cl₂).



(5S)-7-(tert-Butyldiphenylsilyloxy)-2,4-dimethylhept-1-en-3-one (**80**).

A solution of alcohol **29b** (335 mg, 0.85 mmol, 1.0 equiv.) in CH₂Cl₂ (4 mL) was cooled to -5 °C. Et₃N (351 μL, 2.53 mmol, 3 equiv.), followed by SO₃Pyr (403 mg, 2.53 mmol, 3 equiv.) in DMSO (4 mL) were added dropwise. The mixture was stirred for 1.5 h at -5°C and quenched with aqueous 1 M KHSO₄ solution (1 mL). The phases were partitioned between brine and Et₂O (1 : 1, 60 mL) and the aqueous layer was extracted with Et₂O (3 x 10 mL). The combined organic fractions were concentrated to 5 mL under reduced pressure, filtered over a plug of silica and excess solvent was removed *in vacuo* to afford ketone **80** (331 mg, 99%).

¹H-NMR (400MHz, CDCl₃): δ 7.69-7.62 (m, 4H), 7.45-7.34 (m, 6H), 5.89 (br, 1H), 5.73 (br, 1H), 3.63 (t, *J* = 6.2 Hz, 2H), 3.26-3.15 (m, 1H), 1.87 (s, 3H), 1.78-1.68 (m, 1H), 1.56-1.40 (m, 3H), 1.06 (d, *J* = 7.1 Hz, 3H), 1.04 (s, 9H); ¹³C-NMR (100MHz, CDCl₃): δ 206.1, 144.1, 135.6, 134.0, 129.5, 127.6, 123.9, 63.7, 39.2, 30.2, 30.1, 26.9, 19.2, 18.0, 17.5; IR (film): 3071, 2931, 2858, 1675, 1629, 1589, 1461, 1428, 1378, 1189, 1112 cm⁻¹; HRMS (ESI) (*m/z*): [M-tBu]⁺ calcd for C₂₁H₂₅O₂Si, 337.1624, found, 337.1617; [α]_D²⁰ +8.3 (c 1.15, CH₂Cl₂).



(5S)-7-(tert-Butyldiphenylsilyloxy)-2,4-dimethylhept-1-en-3-yl 4-methylhex-5-enoate (**37a**) and (5S)-7-(tert-butylidiphenylsilyloxy)-2,4-dimethylhept-1-en-3-yl 4-methylhex-5-enoate (**37b**).

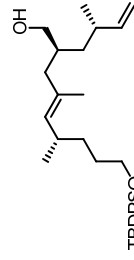
A mixture of acid **30** (65 mg, 0.507 mmol, 1.0 equiv.), alcohol **29** (221 mg, 0.557 mmol, 1.1 equiv.), DIC (49 μL, 0.557 mmol, 1.1 equiv.) and DMAP (62 mg, 0.507 mmol, 1.0 equiv.) in CH₂Cl₂ (4 mL) was stirred at ambient temperature overnight. The reaction was diluted with CH₂Cl₂ (70

mL), quenched with 1% HCl (10 mL) and washed with brine (20 mL). The organic layer was dried over MgSO₄, concentrated and the residue was purified by flash chromatography (hexane : ethylacetate = 50 : 1) to furnish ester **37a** or **37b** (236 mg, 92%).

37a: ¹H-NMR (400MHz, CDCl₃): δ 7.69-7.64 (m, 4H), 7.45-7.35 (m, 6H), 5.62 (ddd, *J* = 17.3, 10.0, 7.6 Hz, 1H), 4.98-4.90 (m, 4H), 4.95 (d, *J* = 7.1 Hz, 1H), 3.64 (dt, *J* = 6.0, 1.9 Hz, 2H), 2.36-2.22 (m, 2H), 2.18-2.06 (m, 1H), 1.81-1.72 (m, 1H), 1.69 (s, 3H), 1.68-1.55 (m, 4H), 1.53-1.42 (m, 1H), 1.15-1.06 (m, 1H), 1.05 (s, 9H), 0.99 (d, *J* = 6.8 Hz, 3H), 0.83 (d, *J* = 6.8 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 173.1, 143.5, 142.2, 135.6, 134.1, 129.5, 127.6, 113.9, 113.6, 81.4, 64.1, 37.5, 34.2, 32.3, 31.4, 30.0, 27.9, 26.9, 20.1, 19.2, 18.4, 15.8; IR (film): 3072, 2931, 2859, 1737, 1653, 1462, 1428, 1378, 1254, 1178, 1111 cm⁻¹; HRMS (ESI) (*m/z*): [M+*t*Bu]⁺ calcd for C₂₈H₃₇O₃Si, 449.2512, found, 449.2519; [α]_D²⁰ -0.6 (c 0.90, CH₂Cl₂).

37b: ¹H-NMR (400MHz, CDCl₃): δ 7.68-7.63 (m, 4H), 7.45-7.34 (m, 6H), 5.64 (ddd, *J* = 17.4, 10.1, 7.6 Hz, 1H), 5.02 (d, *J* = 6.3 Hz, 1H), 5.00-4.92 (m, 2H), 4.91-4.86 (m, 2H), 3.64 (t, *J* = 6.3 Hz, 2H), 2.31 (ddd, *J* = 7.0, 8.7, 3.3 Hz, 2H), 2.20-2.08 (m, 1H), 1.80-1.70 (m, 1H), 1.69 (s, 3H), 1.69-1.57 (m, 2H), 1.57-1.48 (m, 2H), 1.47-1.37 (m, 1H), 1.20-1.10 (m, 1H), 1.04 (s, 9H), 1.01 (d, *J* = 6.8 Hz, 3H), 0.83 (d, *J* = 6.6 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 173.0, 143.5, 142.3, 135.6, 134.0, 129.5, 127.6, 113.6, 113.1, 80.3, 63.9, 37.5, 34.3, 32.3, 31.4, 30.0, 29.2, 26.8, 20.1, 19.2, 18.7, 14.5; IR (film): 3072, 2933, 2858, 1736, 1428, 1379, 1176, 1112 cm⁻¹; HRMS (ESI) (*m/z*): [M+*t*Bu]⁺ calcd for C₂₈H₃₇O₃Si, 449.2512, found, 449.2517; [α]_D²⁰ +4.0 (c 1.05, CH₂Cl₂).

55



(2S,6S,E)-9-(tert-Butyldiphenylsilyloxy)-4,6-dimethyl-2-((S)-2-methylbut-3-enyl)non-4-en-1-ol (39).

Freshly prepared LDA (1 M in THF, 0.49 mL, 0.49 mmol, 1.25 equiv.) was cooled to -78 °C and freshly distilled HMPA (120 μL) was added *via* syringe. After 5 min ester **37a** (190 mg, 0.374 mmol, 1.0 equiv.) was added as a solution in THF (1 mL). After 20 min TBSCl (4 M in THF, 103 μL, 0.41 mmol, 1.1 equiv.) was added to the reaction mixture and stirring was continued for 20 min

at -78 °C. The solution was allowed to warm to ambient temperature and refluxed for 2 h. The reaction mixture was diluted at room temperature by the addition of pentane : Et₂O (1 : 1, 70 mL) and washed with brine (30 mL). The aqueous layer was extracted with Et₂O (3 x 15 mL), the combined organic fractions were dried over MgSO₄ and the solvent was removed *in vacuo*.

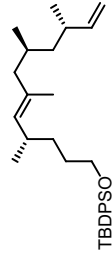
The crude TBS-ester was dissolved in HMPA (1 mL) followed by the addition of KHCO₃ (56 mg, 0.562 mmol, 1.5 equiv.) and KF·H₂O (33 mg, 0.562 mmol, 1.5 equiv.). After 30 min MeI (47 μL, 0.749 mmol, 2.0 equiv.) was added and stirring was continued over night. The reaction mixture was partitioned between Et₂O (50 mL) and H₂O (20 mL), the aqueous layer was extracted with Et₂O (2 x 10 mL) and the organic fraction was dried over MgSO₄. The mixture was filtered and concentrated to about 5 mL.

The flask was transferred to an ice-bath and LAH (30 mg, 0.785 mmol, 2.1 equiv.) was added. After 30 min at 0 °C the reaction mixture was quenched by the slow addition of ethylacetate, diluted with Et₂O (50 mL) and washed with 1% HCl (20 mL). The aqueous layer was extracted with Et₂O (3 x 20 mL), the combined organic fractions were washed with brine (30 mL), dried over MgSO₄ and the solvent was removed *in vacuo*. Purification by column chromatography (hexane : ethylacetate from 10 : 1 to 5 : 1) afforded alcohol **39** as a colorless oil (138 mg, 63%, dr = 5 : 1 as determined by ¹H-NMR).

S-39: ¹H-NMR (400MHz, CDCl₃): δ 7.69-7.64 (m, 4H), 7.44-7.35 (m, 6H), 5.66 (ddd, *J* = 17.2, 10.1, 8.1 Hz, 1H), 5.01-4.86 (m, 2H), 4.93 (d, *J* = 10.3 Hz, 1H), 3.63 (t, *J* = 6.4 Hz, 2H), 3.50-3.45 (m, 2H), 2.37-2.27 (m, 1H), 2.27-2.17 (m, 1H), 2.01 (dd, *J* = 13.3, 8.7, 1H), 1.92 (dd, *J* = 13.5, 8.7 Hz, 1H), 1.76-1.66 (m, 1H), 1.56 (d, *J* = 1.3 Hz, 3H), 1.56-1.46 (m, 2H), 1.41-1.34 (m, 1H), 1.39-1.30 (m, 1H), 1.26-1.20 (m, 1H), 1.20-1.12 (m, 1H), 1.05 (s, 9H), 0.99 (d, *J* = 6.8 Hz, 3H), 0.90 (d, *J* = 6.8 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 144.7, 135.6, 134.2, 133.5, 132.4, 129.5, 127.6, 112.8, 66.3, 64.1, 42.4, 38.5, 35.9, 35.7, 33.7, 32.2, 30.6, 26.9, 21.1, 21.0, 19.2, 16.2; IR (film): 3394, 3072, 2930, 1639, 1590, 1453, 1387, 1112 cm⁻¹; HRMS (ESI) (*m/z*): [M+*t*Bu]⁺ calcd for C₂₈H₃₉O₂Si, 435.2719, found, 435.2722; [α]_D²⁰ +7.9 (c 1.00, CH₂Cl₂).

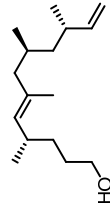
R-39: ¹H-NMR (400MHz, CDCl₃): δ 7.69-7.64 (m, 4H), 7.45-7.35 (m, 6H), 5.63 (ddd, *J* = 17.4, 10.0, 7.7 Hz, 1H), 5.01-4.88 (m, 3H), 3.63 (t, *J* = 6.4 Hz, 2H), 3.59-3.45 (m, 2H), 2.38-2.28 (m, 1H), 2.31-2.19 (m, 1H), 2.02 (dd, *J* = 13.4, 8.1, 1H), 1.93 (dd, *J* = 13.4, 6.6 Hz, 1H), 1.79-1.69 (m, 1H), 1.59-

1.56 (m, 2H), 1.57 (s, 3H), 1.43-1.33 (m, 1H), 1.30-1.18 (m, 3H), 1.05 (s, 9H), 0.98 (d, $J = 6.6$ Hz, 3H), 0.91 (d, $J = 6.8$ Hz, 3H); $^{13}\text{C-NMR}$ (100MHz, CDCl_3): δ 144.7, 135.6, 134.2, 133.5, 132.6, 129.5, 127.6, 112.9, 65.9, 64.1, 43.2, 38.1, 35.7, 35.6, 33.7, 32.1, 30.6, 26.9, 21.3, 21.1, 19.2, 16.2; IR (film): 3356, 3072, 2930, 2859, 1640, 1590, 1472, 1428, 1388, 1112 cm^{-1} ; HRMS (ESI) (m/z): $[\text{M}+\text{tBu}]^+$ calcd for $\text{C}_{28}\text{H}_{39}\text{O}_2\text{Si}$, 435.2719, found, 435.2716; $[\alpha]_D^{20} +8.8$ (c 0.90, CH_2Cl_2).



tert-Butyldiphenyl((4S,8S,10S,E)-4,6,8,10-tetramethyldodeca-5,11-dienyloxy)silane (35).

Alcohol **39** (100 mg, 0.20 mmol, 1.0 equiv.) was dissolved in CH_2Cl_2 (5 mL), cooled to 0 °C and treated with Et_3N (34 μL , 0.24 mmol, 1.2 equiv.). After 5 min MsCl (19 μL , 0.24 mmol, 1.2 equiv.) was added and stirring was continued for 30 min. The solution was poured onto H_2O (10 mL), extracted with CH_2Cl_2 (3 x 5 mL), washed with brine and dried over MgSO_4 . Evaporation of the solvent gave the crude mesylate, which was immediately dissolved in Et_2O (2 mL). LAH (11 mg, 0.30 mmol, 1.5 equiv.) was carefully added to the ice cooled solution and the cloudy mixture was allowed to warm to room temperature over 30 min. After 2 h the reaction mixture was quenched at 0 °C by slow addition of ethylacetate, diluted with Et_2O (20 mL) and washed with 1% HCl (10 mL). The aqueous layer was extracted with Et_2O (3 x 5 mL), and the combined organic fractions were washed with brine, dried over MgSO_4 and the solvent was removed *in vacuo*. Purification by flash chromatography (hexane : ethylacetate = 50 : 1) afforded alkene **35** as an oil (86 mg, 90%). Analytical data matched those reported on pp. 16.

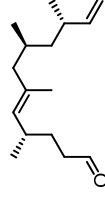


(4S,8S,10S,E)-4,6,8,10-Tetramethyldodeca-5,11-dien-1-ol (81).

A solution of alkene **35** (1.70 g, 3.57 mmol, 1.0 equiv.) in THF (70 mL) was treated with TBAF (1 M in THF, 4.27 mL, 4.28 mmol, 1.2 equiv.) and stirred overnight at room temperature. Finally

the reaction was quenched with 50 mL of NH_4Cl and the aqueous layer was extracted with Et_2O (3 x 30 mL). The combined organic extracts were dried over MgSO_4 , filtered and evaporated to dryness. Purification by column chromatography using gradient elution (hexane : ethylacetate from 10 : 1 to 5 : 1) furnished alcohol **81** as a yellow oil (739 mg, 87%).

$^1\text{H-NMR}$ (400MHz, CDCl_3): δ 5.69 (ddd, $J = 17.4$, 10.2, 7.4 Hz, 1H), 4.99-4.86 (m, 2H), 4.85 (d, $J = 9.6$ Hz, 1H), 3.61 (q, $J = 6.1$ Hz, 2H), 2.40-2.30 (m, 1H), 2.27-2.18 (m, 1H), 2.07-1.97 (m, 1H), 1.70-1.62 (m, 2H), 1.58-1.48 (m, 2H), 1.55 (d, $J = 1.3$ Hz, 3H), 1.43-1.33 (m, 1H), 1.28-1.18 (m, 1H), 1.19-1.09 (m, 2H), 0.95 (d, $J = 6.8$ Hz, 3H), 0.93 (d, $J = 6.8$ Hz, 3H), 0.79 (d, $J = 6.1$ Hz, 3H); $^{13}\text{C-NMR}$ (100MHz, CDCl_3): δ 145.4, 132.7, 132.6, 112.0, 63.3, 47.8, 44.2, 35.3, 33.8, 32.2, 30.9, 28.1, 21.4, 20.1, 19.8, 16.1; IR (film): 3331, 2951, 2360, 1458, 1375, 1060 cm^{-1} ; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{16}\text{H}_{30}\text{O}$, 238.2297, found, 238.2296; $[\alpha]_D^{20} +17.9$ (c 1.55, CH_2Cl_2).

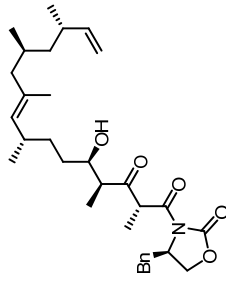


(4S,8S,10S,E)-4,6,8,10-tetramethyldodeca-5,11-dienal (25).

A 50 mL flask was charged with alcohol **81** (925 mg, 3.88 mmol, 1.0 equiv.) and DMSO (19 mL, 0.2 M). IBX (3.8 g, 13.58 mmol, 3.5 equiv.) was added and stirring was continued for 2 h at ambient temperature. The solution was diluted with Et_2O : hexane = 1 : 1 (50 mL) and H_2O (30 mL) to precipitate unreacted IBX. The mixture was filtered over Celite, the layers were separated and the aqueous layer was extracted with Et_2O : hexane = 1 : 1 (3 x 20 mL). The combined organic layers were dried over anhydrous MgSO_4 , filtered and concentrated *in vacuo*. The crude product was filtered over a plug of silica to give 870 mg (95%) of aldehyde **25**.

$^1\text{H-NMR}$ (400MHz, CDCl_3): δ 9.75 (t, $J = 1.8$ Hz, 1H), 5.69 (ddd, $J = 17.4$, 10.0, 7.5 Hz, 1H), 4.99-4.87 (m, 2H), 4.82 (d, $J = 9.6$ Hz, 1H), 2.42-2.30 (m, 3H), 2.29-2.18 (m, 1H), 2.07-1.98 (m, 1H), 1.73-1.62 (m, 3H), 1.53 (s, 3H), 1.52-1.41 (m, 1H), 1.19-1.09 (m, 2H), 0.96 (d, $J = 6.6$ Hz, 3H), 0.95 (d, $J = 6.8$ Hz, 3H), 0.79 (d, $J = 6.3$ Hz, 3H); $^{13}\text{C-NMR}$ (100MHz, CDCl_3): δ 202.9, 145.3, 134.0, 131.5, 112.0, 47.8, 44.1, 42.3, 35.3, 32.0, 29.8, 28.0, 21.3, 20.1, 19.8, 16.1; IR (film): 2957, 2924,

2869, 2714, 1728, 1640, 1456, 1375, 1112 cm^{-1} ; HRMS (ESI) (m/z): $[M]^+$ calcd for $\text{C}_{16}\text{H}_{28}\text{O}_5$, 236.2140, found, 236.2135; $[\alpha]_D^{20} + 16.1$ (c 1.20, CH_2Cl_2).

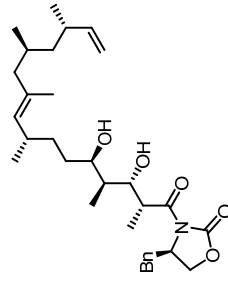


(2R,4S,5R,8S,12S,14S,E)-1-((R)-4-Benzyl-2-oxoxazolidin-3-yl)-5-hydroxy-2,4,8,10,12,14-hexamethylhexadeca-9,15-diene-1,3-dione (82).

A 100 mL Schlenk flask was charged with acid free $\text{Sn}(\text{OTf})_2$ (3.77 g, 9.05 mmol, 2.2 equiv.) and CH_2Cl_2 (30 mL, 0.3 M). The white suspension was treated at -20°C with Et_3N (1.19 mL, 8.67 mmol, 2.1 equiv.) whereupon the mixture turned pale yellow. After 5 min β -keto imide **26** (1.82 g, 6.29 mmol, 1.5 equiv.) in CH_2Cl_2 (9 mL, 0.7 M) was added dropwise and the clear solution was stirred for 1 h at -20°C . Aldehyde **25** (960 mg, 4.06 mmol, 1.0 equiv.) was dissolved in CH_2Cl_2 (6.5 mL, 0.6 M) and slowly added at -78°C . After 1 h at -78°C TLC analysis showed complete consumption of the starting material and the yellow-orange solution was poured onto a cooled and vigorously stirred mixture of CH_2Cl_2 and 1 M NaHSO_4 (100 mL, 1 : 1). After 20 min at room temperature the aqueous phase was extracted with CH_2Cl_2 (3 x 25 mL), the organic phase was washed with saturated aqueous NaHCO_3 , dried over MgSO_4 and concentrated *in vacuo*. Purification of the residue by gradient flash chromatography (hexane : ethylacetate from 5 : 1 to 1 : 1) yielded **82** as a viscous oil (1.61 g, 76%, dr = 10 : 1 as determined by $^1\text{H-NMR}$).

$^1\text{H-NMR}$ (400MHz, CDCl_3): δ 7.37-7.27 (m, 3H), 7.22-7.17 (m, 2H), 5.69 (ddd, J = 17.4, 10.0, 7.0 Hz, 1H), 4.99-4.92 (m, 1H), 4.92-4.85 (m, 2H), 4.85-4.81 (m, 1H), 4.79-4.71 (m, 1H), 4.29-4.23 (m, 1H), 4.21-4.16 (m, 1H), 3.93-3.85 (br, 1H), 3.31 (dd, J = 13.3, 2.4 Hz, 1H), 2.82-2.73 (m, 2H), 2.44 (br, OH), 2.41-2.29 (m, 1H), 2.28-2.18 (m, 1H), 2.07-1.97 (m, 1H), 1.71-1.59 (m, 2H), 1.57-1.49 (m, 1H), 1.54 (d, J = 1.3 Hz, 3H), 1.47 (d, J = 7.1 Hz, 3H), 1.37-1.29 (m, 2H), 1.29-1.23 (m, 1H), 1.21 (d, J = 7.1 Hz, 3H), 1.19-1.11 (m, 2H), 0.95 (d, J = 6.8 Hz, 3H), 0.93 (d, J = 6.8 Hz, 3H),

0.79 (d, J = 6.1 Hz, 3H); $^{13}\text{C-NMR}$ (100MHz, CDCl_3): δ 169.7, 152.9, 145.4, 135.0, 132.8, 132.5, 129.4, 129.0, 127.4, 112.0, 71.2, 66.5, 55.3, 51.9, 48.4, 47.8, 44.2, 38.0 (2xCH), 35.3, 34.0, 32.2, 32.0, 28.1, 21.4, 20.1, 19.8, 16.1, 12.9, 9.9; IR (film): 3536, 2955, 1782, 1715, 1455, 1360, 1215, 1122 cm^{-1} ; HRMS (ESI) (m/z): $[M+\text{Na}]^+$ calcd for $\text{C}_{32}\text{H}_{47}\text{O}_5\text{NNa}$, 548.3352, found, 548.3346; $[\alpha]_D^{20}$ - 54.9 (c 1.00, CH_2Cl_2).

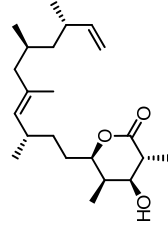


(R)-4-Benzyl-3-((2R,3S,4S,5R,8S,12S,14S,E)-3,5-dihydroxy-2,4,8,10,12,14-hexamethylhexadeca-9,15-dienyl)oxazolidin-2-one (40).

$\text{Me}_4\text{NBH}(\text{OAc})_3$ (3.74 g, 14.25 mmol, 5 equiv.) was dissolved in 400 mL MeCN : AcOH = 1.9 : 1, cooled to -32°C and aldol product **82** (1.5 g, 2.85 mmol, 1.0 equiv.) in MeCN (3 mL) was added dropwise. The reaction was stirred for 3 h at -32°C , allowed to warm to 0°C overnight, diluted with CH_2Cl_2 (70 mL) and quenched by the addition of saturated aqueous Rochelle's salt (40 mL). Saturated aqueous NaHCO_3 (50 mL) was carefully added to the vigorously stirred solution over 20 min. After 1 h no more gas evolution was observed, the aqueous layer was extracted with CH_2Cl_2 (3 x 30 mL), and the combined organic fractions were dried over MgSO_4 , filtered and concentrated *in vacuo*. Purification by flash chromatography (hexane : ethylacetate from 3 : 1 to 2 : 1) afforded diol **40** as a viscous oil (1.14 g, 76%, dr \geq 20 : 1 as determined by $^1\text{H-NMR}$).

$^1\text{H-NMR}$ (400MHz, CDCl_3): δ 7.37-7.27 (m, 3H), 7.23-7.18 (m, 2H), 5.69 (ddd, J = 17.4, 10.2, 7.4 Hz, 1H), 4.99-4.87 (m, 2H), 4.87 (d, J = 8.8 Hz, 1H), 4.73-4.66 (m, 1H), 4.26-4.17 (m, 2H), 3.96 (dd, J = 9.0, 2.4 Hz, 1H), 3.88 (dq, J = 7.0, 2.5 Hz, 1H), 3.80 (dt, J = 9.3, 2.3 Hz, 1H), 3.25 (dd, J = 13.4, 9.3 Hz, 1H), 2.79 (dd, J = 13.4, 9.3 Hz, 1H), 2.42-2.30 (m, 1H), 2.29-2.17 (m, 1H), 2.07-1.98 (m, 1H), 1.87-1.77 (m, 1H), 1.71-1.59 (m, 2H), 1.55-1.45 (m, 1H), 1.54 (d, J = 1.1 Hz, 3H), 1.45-1.29 (m, 2H), 1.41-1.30 (m, 1H), 1.27 (d, J = 7.1 Hz, 3H), 1.19-1.09 (m, 2H), 0.95 (d, J = 6.6 Hz,

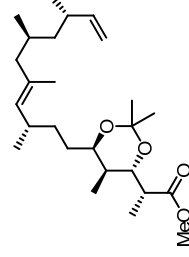
3H), 0.93 (d, $J = 6.1$ Hz, 3H), 0.83 (d, $J = 7.1$ Hz, 3H), 0.79 (d, $J = 6.1$ Hz, 3H); ^{13}C -NMR (100MHz, CDCl_3): δ 178.0, 152.8, 145.4, 134.9, 132.8, 132.6, 129.4, 129.0, 127.5, 112.0, 73.6 (2xCH), 66.2, 55.1, 47.8, 44.2, 39.6, 39.2, 37.8, 35.3, 34.5, 32.3, 31.0, 28.1, 21.4, 20.1, 19.7, 16.1, 11.5, 10.1; IR (film): 3475, 2956, 1783, 1700, 1455, 1387, 1211, 1107, cm^{-1} ; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{32}\text{H}_{49}\text{O}_5\text{N}$, 527.3611, found, 527.3615; $[\alpha]_D^{20}$ -23.9 (c 0.90, CH_2Cl_2).



(3R,4S,5R,6R)-4-Hydroxy-3,5-dimethyl-6-((3S,7S,9S,E)-3,5,7,9-tetramethylundeca-4,10-dienyl)tetrahydro-2H-pyran-2-one (83).

A solution of diol **40** (1.14 g, 2.16 mmol, 1.0 equiv.) in $\text{THF}/\text{H}_2\text{O} = 3 : 1$ (22 mL 0.1 M) was treated at 0°C with H_2O_2 (30% in H_2O , 864 μL). LiOH (145 mg, 3.46 mmol, 1.6 equiv.) was added in one portion and stirring was continued for 1 h at ambient temperature. The reaction was acidified with 1 N HCl and stirred for further 5 min. The mixture was diluted with H_2O (50 mL), extracted with Et_2O (4 x 30 mL), dried over MgSO_4 and concentrated under reduced pressure. Purification by flash chromatography (hexane : ethylacetate from 5 : 1 to 3 : 1) gave lactone **83** (700 mg, 92%) as an oil.

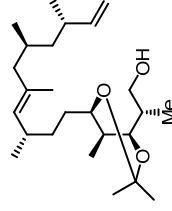
^1H -NMR (400MHz, CDCl_3): δ 5.68 (ddd, $J = 17.4, 10.0, 7.5$ Hz, 1H), 4.98-4.86 (m, 2H), 4.85 (d, $J = 9.4$ Hz, 1H), 4.16 (ddd, $J = 8.2, 5.4, 2.5$ Hz, 1H), 3.79 (dd, $J = 10.2, 4.2$ Hz, 1H), 2.44 (dq, $J = 10.4, 7.1$ Hz, 1H), 2.39-2.30 (m, 1H), 2.27-2.17 (m, 1H), 2.14-2.05 (m, 1H), 2.07-1.98 (m, 1H), 1.83-1.72 (m, 1H), 1.71-1.57 (m, 2H), 1.53 (d, $J = 1.2$ Hz, 3H), 1.48-1.35 (m, 1H), 1.42-1.29 (m, 2H), 1.38 (d, $J = 7.1$ Hz, 3H), 1.17-1.09 (m, 2H), 0.94 (d, $J = 6.8$ Hz, 3H), 0.93 (d, $J = 6.8$ Hz, 6H), 0.77 (d, $J = 6.3$ Hz, 3H); ^{13}C -NMR (100MHz, CDCl_3): δ 173.7, 145.3, 133.2, 132.1, 112.0, 78.0, 73.9, 47.7, 44.2, 39.8, 37.3, 35.3, 33.2, 32.1, 30.2, 28.0, 21.3, 20.1, 19.7, 16.1, 14.3, 4.4; IR (film): 3448, 2956, 2915, 1714, 1629, 1456, 1375, 1293, 1215, 1103, 1038 cm^{-1} ; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{27}\text{H}_{38}\text{O}_3$, 350.2821, found, 350.2832; $[\alpha]_D^{20}$ +44.7 (c 1.50, CH_2Cl_2).



(R)-Methyl 2-((4S,5S,6R)-2,2,5-trimethyl-6-((3S,7S,9S,E)-3,5,7,9-tetramethylundeca-4,10-dienyl)-1,3-dioxan-4-yl)propanoate (84).

Lactone **83** (700 mg, 1.99 mmol, 1.0 equiv.) was dissolved in 2,2-dimethoxypropane (20 mL), treated with camphorsulfonic acid (46 mg, 0.19 mmol, 0.1 equiv.) and stirred overnight at room temperature. The solution was diluted with Et_2O (100 mL), neutralized with saturated aqueous NaHCO_3 (40 mL), extracted with Et_2O (3 x 30 mL), washed with brine (50 mL) and dried over MgSO_4 . The solvent was removed *in vacuo* and the residue was purified by flash chromatography (hexane : ethylacetate = 10 : 1) to yield ester **84** (757 mg, 90%).

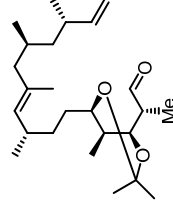
^1H -NMR (400MHz, CDCl_3): δ 5.69 (ddd, $J = 17.4, 10.1, 7.5$ Hz, 1H), 4.99-4.87 (m, 2H), 4.86 (d, $J = 9.4$ Hz, 1H), 3.72 (dd, $J = 9.1, 4.3$ Hz, 1H), 3.68 (s, 3H), 3.61 (dd, $J = 7.7, 4.9$ Hz, 1H), 2.60-2.52 (m, 1H), 2.41-2.29 (m, 1H), 2.27-2.17 (m, 1H), 2.07-1.97 (m, 1H), 1.84-1.74 (m, 1H), 1.71-1.60 (m, 2H), 1.54 (d, $J = 1.0$ Hz, 3H), 1.45-1.37 (m, 1H), 1.33-1.20 (m, 2H), 1.30 (s, 3H), 1.29 (s, 3H), 1.28-1.21 (m, 1H), 1.22-1.07 (m, 2H), 1.19 (d, $J = 7.1$ Hz, 3H), 0.95 (d, $J = 6.6$ Hz, 3H), 0.92 (d, $J = 6.6$ Hz, 3H), 0.81 (d, $J = 6.8$ Hz, 3H), 0.79 (d, $J = 6.3$ Hz, 3H); ^{13}C -NMR (100MHz, CDCl_3): δ 174.9, 145.4, 132.7, 132.5, 112.0, 100.5, 75.4, 69.3, 51.6, 47.8, 44.2, 43.0, 36.8, 35.3, 33.8, 32.3, 28.5, 28.1, 25.0, 23.7, 21.3, 20.1, 19.7, 16.1, 11.9, 11.4; IR (film): 2926, 1743, 1494, 1455, 1380, 1227, 1112, 1050 cm^{-1} ; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{56}\text{H}_{86}\text{O}_4$, 422.3396, found, 422.3394; $[\alpha]_D^{20}$ -12.8 (c 1.00, CH_2Cl_2).



(S)-2-((4*R*,5*S*,6*R*)-2,2,5-Trimethyl-6-((3*S*,7*S*,9*S*,*E*)-3,5,7,9-tetramethylundeca-4,10-dienyl)-1,3-dioxan-4-yl)propan-1-ol (85).

Ester **84** (730 mg, 1.73 mmol, 1.0 equiv.) in Et₂O (17 mL) was cooled in an ice-bath and LAH (138 mg, 3.63 mmol, 2.1 equiv.) was added carefully in small portions. After 30 min at 0 °C TLC analysis showed complete consumption of the starting material and the reaction mixture was quenched by slow addition of ethylacetate, diluted with Et₂O (80 mL) and washed with 1% HCl (100 mL). The aqueous layer was extracted with Et₂O (3 x 30 mL), and the combined organic fractions were washed with brine (50 mL), dried over MgSO₄ and concentrated *in vacuo*. Purification by column chromatography (hexane : ethylacetate = 2 : 1) afforded alcohol **85** as a colorless oil (677 mg, 99%).

¹H-NMR (400MHz, CDCl₃): δ 5.69 (ddd, *J* = 17.3, 10.0, 7.5 Hz, 1H), 4.98-4.87 (m, 2H), 4.87 (d, *J* = 8.1 Hz, 1H), 3.72 (dd, *J* = 8.8, 4.3 Hz, 1H), 3.69-3.63 (m, 2H), 3.50 (dd, *J* = 7.8, 2.8 Hz, 1H), 2.43 (t, *J* = 4.9 Hz, 1H), 2.40-2.30 (m, 1H), 2.29-2.17 (m, 1H), 2.07-1.98 (m, 1H), 1.89-1.75 (m, 2H), 1.71-1.60 (m, 2H), 1.55 (d, *J* = 1.2 Hz, 3H), 1.47-1.38 (m, 1H), 1.34 (s, 3H), 1.32-1.23 (m, 2H), 1.31 (s, 3H), 1.31-1.23 (m, 1H), 1.20-1.07 (m, 2H), 0.97 (d, *J* = 7.1 Hz, 3H), 0.95 (d, *J* = 6.6 Hz, 3H), 0.92 (d, *J* = 6.6 Hz, 3H), 0.81 (d, *J* = 6.6 Hz, 3H), 0.79 (d, *J* = 6.3 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 145.4, 132.7, 132.5, 112.0, 100.4, 77.7, 69.6, 67.3, 47.8, 44.2, 37.4, 35.8, 35.3, 33.8, 32.2, 28.4, 28.1, 25.2, 23.7, 21.3, 20.1, 19.7, 16.1, 12.2, 10.6; IR (film): 3424, 3074, 2958, 1639, 1457, 1380, 1226, 1167, 1019 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₂₅H₄₆O₃, 394.3447, found, 394.3445; [α]_D²⁰ +1.9 (*c* 1.05, CH₂Cl₂).

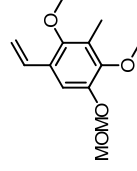


(R)-2-((4*R*,5*S*,6*R*)-2,2,5-Trimethyl-6-((3*S*,7*S*,9*S*,*E*)-3,5,7,9-tetramethylundeca-4,10-dienyl)-1,3-dioxan-4-yl)propanal (24).

To a solution of alcohol **85** (110 mg, 0.28 mmol, 1.0 equiv.) in DMSO (2 mL), IBX (273 mg, 0.98 mmol, 3.5 equiv.) was added over a period of 20 min and stirring was continued for 1.5 h at

ambient temperature. The solution was diluted with Et₂O : hexane = 1 : 1 (60 mL) and H₂O (20 mL) to precipitate unreacted IBX. The mixture was filtered over Celite, the layers were separated and the aqueous layer was extracted with Et₂O : hexane = 1 : 1 (3 x 20 mL). The combined organic layers were dried over anhydrous MgSO₄, filtered and concentrated *in vacuo*.

The crude product was filtered over a plug of silica to give aldehyde **24** as an oil (108 mg, 99%). ¹H-NMR (400MHz, CDCl₃): δ 9.70 (s, 1H), 5.69 (ddd, *J* = 17.4, 10.2, 7.4, 1H), 4.99-4.86 (m, 2H), 4.86 (d, *J* = 9.3 Hz, 1H), 3.76 (dd, *J* = 8.0, 3.2 Hz, 1H), 3.74-3.69 (m, 1H), 2.41 (dq, *J* = 7.0, 3.2 Hz, 1H), 2.38-2.29 (m, 1H), 2.29-2.16 (m, 1H), 2.07-1.97 (m, 1H), 1.88-1.76 (m, 1H), 1.71-1.60 (m, 2H), 1.54 (d, *J* = 1.3 Hz, 3H), 1.49-1.40 (m, 1H), 1.32 (s, 3H), 1.32-1.20 (m, 2H), 1.30-1.24 (m, 1H), 1.29 (s, 3H), 1.20-1.08 (m, 2H), 1.14 (d, *J* = 7.1 Hz, 3H), 0.95 (d, *J* = 6.6 Hz, 3H), 0.93 (d, *J* = 6.6 Hz, 3H), 0.85 (d, *J* = 6.8 Hz, 3H), 0.79 (d, *J* = 6.3 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 204.4, 145.4, 132.6, 112.0, 100.6, 73.7, 69.4, 48.8, 47.8, 44.2, 36.2, 35.3, 33.8, 32.2, 28.4, 28.1, 24.8, 23.6, 21.3, 20.1, 19.7, 16.1, 12.0, 7.8; IR (film): 2957, 2362, 1734, 1639, 1457, 1381, 1225, 1165, 1019 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₂₅H₄₄O₃, 392.3290, found, 392.3298; [α]_D²⁰ -26.0 (*c* 1.50, CH₂Cl₂).



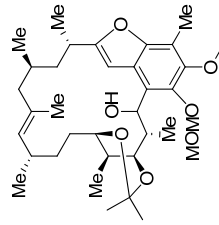
2,4-Dimethoxy-1-(methoxymethoxy)-3-methyl-5-vinylbenzene (23).

Bromide **41** (1 g, 3.43 mmol, 1.0 equiv.) was dissolved in Et₂O (34 mL, 0.1 M), cooled to -78 °C and *t*-BuLi (4 mL, 6.86 mmol, 2 equiv.) was added dropwise *via* syringe. The white suspension was stirred for 20 min at -78 °C, 15 min at 0 °C and subsequently treated with DMF (534 μL, 6.86 mmol, 2 equiv.). The solution was allowed to warm to ambient temperature over 1 h, diluted with Et₂O (70 mL) and quenched with saturated aqueous NH₄Cl (20 mL). The organic extracts were dried over MgSO₄, concentrated and purified by flash chromatography (hexane : ethylacetate from 10 : 1 to 5 : 1) to give pure aldehyde (687 mg, 83%).

To (methyl)triphenylphosphonium bromide (654 mg, 1.83 mmol, 2.2 equiv.) in THF (3.7 mL) *t*-BuOK (196 mg, 1.75 mmol, 2.1 equiv.) was added in 3 portions at 0 °C. The cloudy-orange

suspension was stirred for 45 min at room temperature, recooled to 0 °C and the above aldehyde (200 mg, 0.83 mmol, 1.0 equiv.) was added as a solution in THF (2.7 mL). The mixture was warmed to ambient temperature within 30 min, whereupon TLC analysis showed complete consumption of the starting material. The solution was diluted with 50 mL Et₂O / hexane (1 : 1), washed with saturated aqueous NH₄Cl (10 mL) and extracted with diethyl ether (3 x 10 mL). The organic layer was dried over MgSO₄ and the solvent was removed *in vacuo*. The residue was purified by column chromatography (hexane : ethylacetate from 5 : 1 to 3 : 1) to furnish arene **23** (194 mg, 98%) as an oil.

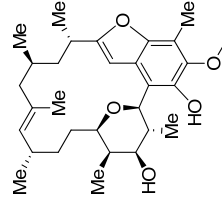
¹H-NMR (400MHz, CDCl₃): δ 7.14 (s, 1H), 6.94 (dd, *J* = 17.7, 11.1 Hz, 1H), 5.67 (dd, *J* = 17.7, 1.3 Hz, 1H), 5.25 (dd, *J* = 11.0, 1.3 Hz, 1H), 5.20 (s, 2H), 3.82 (s, 3H), 3.68 (s, 3H), 3.53 (s, 3H), 2.21 (s, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 151.5, 148.9, 146.9, 131.2, 126.4, 125.6, 114.0, 111.2, 95.6, 61.0, 60.4, 56.1, 9.3; IR (film): 2934, 2361, 1482, 1396, 1322, 1241, 1154, 1121, 1093, 1061, 1040 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₁₃H₁₈O₄, 238.1205, found, 238.1203.



Olefin **58**.

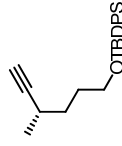
To a stirred refluxing solution of **56** (35 mg, 0.059 mmol, 1.0 equiv.) in CH₂Cl₂ (5 mL) was added portionwise dipotassium azodicarboxylate (58 mg, 0.299 mmol, 5 equiv.), followed by AcOH (9 µL, 0.150 mmol, 2.5 equiv.) in CH₂Cl₂ (1 mL). After 36 h the mixture was cooled to room temperature, filtered through silica and the filtrate was concentrated *in vacuo*. The crude product was purified by flash chromatography (hexane : ethylacetate = 10 : 1) to give **58** as a white foam (21 mg, 60%). ¹H-NMR (600MHz, CDCl₃): δ 6.27 (s, 1H), 5.16 (d, *J* = 5.7 Hz, 1H), 5.10-5.05 (m, 1H), 5.10(d, *J* = 5.7 Hz, 1H), 4.85 (d, *J* = 5.7 Hz, 1H), 4.85 (d, *J* = 9.1 Hz, 1H), 3.81-3.74 (m, 1H+OH), 3.78 (s, 3H), 3.59 (s, 3H), 3.55-3.50 (m, 1H), 3.04-2.95 (m, 1H), 2.43-2.33 (m, 1H),

2.40 (s, 3H), 2.20-2.12 (br, 1H), 2.12-2.04 (br, 1H), 2.01-1.95 (m, 1H), 1.97-1.91 (br, 1H), 1.84-1.77 (br, 1H), 1.69-1.62 (m, 1H), 1.61 (s, 3H), 1.55-1.47 (br, 1H), 1.41-1.33 (m, 1H), 1.31-1.22 (m, 1H), 1.28 (d, *J* = 6.8 Hz, 3H), 1.27-1.19 (m, 1H), 1.20 (s, 3H), 1.14 (d, *J* = 6.4 Hz, 3H), 1.07-1.01 (br, 3H), 0.99 (d, *J* = 6.8 Hz, 3H), 0.92 (d, *J* = 6.8 Hz, 3H), 0.91-0.84 (br, 3H); ¹³C-NMR (150MHz, CDCl₃): δ 164.6, 149.9, 147.6, 145.2, 132.2, 131.3, 124.6, 121.4, 119.4, 114.3, 100.3, 99.4, 98.7, 71.6, 73.0, 71.2, 60.5, 57.7, 45.9, 42.6, 40.7, 34.7, 34.1, 32.4, 31.1, 28.3, 27.8, 24.3, 20.3, 19.1, 18.8, 12.1, 10.5, 9.1; IR (film): 3435, 2930, 1654, 1559, 1490, 1458, 1378, 1248, 1158, 1109, 1001 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₃₅H₅₄O₇Na, 609.3767, found, 609.3781; [α]_D²⁰ -9.8 (c 1.0, CH₂Cl₂).



Tetrahydropyran **5**.

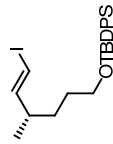
Compound **58** (20 mg, 0.034 mmol) was dissolved in MeOH (1 mL) and treated with a drop of 3N HCl. The reaction mixture was stirred for 4 h at room temperature, diluted with H₂O (10 mL) and extracted with CH₂Cl₂ (4 x 10 mL). The organic extracts were dried over MgSO₄, concentrated *in vacuo* and purified by flash chromatography (hexane : ethyl acetate from 2 : 1 to 1 : 1) to afford tetrahydropyran **5** (16 mg, 96%) as a white foam. All analytical data matched with those of the material described previously (confer conversion of **60** to **5**).



(S)-tert-Butyl(4-methylhex-5-ynyloxy)diphenylsilane (**86**).

A solution of trimethylsilyl diazomethane (2 M in Et₂O, 6.73 mL, 13.45 mmol, 1.2 equiv.) was diluted with THF (33 mL) and cooled to -78 °C. *n*-BuLi (2.5 M in hexane, 5.38 mL, 13.45 mmol, 1.2 equiv.) was added dropwise and stirring was continued for 45 min. A solution of aldehyde **36** (3.97 g, 11.21 mmol, 1.0 equiv.) in THF (11 mL) was added over a period of 20 min. After 1 h at -78 °C the reaction mixture was stirred for 30 min at 0 °C and for 1 h at ambient temperature. As TLC analysis showed complete consumption of the starting material, the mixture was partitioned between saturated aqueous NH₄Cl (100 mL) and Et₂O (100 mL). The aqueous layer was extracted with Et₂O (100 mL), the organic phase was washed with brine, dried over MgSO₄, and concentrated under reduced pressure. Flash chromatography (hexane : ethylacetate = 50 : 1) afforded 3.24 g (83%) of alkyne **86**.

¹H-NMR (400MHz, CDCl₃): δ 7.69-7.65 (m, 4H), 7.45-7.35 (m, 6H), 3.69 (t, *J* = 6.2 Hz, 2H), 2.48-2.38 (m, 1H), 2.02 (d, *J* = 2.3 Hz, 1H), 1.81-1.62 (m, 2H), 1.60-1.46 (m, 2H), 1.18 (d, *J* = 6.8 Hz, 3H), 1.05 (s, 9H); ¹³C-NMR (100MHz, CDCl₃): δ 135.6, 134.1, 129.5, 127.6, 89.0, 68.2, 63.6, 33.0, 30.2, 26.9, 25.4, 21.0, 19.2; IR (film): 2934, 2856, 1427, 1111, 889 cm⁻¹; HRMS (ESI) (*m/z*): [M-*t*Bu]⁺ calcd for C₁₉H₂₂OSi, 293.1362, found, 293.1355; [α]_D²⁰ +12.3 (c 1.1, CH₂Cl₂).

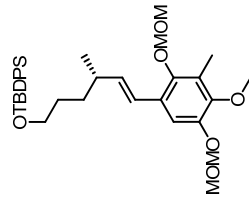


(*S,E*)-tert-Butyl(6-iodo-4-methylhex-5-enyloxy)diphenylsilane (87**).**

A 25 mL Schlenk flask was charged with Cp₂ZrHCl (207 mg, 0.804 mmol, 1.2 equiv.) followed by 2.7 mL THF (0.35 M). The white suspension was cooled in an ice bath and alkyne **86** (235 mg, 0.670 mmol) in THF (0.7 mL) was added slowly *via* cannula. The mixture was stirred for 1 h until a clear, slightly yellow solution has formed. The reaction mixture was cooled in a dry-ice-acetone bath and iodine (238 mg, 0.938 mmol, 1.4 equiv.) in THF (1 mL) was added slowly until the violet color persisted. The reaction was quenched by the addition of 1 M Na₂S₂O₃ solution. Stirring was continued until both layers became colorless. The aqueous phase was extracted with Et₂O (4 x 20 mL), the combined organic layers were washed with brine, dried over MgSO₄, filtered and evaporated to dryness. Chromatography (hexane) yielded 243 mg (76%) of vinyl

iodide **87.**

¹H-NMR (400MHz, CDCl₃): δ 7.69-7.64 (m, 4H), 7.46-7.36 (m, 6H), 6.38 (dd, *J* = 14.4, 8.1 Hz, 1H), 5.91 (dd, *J* = 14.4, 0.8 Hz, 1H), 3.64 (t, *J* = 6.3 Hz, 2H), 2.20-2.08 (m, 1H), 1.59-1.46 (m, 2H), 1.40-1.32 (m, 2H), 1.05 (s, 9H), 0.98 (d, *J* = 6.8 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 152.1, 135.6, 134.0, 129.5, 127.6, 73.4, 63.8, 40.4, 32.2, 30.0, 26.9, 19.7, 19.2; IR (film): 2931, 2857, 1684, 1653, 1559, 1507, 1472, 1428, 1112 cm⁻¹; HRMS (ESI) (*m/z*): [M-*t*Bu]⁺ calcd for C₁₉H₂₂OSi, 421.0485, found, 421.0482; [α]_D²⁰ +25.6 (c 1.4, CH₂Cl₂).

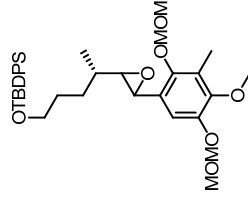


(*S,E*)-tert-Butyl(6-(4-methoxy-2,5-bis(methoxymethoxy)-3-methylphenyl)-4-methylhex-5-enyloxy)diphenylsilane (62**).**

To flame-dried anhydrous ZnCl₂ (128 mg, 0.940 mmol, 1.8 equiv.) aryl bromide **45** (302 mg, 0.940 mmol, 1.8 equiv.) in degassed Et₂O (9 mL) was added. The suspension was cooled to -78 °C and *t*-BuLi (1.7 M in pentane, 1.1 mL, 1.88 mmol, 3.6 equiv.) was added rapidly. The pale yellow solution was stirred for 30 min at -78 °C and 1 h at 0 °C. To this white suspension vinyl iodide **9** (250 mg, 0.522 mmol, 1.0 equiv.) along with Pd(PPh₃)₄ (30 mg, 0.026 mmol, 0.05 equiv.) in degassed THF (3 mL) was added slowly. The clear orange solution was stirred for 2 h at 0 °C, diluted with Et₂O (10 mL) and quenched by the addition of NH₄Cl (10 mL). The phases were separated, the organic phase was extracted with a 1 : 1 mixture of Et₂O-hexane (4 x 10 mL), washed with brine, dried over MgSO₄ and evaporated to dryness.

Purification by flash chromatography (hexane : ethylacetate = 20 : 1) afforded 270 mg **62** (208 mg, 67%, determined by ¹H-NMR) along with dehalogenated **45**. This mixture was used in the next step without further purification. A small sample was purified by HPLC to furnish analytically pure **62**.

¹H-NMR (400MHz, CDCl₃): δ 7.69-7.64 (m, 4H), 7.44-7.33 (m, 6H), 7.06 (s, 1H), 6.53 (d, *J* = 15.9 Hz, 1H), 5.96 (dd, *J* = 15.9, 8.1 Hz, 1H), 5.19 (s, 2H), 4.88 (s, 2H), 3.80 (s, 3H), 3.66 (t, *J* = 6.3 Hz, 2H), 3.57 (s, 3H), 3.53 (s, 3H), 2.33-2.22 (m, 1H), 2.23 (s, 3H), 1.64-1.52 (m, 2H), 1.49-1.39 (m, 2H), 1.07 (d, *J* = 6.8 Hz, 3H), 1.05 (s, 9H); ¹³C-NMR (100MHz, CDCl₃): δ 148.4, 148.0, 147.1, 137.3, 135.6, 134.1, 129.5, 127.6, 127.0, 126.0, 123.0, 111.3, 99.7, 95.6, 64.0, 60.5, 57.6, 56.3, 37.3, 33.1, 30.5, 26.7, 20.7, 19.2, 10.2; IR (film): 2933, 1653, 1559, 1481, 1428, 1394, 1156, 1112, 1050, 971 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₃₅H₄₈O₆Si, 592.3220, found, 592.3226; [α]_D²⁰ +24.9 (c 1.0, CH₂Cl₂).

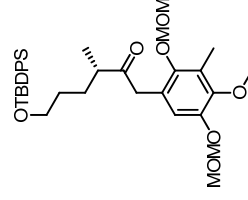


tert-Butyl((4S)-4-(3-(4-methoxy-2,5-bis(methoxymethoxy)-3-methylphenyl)oxiran-2-yl)pentyl)oxy)phenylsilane (88).

Alkene **62** (50 mg, 0.084 mmol, 1.0 equiv.) was dissolved in acetone (1 mL) and treated with DMDO (0.07 M - 0.09 M in acetone, 4 mL, 0.032 mmol, 3.8 equiv.) at ambient temperature. After stirring for 15 min, the reaction mixture was concentrated and loaded directly onto a column of silica. Flash chromatography (hexane : ethylacetate = 3 : 1) gave epoxide **88** as a pale oil (50 mg, 99%, 1.1 : 1 mixture of diastereomers).

¹H-NMR (The asterisk denotes the minor diastereomer, 400MHz, CDCl₃): δ 7.70-7.63 (m, 4H), 7.45-7.32 (m, 6H), 6.72 (s, 1H), 5.19-5.12 (m, 2H), 4.99-4.91 (m, 2H), 3.96 (d, *J* = 2.27 Hz, 1H), 3.91* (d, *J* = 2.27 Hz, 1H), 3.81 (s, 3H), 3.71-3.63 (m, 2H), 3.57* (s, 3H), 3.52 (s, 3H), 3.50 (s, 3H), 2.70-2.65 (m, 1H), 2.23* (s, 3H), 2.22 (s, 3H), 1.73-1.59 (m, 2H), 1.57-1.48 (m, 1H), 1.55-1.34 (m, 2H), 1.06* (d, *J* = 6.6 Hz, 3H), 1.05* (s, 9H), 1.04 (s, 9H), 1.00 (d, *J* = 6.6 Hz, 3H); ¹³C-NMR (The asterisk denotes the minor diastereomer, 100MHz, CDCl₃): δ 149.5, 148.5, 147.3, 135.6*, 135.5, 134.1*, 134.0, 129.5, 127.6, 126.7, 126.6*, 125.8, 110.5, 100.1, 95.5, 67.2, 67.1*, 64.1, 64.0*,

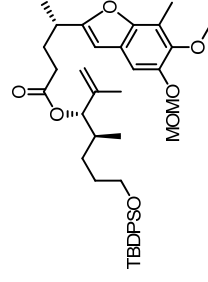
60.4, 57.5, 56.3, 54.0, 53.1*, 35.9, 35.7*, 30.5*, 30.1, 29.9, 26.9, 19.2, 16.6, 15.8*, 10.2, 10.1*; IR (film): 2933, 1734, 1559, 1486, 1429, 1394, 1360, 1238, 1156, 1112, 1050 cm⁻¹; HRMS (ESI) (*m/z*): [M+Na]⁺ calcd for C₃₅H₄₈O₇SiNa, 631.3076, found, 631.3069.



(S)-6-(tert-Butyldiphenylsilyloxy)-1-(4-methoxy-2,5-bis(methoxymethoxy)-3-methylphenyl)-3-methylhexan-2-one (48).

In a 5 mL Schlenk flask, Pd(OAc)₂ (9 mg, 0.038 mmol, 0.1 equiv.) was added to degassed *t*-BuOH (1.5 mL) followed by the addition of PBu₃ (28 μL, 0.113 mmol, 0.3 equiv.). After 5 min epoxide **88** (230 mg, 0.377 mmol, 1.0 equiv.) was added to the reaction mixture and the solution was refluxed for 1.5 h. The mixture was allowed to cool to room temperature, at which point excess *t*-BuOH was evaporated. The crude product was purified by flash chromatography (hexane : ethylacetate from 10 : 1 to 3 : 1) to yield ketone **48** as a yellow oil (185 mg, 81%).

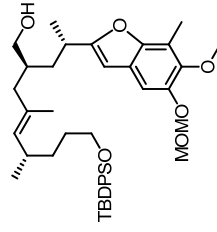
All analytical data matched with those of the material described previously (confer conversion of **44/45** to **48**).



(S)-((3S,4S)-7-(tert-Butyldiphenylsilyloxy)-2,4-dimethylhept-1-en-3-yl) 4-(6-methoxy-5-(methoxymethoxy)-7-methylbenzofuran-2-yl)pentanoate (63).

A mixture of acid **43** (1.39 g, 4.30 mmol, 1.1 equiv.), alcohol **29a** (1.55 g, 3.91 mmol, 1.0 equiv.), EDCI·HCl (1.19 g, 6.25 mmol, 1.6 equiv.) and DMAP (954 mg, 7.82 mmol, 2 equiv.) in CH₂Cl₂ (8 mL) was stirred at ambient temperature for 2 h. The reaction was diluted with CH₂Cl₂ (70 mL), quenched with 1% HCl (10 mL) and washed with brine (50 mL). The organic layer was dried over MgSO₄, concentrated and the residue was purified by flash chromatography (hexane : ethylacetate from 15 : 1 to 5 : 1) to give ester **63** (2.35 g, 86%).

¹H-NMR (400MHz, CDCl₃): δ 7.68-7.62 (m, 4H), 7.43-7.33 (m, 6H), 7.07 (s, 1H), 6.25 (s, 1H), 5.20 (s, 2H), 4.96 (d, *J* = 7.6 Hz, 1H), 4.92 (s, 2H), 3.83 (s, 3H), 3.63 (dt, *J* = 6.1, 1.9 Hz, 2H), 3.54 (s, 3H), 2.99-2.89 (m, 1H), 2.40 (s, 3H), 2.36-2.29 (m, 2H), 2.09-1.99 (m, 1H), 1.99-1.89 (m, 1H), 1.80-1.72 (m, 1H), 1.69-1.60 (m, 1H), 1.68 (s, 3H), 1.64-1.57 (m, 1H), 1.52-1.42 (m, 1H), 1.30 (d, *J* = 7.1 Hz, 3H), 1.14-1.05 (m, 1H), 1.02 (s, 9H), 0.82 (d, *J* = 6.8 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 172.7, 162.7, 149.3, 147.0, 145.7, 142.2, 135.5, 134.1, 129.5, 127.6, 123.3, 115.4, 114.0, 105.1, 101.4, 96.2, 81.6, 64.0, 61.0, 56.1, 34.2, 33.1, 32.1, 30.5, 29.9, 27.9, 26.8, 19.2, 18.9, 18.4, 15.8, 9.1; IR (film): 2932, 1734, 1559, 1457, 1153, 1113, 1044 cm⁻¹; HRMS (ESI) (*m/z*) : [M]⁺ calcd for C₄₂H₅₆O₇Si, 700.3795, found, 700.3814; [α]_D²⁰ +13.9 (c 0.90, CH₂Cl₂).



(2S,6S,E)-9-(tert-Butyldiphenylsilyloxy)-2-((S)-2-(6-methoxy-5-(methoxymethoxy)-7-methylbenzofuran-2-yl)propyl)-4,6-dimethylnon-4-en-1-ol (64).

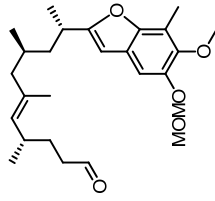
LHMDS (1 M in THF, 13.1 mL, 13.12 mmol, 4 equiv.) was cooled to -78 °C and freshly distilled HMPA (3 mL) was slowly added *via* cannula. After 5 min ester **63** (2.3 g, 3.28 mmol, 1.0 equiv.) was added as a solution in THF : HMPA = 4 : 1 (10 mL), whereupon the cloudy orange solution turned clear. After 45 min a freshly prepared TBSCl-solution (3 M in THF, 6.56 mL, 19.68 mmol, 6 equiv.) was added to the reaction mixture and stirring was continued for 20 min at -78 °C. The

solution was allowed to warm to 0 °C over 15 min, stirred for additional 5 min at ambient temperature and partitioned between H₂O (100 mL) and Et₂O (3 x 70 mL). The combined organic layers were washed with brine, dried over MgSO₄ and evaporated to dryness. The crude ketene silyl acetal was dissolved in THF (15 mL) and refluxed for 3 h. The mixture was allowed to cool to 40 °C, concentrated under reduced pressure and Et₂O (33 mL) was added. The flask was immersed in an ice-bath and LAH (4 M in Et₂O, 1.64 mL, 6.56 mmol, 2 equiv.) was added carefully *via* cannula. After 30 min at room temperature TLC analysis showed complete consumption of the starting material and the reaction mixture was quenched at 0 °C by slow addition of ethylacetate, diluted with Et₂O (50 mL) and washed with 1% HCl (100 mL). The aqueous layer was extracted with Et₂O (3 x 50 mL), the combined organic fractions were washed with brine (50 mL), dried over MgSO₄ and the solvent was removed *in vacuo*. Purification by column chromatography (hexane : ethylacetate from 10 : 1 to 3 : 1) afforded alcohol **64** as a colorless oil (1.89 g, 84%, dr = 4 : 1 as determined by ¹H-NMR).

S-64: ¹H-NMR (400MHz, CDCl₃): δ 7.70-7.62 (m, 4H), 7.44-7.33 (m, 6H), 7.08 (s, 1H), 6.28 (s, 1H), 5.21 (s, 2H), 4.95 (d, *J* = 9.1 Hz, 1H), 3.84 (s, 3H), 3.63 (t, *J* = 6.4 Hz, 2H), 3.54 (s, 3H), 3.47 (d, *J* = 5.1 Hz, 2H), 3.08-2.97 (m, 1H), 2.42 (s, 3H), 2.37-2.25 (m, 1H), 2.09-1.99 (m, 2H), 1.84 (ddd, *J* = 13.8, 8.0, 6.0 Hz, 1H), 1.76-1.66 (m, 1H), 1.57-1.43 (m, 2H), 1.55 (s, 3H), 1.49-1.40 (m, 1H), 1.42-1.33 (m, 1H), 1.31 (d, *J* = 6.8 Hz, 3H), 1.28-1.18 (m, 1H), 1.04 (s, 9H), 0.92 (d, *J* = 6.8 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 164.0, 149.2, 146.9, 145.6, 135.6, 134.1, 133.7, 132.1, 129.5, 127.5, 123.4, 115.3, 105.1, 100.9, 96.2, 66.0, 64.1, 61.0, 56.1, 42.6, 37.7, 36.1, 33.7, 32.2, 31.6, 30.6, 26.9, 21.2, 20.2, 19.2, 16.2, 9.1; IR (film): 3675, 3629, 2932, 1653, 1457, 1153, 1112, 1044 cm⁻¹; HRMS (ESI) (*m/z*) : [M]⁺ calcd for C₄₂H₅₈O₆Si, 686.4003, found, 686.3982; [α]_D²⁰ +15.8 (c 1.25, CH₂Cl₂).

R-64: ¹H-NMR (400MHz, CDCl₃): δ 7.70-7.63 (m, 4H), 7.44-7.33 (m, 6H), 7.06 (s, 1H), 6.25 (s, 1H), 5.20 (s, 2H), 4.94 (d, *J* = 9.1 Hz, 1H), 3.83 (s, 3H), 3.63 (t, *J* = 6.4 Hz, 2H), 3.57-3.51 (br, 2H), 3.54 (s, 3H), 3.11-3.00 (m, 1H), 2.40 (s, 3H), 2.36-2.26 (m, 1H), 2.06-1.93 (m, 2H), 1.78-1.66 (m, 2H), 1.61-1.51 (m, 1H), 1.58-1.46 (m, 2H), 1.50 (s, 3H), 1.41-1.33 (m, 1H), 1.29 (d, *J* = 7.07 Hz, 3H), 1.26-1.20 (m, 1H), 1.04 (s, 9H), 0.89 (d, *J* = 6.6 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 163.8, 149.1, 147.0, 145.6, 135.6, 134.2, 133.7, 132.3, 129.5, 127.5, 123.4, 115.3, 105.1, 101.0, 96.2,

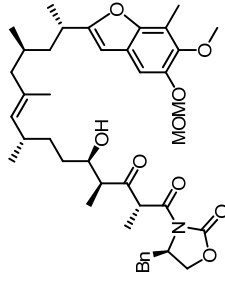
IR (film): 3629, 3422, 2928, 1654, 1606, 1458, 1340, 1219, 1153, 1116, 1044 cm^{-1} ; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{26}\text{H}_{40}\text{O}_5$, 432.2876, found, 432.2862; $[\alpha]_D^{20}$ +19.8 (c 1.0, CH_2Cl_2).



(4S,8S,10S,E)-10-(6-Methoxy-5-(methoxymethoxy)-7-methylbenzofuran-2-yl)-4,6,8-trimethylundec-5-enal (65).

Alcohol **90** (265 mg, 0.61 mmol, 1.0 equiv.) was dissolved in DMSO (3 mL, 0.2 M) and IBX (429 mg, 1.53 mmol, 2.5 equiv.) was added over a period of 20 min. After stirring for 2 h at room temperature the solution was diluted with Et_2O : hexane = 1 : 1 (30 mL) and H_2O (30 mL) to precipitate unreacted IBX. The mixture was filtered over Celite, the layers were separated and the aqueous layer was extracted with Et_2O : hexane = 1 : 1 (3 x 20 mL). The combined organic layers were dried over anhydrous MgSO_4 , filtered and concentrated *in vacuo*. The crude product was filtered over a plug of silica to give aldehyde **65** as a colorless oil (245 mg, 93%).

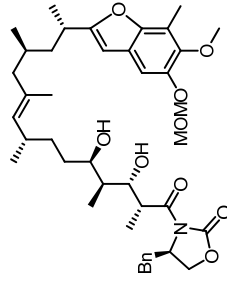
$^1\text{H-NMR}$ (600MHz, CDCl_3): δ 9.74 (t, J = 1.7 Hz, 1H), 7.07 (s, 1H), 6.24 (d, J = 0.8 Hz, 1H), 5.21 (s, 2H), 4.84 (dd, J = 9.7, 1.0 Hz, 1H), 3.84 (s, 3H), 3.54 (s, 3H), 3.03-2.96 (m, 1H), 2.42 (s, 3H), 2.42-2.34 (m, 3H), 2.07 (dd, J = 12.9, 5.9 Hz, 1H), 1.76 (dd, J = 13.4, 8.7, 1H), 1.72-1.64 (m, 2H), 1.60-1.55 (m, 1H), 1.54 (d, J = 1.1 Hz, 3H), 1.50-1.43 (m, 2H), 1.27 (d, J = 6.8 Hz, 3H), 0.96 (d, J = 6.4 Hz, 3H), 0.82 (d, J = 6.4 Hz, 3H); $^{13}\text{C-NMR}$ (150MHz, CDCl_3): δ 202.9, 164.7, 149.1, 146.9, 145.4, 133.7, 131.7, 123.5, 115.3, 104.9, 100.4, 96.1, 61.0, 56.1, 48.0, 43.0, 42.3, 32.0, 31.3, 29.7, 28.2, 21.3, 19.6, 19.2, 16.1, 9.1; IR (film): 2924, 1718, 1684, 1653, 1559, 1457, 1116 cm^{-1} ; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{26}\text{H}_{38}\text{O}_5$, 430.2719, found, 430.2705; $[\alpha]_D^{20}$ +16.4 (c 0.75, CH_2Cl_2).



(2R,4S,5R,8S,12S,14S,E)-1-((R)-4-Benzyl-2-oxoxazolidin-3-yl)-5-hydroxy-14-(6-methoxy-5-(methoxymethoxy)-7-methylbenzofuran-2-yl)-2,4,8,10,12-pentamethylpentadec-9-ene-1,3-dione (91).

A 25 mL Schlenk flask was charged with acid free $\text{Sn}(\text{OTf})_2$ (502 mg, 1.21 mmol, 1.7 equiv.) and CH_2Cl_2 (4 mL, 0.3 M). The white suspension was treated at -20°C with Et_3N (167 μL , 1.21 mmol, 1.7 equiv.) whereupon the mixture turned pale yellow. After 5 min β -keto imide **26** (338 mg, 1.17 mmol, 1.6 equiv.) in CH_2Cl_2 (2 mL, 0.6 M) was added dropwise and the clear solution was stirred for 1 h at -20°C . Aldehyde **65** (308 mg, 0.72 mmol, 1.0 equiv.) was dissolved in CH_2Cl_2 (1.2 mL, 0.6 M) and added slowly at -78°C . After 45 min at -78°C TLC analysis showed complete consumption of the starting material and the yellow-orange solution was poured onto a cooled and vigorously stirred mixture of CH_2Cl_2 and 1 M NaHSO_4 (50 mL, 1 : 1). After 20 min at room temperature the aqueous phase was extracted with CH_2Cl_2 (3 x 15 mL), the organic phase was washed with saturated aqueous NaHCO_3 , dried over MgSO_4 and concentrated *in vacuo*. Purification of the residue by gradient flash chromatography (hexane : ethylacetate from 3 : 1 to 1 : 1) yielded **91** as a viscous oil (448 mg, 87%, dr = 6 : 1 as determined by HPLC and $^1\text{H-NMR}$). $^1\text{H-NMR}$ (400MHz, CDCl_3): δ 7.36-7.27 (m, 3H), 7.22-7.17 (m, 2H), 7.07 (s, 1H), 6.24 (s, 1H), 5.21 (s, 2H), 4.88 (d, J = 8.8 Hz, 1H), 4.85 (q, J = 7.2 Hz, 1H), 4.78-4.70 (m, 1H), 4.28-4.21 (m, 1H), 4.17 (dd, J = 9.1, 3.0 Hz, 1H), 3.92-3.85 (m, 1H), 3.84 (s, 3H), 3.54 (s, 3H), 3.30 (dd, J = 13.3, 3.2 Hz, 1H), 3.05-2.94 (m, 1H), 2.81-2.72 (m, 2H), 2.45 (d, J = 3.0 Hz, OH), 2.42 (s, 3H), 2.40-2.29 (m, 1H), 2.07 (dd, J = 12.4, 5.1 Hz, 1H), 1.78 (dd, J = 12.4, 8.2 Hz, 1H), 1.71-1.63 (m, 1H), 1.60-1.44 (m, 2H), 1.54 (d, J = 1.3 Hz, 3H), 1.54-1.41 (m, 2H), 1.46 (d, J = 7.3 Hz, 3H), 1.37-1.26 (m, 2H), 1.27 (d, J = 6.8 Hz, 3H), 1.20 (d, J = 7.3 Hz, 3H), 0.94 (d, J = 6.8 Hz, 3H), 0.82 (d, J = 6.3 Hz, 3H); $^{13}\text{C-NMR}$ (100MHz, CDCl_3): δ 212.2, 170.3, 164.8, 153.6, 149.2, 146.9, 145.5, 135.0, 132.8,

132.6, 129.4, 129.0, 127.4, 123.5, 115.3, 105.0, 100.4, 96.2, 71.2, 66.5, 61.0, 56.1, 55.3, 51.9, 48.4, 48.1, 43.0, 38.0, 34.0, 32.3, 32.0, 31.3, 28.2, 21.4, 19.5, 19.2, 16.1, 12.9, 9.9, 9.1; IR (film): 3567, 2956, 1773, 1701, 1685, 1653, 1648, 1559, 1458, 1358, 1217, 1116 cm^{-1} ; HRMS (ESI) (m/z): $[M]^+$ calcd for $\text{C}_{42}\text{H}_{57}\text{O}_9\text{NNa}$, 742.3831, found, 742.3948; $[\alpha]_D^{20}$ -41.4 (c 1.0, CH_2Cl_2).

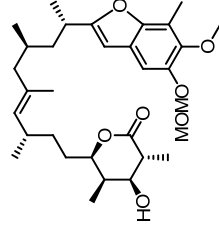


(R)-4-Benzyl-3-((2R,3S,4S,5R,8S,12S,14S,E)-3,5-dihydroxy-14-(6-methoxy-5-(methoxymethoxy)-7-methylbenzofuran-2-yl)-2,4,8,10,12-pentamethylpentadec-9-enyl)oxazolidin-2-one (92).

$\text{Me}_4\text{NBH}(\text{OAc})_3$ (73 mg, 0.278 mmol, 5 equiv.) was dissolved in 9 mL MeCN : AcOH = 1.9 : 1, cooled to -32°C and aldol product **91** (main diastereomer, 40 mg, 0.056 mmol, 1.0 equiv.) in MeCN (1 mL) was added dropwise. The reaction was stirred for 3 h at -32°C , allowed to warm to 0°C overnight, diluted with CH_2Cl_2 (20 mL) and quenched with a saturated aqueous solution of Rochelle's salt (10 mL). Saturated aqueous NaHCO_3 was carefully added to the vigorously stirred solution over 20 min. After 1 h no more gas evolution was observed and the two-phase mixture was partitioned between CH_2Cl_2 (20 mL) and H_2O (20 mL). The aqueous layer was extracted with CH_2Cl_2 (3 x 10 mL), and the combined organic fractions were dried over MgSO_4 , filtered and concentrated *in vacuo*. Purification by flash chromatography (hexane : ethylacetate from 3 : 1 to 2 : 1) afforded diol **92** as a viscous oil (29 mg, 72%, dr $\geq 20 : 1$ as determined by ^1H -NMR).

^1H -NMR (400MHz, CDCl_3): δ 7.37-7.27 (m, 3H), 7.22-7.17 (m, 2H), 7.07 (s, 1H), 6.24 (s, 1H), 5.20 (s, 2H), 4.89 (d, $J = 9.3$ Hz, 1H), 4.72-4.65 (m, 1H), 4.25-4.16 (m, 2H), 3.98-3.93 (m, 1H), 3.87 (dq, $J = 7.24, 2.4$ Hz, 1H), 3.83 (s, 3H), 3.82-3.75 (br, 1H), 3.59 (d, $J = 2.5$ Hz, OH), 3.54 (s, 3H), 3.24 (dd, $J = 13.4, 3.3$ Hz, 1H), 3.05-2.94 (m, 1H), 2.79 (dd, $J = 13.4, 9.3$ Hz, 1H), 2.55 (d, $J = 6.3$ Hz,

OH), 2.41 (s, 3H), 2.40-2.31 (m, 1H), 2.08 (dd, $J = 12.3, 5.2$ Hz, 1H), 1.87-1.78 (m, 1H), 1.75 (dd, $J = 12.6, 8.3$ Hz, 1H), 1.71-1.62 (m, 1H), 1.62-1.44 (m, 2H), 1.58-1.45 (m, 2H), 1.55 (d, $J = 1.3$ Hz, 3H), 1.49-1.29 (m, 2H), 1.27 (d, $J = 6.8$ Hz, 6H), 0.94 (d, $J = 6.6$ Hz, 3H), 0.82 (d, $J = 6.8$ Hz, 6H); ^{13}C -NMR (100MHz, CDCl_3): δ 178.1, 164.8, 152.8, 149.2, 146.9, 145.2, 134.9, 133.1, 132.4, 129.4, 129.0, 127.5, 123.5, 115.3, 105.0, 100.4, 96.2, 73.7, 73.6, 66.2, 61.0, 56.1, 55.0, 48.1, 43.1, 39.6, 39.2, 37.8, 34.5, 32.3, 31.3, 31.0, 28.2, 21.4, 19.5, 19.2, 16.1, 11.5, 10.1, 9.1; IR (film): 3678, 3452, 2925, 1783, 1700, 1559, 1455, 1386, 1217, 1153, 1116, 1043 cm^{-1} ; HRMS (ESI) (m/z): $[M]^+$ calcd for $\text{C}_{42}\text{H}_{59}\text{O}_9\text{NNa}$, 744.4088, found, 744.4098; $[\alpha]_D^{20}$ -8.4 (c 1.05, CH_2Cl_2).

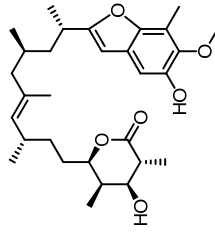


(3R,4S,5R,6R)-4-Hydroxy-6-((3S,5S,9S,E)-9-(6-methoxy-5-(methoxymethoxy)-7-methylbenzofuran-2-yl)-3,5,7-trimethyldec-4-enyl)-3,5-dimethyltetrahydro-2H-pyran-2-one (66).

A solution of diol **92** (635 mg, 0.88 mmol, 1.0 equiv.) in THF / H_2O = 3 : 1 (9 mL 0.1 M) was treated at 0°C with H_2O_2 (30% in H_2O , 352 μL). LiOH (59 mg, 1.41 mmol, 1.6 equiv.) was added in one portion and stirring was continued for 1 h at ambient temperature. The mixture was acidified with 1 N HCl (5 mL), stirred for further 5 min and then diluted with H_2O (30 mL), extracted with Et_2O (3 x 20 mL), dried over MgSO_4 and concentrated under reduced pressure. Purification by flash chromatography (hexane : ethylacetate from 3 : 1 to 1 : 1) gave lactone **66** (458 mg, 96%) as an oil.

^1H -NMR (400MHz, CDCl_3): δ 7.07 (s, 1H), 6.24 (s, 1H), 5.21 (s, 2H), 4.87 (d, $J = 9.1$ Hz, 1H), 4.14 (ddd, $J = 8.1, 5.4, 2.3$ Hz, 1H), 3.84 (s, 3H), 3.77 (dd, $J = 10.2, 4.2$ Hz, 1H), 3.54 (s, 3H), 3.06-2.95 (m, 1H), 2.48-2.40 (m, 1H), 2.42 (s, 3H), 2.40-2.31 (m, 1H), 2.11-2.02 (m, 2H), 1.94-1.84 (m, OH), 1.82-1.69 (m, 2H), 1.73-1.62 (m, 1H), 1.62-1.52 (m, 1H), 1.54 (d, $J = 1.3$ Hz, 3H), 1.52-1.43 (m,

1H), 1.46-1.35 (m, 1H), 1.42-1.29 (m, 2H), 1.38 (d, $J = 7.1$ Hz, 3H), 1.27 (d, $J = 7.1$ Hz, 3H), 0.94 (d, $J = 6.6$ Hz, 3H), 0.92 (d, $J = 6.6$ Hz, 3H), 0.82 (d, $J = 6.3$ Hz, 3H); $^{13}\text{C-NMR}$ (100MHz, CDCl_3): δ 173.6, 164.7, 149.2, 146.8, 145.5, 132.9, 132.3, 123.5, 115.3, 105.0, 100.4, 96.2, 79.9, 73.9, 61.0, 56.1, 47.9, 43.0, 39.8, 37.3, 33.1, 32.1, 31.3, 30.2, 28.2, 21.3, 19.6, 19.2, 16.2, 14.2, 9.1, 4.4; IR (film): 3454, 2927, 1733, 1700, 1559, 1456, 1219, 1153, 1115, 1043 cm^{-1} ; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{32}\text{H}_{48}\text{O}_5\text{Na}$, 567.3298, found, 567.3310; $[\alpha]_D^{20} + 50.7$ (c 0.55, CH_2Cl_2).

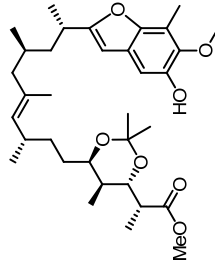


(3R,4S,5R,6R)-4-Hydroxy-6-((3S,7S,9S,E)-9-(5-hydroxy-6-methoxy-7-methylbenzofuran-2-yl)-3,5,7-trimethyldec-4-enyl)-3,5-dimethyltetrahydro-2H-pyran-2-one (93).

Lactone **66** (93 mg, 0.170 mmol, 1.0 equiv.) in dioxane (2 mL) was heated to 50 °C and 2 drops of 3 N HCl were added. After stirring for 2 h the reaction mixture was poured onto H_2O (10 mL), extracted with Et_2O (4 x 20 mL), washed with brine (15 mL), dried over MgSO_4 and concentrated. For analytical purposes a small sample of crude phenol **93** was purified by flash chromatography (hexane : ethylacetate = 3 : 1).

$^1\text{H-NMR}$ (400MHz, CDCl_3): δ 6.86 (s, 1H), 6.23 (s, 1H), 5.46 (s, OH), 4.87 (d, $J = 9.1$ Hz, 1H), 4.15 (ddd, $J = 8.4, 5.4, 2.6$ Hz, 1H), 3.83 (s, 3H), 3.82-3.75 (m, 1H), 3.04-2.94 (m, 1H), 2.47-2.40 (m, 1H), 2.45 (s, 3H), 2.40-2.30 (m, 1H), 2.11-2.02 (m, 2H), 1.81-1.70 (m, 2H+OH), 1.70-1.62 (m, 1H), 1.62-1.53 (m, 1H), 1.54 (d, $J = 1.3$ Hz, 3H), 1.51-1.42 (m, 1H), 1.47-1.36 (m, 1H), 1.42-1.29 (m, 2H), 1.39 (d, $J = 7.3$ Hz, 3H), 1.27 (d, $J = 6.8$ Hz, 3H), 0.94 (d, $J = 6.3$ Hz, 3H), 0.93 (d, $J = 7.1$ Hz, 3H), 0.82 (d, $J = 6.3$ Hz, 3H); $^{13}\text{C-NMR}$ (100MHz, CDCl_3): δ 173.6, 164.7, 147.8, 145.1, 142.8, 133.0, 132.3, 123.7, 113.7, 101.9, 100.3, 79.9, 74.0, 61.5, 47.9, 43.0, 39.8, 37.3, 33.2, 32.2, 31.3, 30.3, 28.2, 21.3, 19.5, 19.1, 16.1, 14.2, 9.4, 4.4; IR (film): 3443, 2925, 1718, 1653, 1609, 1559, 1507, 1457, 1380, 1217, 1111 cm^{-1} ; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{30}\text{H}_{44}\text{O}_6\text{Na}$, 523.3036,

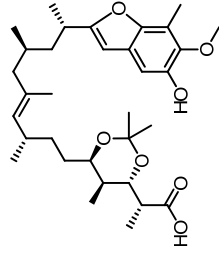
found, 523.3030; $[\alpha]_D^{20} + 55.1$ (c 1.40, CH_2Cl_2).



(R)-Methyl 2-((4S,5S,6R)-6-((3S,7S,9S,E)-9-(5-hydroxy-6-methoxy-7-methylbenzofuran-2-yl)-3,5,7-trimethyldec-4-enyl)-2,2,5-trimethyl-1,3-dioxan-4-yl)propanoate (94).

Phenol **93** was dissolved in 2,2-dimethoxypropane (2 mL), treated with camphorsulfonic acid (4 mg, 0.017 mmol, 0.1 equiv.) and stirred overnight at room temperature. The solution was diluted with Et_2O (10 mL), neutralized with saturated aqueous NaHCO_3 (10 mL), extracted with Et_2O (3 x 10 mL), washed with brine (10 mL) and dried over MgSO_4 . The solvent was removed *in vacuo* and the residue was purified by flash chromatography (hexane : ethylacetate = 5 : 1) to yield ester **94** (82 mg, 85% over 2 steps).

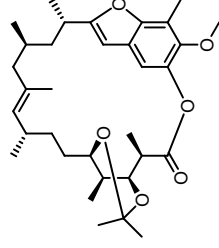
$^1\text{H-NMR}$ (400MHz, CDCl_3): δ 6.87 (s, 1H), 6.23 (s, 1H), 5.43 (s, OH), 4.89 (d, $J = 9.3$ Hz, 1H), 3.83 (s, 3H), 3.75-3.68 (m, 1H), 3.68 (s, 3H), 3.61 (dd, $J = 7.6, 5.1$ Hz, 1H), 3.04-2.94 (m, 1H), 2.60-2.51 (m, 1H), 2.45 (s, 3H), 2.41-2.27 (m, 1H), 2.06 (dd, $J = 12.5, 4.9$ Hz, 1H), 1.82-1.75 (m, 1H), 1.81-1.73 (m, 1H), 1.72-1.64 (m, 1H), 1.61-1.44 (m, 2H), 1.55 (d, $J = 1.3$ Hz, 3H), 1.48-1.36 (m, 1H), 1.33-1.21 (m, 2H), 1.30 (s, 3H), 1.29 (s, 3H), 1.27 (d, $J = 7.1$ Hz, 3H), 1.26-1.19 (m, 1H), 1.18 (d, $J = 7.1$ Hz, 3H), 0.93 (d, $J = 6.6$ Hz, 3H), 0.83 (d, $J = 6.3$ Hz, 3H), 0.80 (d, $J = 6.8$ Hz, 3H); $^{13}\text{C-NMR}$ (100MHz, CDCl_3): δ 174.9, 164.8, 147.5, 145.1, 142.5, 133.0, 132.3, 124.2, 113.8, 101.9, 100.5, 100.3, 75.4, 69.3, 61.5, 51.6, 48.1, 43.0 (2C), 36.8, 33.8, 32.2, 31.3, 28.5, 28.2, 25.0, 23.7, 21.3, 19.5, 19.1, 16.1, 11.9, 11.4, 9.4; IR (film): 3445, 2954, 1738, 1609, 1456, 1380, 1225, 1111, 1021 cm^{-1} ; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{34}\text{H}_{52}\text{O}_7\text{Na}$, 595.3611 found, 595.3607; $[\alpha]_D^{20} - 3.0$ (c 0.50, CH_2Cl_2).



(R)-2-((4S,5S,6R)-6-((3S,7S,9S,E)-9-(5-Hydroxy-6-methoxy-7-methylbenzofuran-2-yl)-3,5,7-trimethyldec-4-enyl)-2,2,5-trimethyl-1,3-dioxan-4-yl)propanoic acid (95).

Ester **94** (69 mg, 0.12 mmol, 1.0 equiv.) was dissolved in THF-MeOH-H₂O (1 mL, 2 : 2 : 1), treated with LiOH (51 mg, 1.21 mmol, 10 equiv.) and stirred for 16 h at room temperature. The mixture was diluted with H₂O (10 mL), acidified to pH 5 with 1% HCl, extracted with CH₂Cl₂ (4 x 10 mL) and dried over MgSO₄. The solvent was evaporated under reduced pressure and the crude material was purified by flash chromatography (hexane : ethylacetate from 2 : 1 to 1 : 1) to afford *seco*-acid **95** (58 mg, 84%).

¹H-NMR (400MHz, CDCl₃): δ 6.87 (s, 1H), 6.23 (s, 1H), 4.87 (d, *J* = 9.3 Hz, 1H), 3.83 (s, 3H), 3.74-3.68 (m, 1H), 3.64 (dd, *J* = 7.6, 3.8 Hz, 1H), 3.04-2.94 (m, 1H), 2.68 (dq, *J* = 7.2, 3.9 Hz, 1H), 2.45 (s, 3H), 2.40-2.29 (m, 1H), 2.06 (dd, *J* = 12.4, 5.6 Hz, 1H), 1.80-1.72 (m, 2H), 1.72-1.63 (m, 1H), 1.61-1.44 (m, 2H), 1.54 (d, *J* = 1.0 Hz, 3H), 1.46-1.38 (m, 1H), 1.36-1.19 (m, 2H), 1.36 (s, 6H), 1.30-1.21 (m, 1H), 1.27 (d, *J* = 7.3 Hz, 3H), 1.21 (d, *J* = 7.3 Hz, 3H), 0.93 (d, *J* = 6.6 Hz, 3H), 0.85 (d, *J* = 6.8 Hz, 3H), 0.82 (d, *J* = 6.3 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 175.6, 164.8, 147.8, 145.1, 142.4, 132.8, 132.4, 124.2, 113.8, 102.0, 101.2, 100.3, 75.3, 69.6, 61.4, 48.0, 43.0 42.7, 36.4, 33.7, 32.3, 31.3, 28.3, 28.2, 25.0, 23.8, 21.3, 19.5, 19.1, 16.1, 12.4, 11.2, 9.4; IR (film): 3312, 2976, 2931, 1709, 1653, 1608, 1457, 1419, 1381, 1225, 1111, 1020 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₃₃H₅₀O₇Na, 581.3454 found, 581.3451; [α]_D²⁰ -6.3 (c 1.00, CH₂Cl₂).



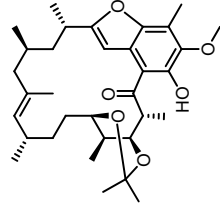
Macrolactone 67.

A 50 mL Schlenk flask was charged with EDCI·HCl (14 mg, 0.072 mmol, 2 equiv.), DMAP (13 mg, 0.107 mmol, 3 equiv.), and DMAP·HCl (11 mg, 0.072 mmol, 2 equiv.). Ethanol-free CHCl₃ (25 mL) was added, the mixture was heated to reflux and *seco*-acid **95** (20 mg, 0.036 mmol, 1.0 equiv.) in CHCl₃ (4 mL) was added *via* syringe pump over 20 h. The reaction was cooled to room temperature and quenched with saturated aqueous NH₄Cl (40 mL). The layers were separated, the aqueous phase was extracted with CH₂Cl₂ (3 x 10 mL), and the combined organic extracts were dried over MgSO₄, filtered through a short pad of silica gel, and concentrated *in vacuo*. Flash column chromatography (hexane : ethylacetate from 20 : 1 to 10 : 1) provided macrolactone **67** along with dimer **67'** as yellow viscous oils (14 mg, 73%, 3 : 1).

67: ¹H-NMR (400MHz, CDCl₃): δ 7.04 (s, 1H), 6.20 (s, 1H), 4.42 (d, *J* = 8.8 Hz, 1H), 3.80 (s, 3H), 3.80-3.74 (m, 1H), 3.56 (t, *J* = 4.9 Hz, 1H), 3.17-3.07 (m, 1H), 3.05 (dq, *J* = 7.1, 4.5 Hz, 1H), 2.45 (s, 3H), 2.30-2.17 (m, 1H), 1.97 (d, *J* = 14.7 Hz, 1H), 1.94-1.86 (m, 1H), 1.63-1.54 (m, 1H), 1.52-1.42 (m, 1H), 1.51 (s, 3H), 1.49-1.23 (m, 2H), 1.44 (s, 3H), 1.41 (s, 3H), 1.41-1.30 (m, 1H), 1.38 (d, *J* = 6.8 Hz, 3H), 1.37-1.21 (m, 1H), 1.33 (d, *J* = 7.3 Hz, 3H), 1.02-0.80 (m, 2H), 0.88 (d, *J* = 6.8 Hz, 3H), 0.82 (d, *J* = 6.3 Hz, 3H), 0.79 (d, *J* = 6.6 Hz, 3H); ¹³C-NMR (100MHz, CDCl₃): δ 172.6, 162.8, 151.4, 146.7, 140.2, 131.4, 129.2, 123.5, 115.5, 110.0, 102.7, 100.6, 78.2, 69.6, 61.5, 43.9, 42.7, 42.4, 33.7, 33.5, 32.0, 31.4, 28.1, 27.9, 26.8, 24.3, 21.5, 20.1, 19.1, 18.9, 14.2, 12.9, 9.2; IR (film): 2930, 1760, 1604, 1456, 1418, 1379, 1226, 1195, 1111, 1024, 1001 cm⁻¹; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₃₃H₄₈O₆Na, 563.3349 found, 563.3352; [α]_D²⁰ -41.7 (c 0.35, CH₂Cl₂).

67' ¹H-NMR (400MHz, CDCl₃): δ 6.92 (s, 1H), 6.25 (s, 1H), 4.88 (d, *J* = 9.3 Hz, 1H), 3.84-3.78 (m, 1H), 3.74 (s, 3H), 3.68 (t, *J* = 6.7 Hz, 1H), 3.04-2.94 (m, 1H), 2.93-2.85 (m, 1H), 2.43 (s, 3H), 2.41-2.30 (m, 1H), 2.05 (dd, *J* = 13.1, 5.6 Hz, 1H), 1.98-1.89 (m, 1H), 1.76 (dd, *J* = 13.3, 8.0 Hz, 1H),

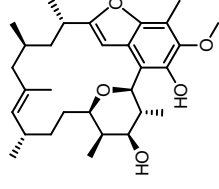
1.69-1.60 (m, 1H), 1.61-1.45 (m, 2H), 1.53 (d, $J = 1.0$ Hz, 3H), 1.51-1.21 (m, 2H), 1.37 (s, 3H), 1.36 (d, $J = 6.8$ Hz, 3H), 1.33 (s, 3H), 1.33-1.23 (m, 2H), 1.26 (d, $J = 6.8$ Hz, 3H), 0.93 (d, $J = 6.6$ Hz, 3H), 0.90 (d, $J = 6.8$ Hz, 3H), 0.82 (d, $J = 6.6$ Hz, 3H); $^{13}\text{C-NMR}$ (100MHz, CDCl_3): δ 173.2, 165.3, 151.4, 146.4, 140.0, 132.8, 132.2, 130.1, 123.8, 115.5, 110.4, 100.4, 76.1, 69.1, 61.4, 47.5, 44.2, 42.9, 36.9, 33.7, 32.4, 31.4, 28.6 (2C), 25.5, 24.2, 21.4, 20.0, 19.8, 16.6, 13.1, 12.5, 9.3; IR (film): 2930, 1762, 1601, 1457, 1418, 1379, 1226, 1195, 1111, 1024, 1001 cm^{-1} ; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{46}\text{H}_{96}\text{O}_{12}\text{Na}$, 1103.6800 found, 1103.6770; $[\alpha]_D^{20}$ -36.6 (c 0.50, CH_2Cl_2).

**Ketone 68.**

Macrolactone **67** (4.0 mg, 7.4 μmol) in cyclohexane (10 mL) was irradiated with a Hannover 700W medium pressure mercury lamp with a quartz filter at room temperature for 50 min. The intensely yellow solution was concentrated and the crude residue was subjected to flash column chromatography (hexane : ethylacetate = 10 : 1) to furnish ketone **68** as a yellow oil (3.1 mg, 75%).

$^1\text{H-NMR}$ (400MHz, CDCl_3): δ 13.73 (s, 1H), 6.77 (s, 1H), 4.67 (d, $J = 9.1$ Hz, 1H), 3.91 (s, 3H), 3.90-3.81 (m, 2H), 3.60 (dd, $J = 8.5, 6.7$ Hz, 1H), 3.14-3.04 (m, 1H), 2.54-2.44 (m, 1H), 2.50 (s, 3H), 2.30 (d, $J = 14.4$ Hz, 1H), 1.86-1.77 (m, 1H), 1.70 (d, $J = 1.0$ Hz, 3H), 1.64-1.56 (m, 1H), 1.50-1.42 (m, 1H), 1.49 (dd, $J = 9.3, 5.6$ Hz, 1H), 1.49-1.21 (m, 1H), 1.44-1.34 (m, 1H), 1.39-1.17 (m, 2H), 1.37 (s, 3H), 1.36 (d, $J = 6.3$ Hz, 3H), 1.35 (s, 3H), 1.34 (d, $J = 6.3$ Hz, 3H), 1.17-1.07 (m, 1H), 0.83 (d, $J = 6.8$ Hz, 3H), 0.82 (d, $J = 6.1$ Hz, 3H), 0.71 (d, $J = 6.8$ Hz, 3H); $^{13}\text{C-NMR}$ (100MHz, CDCl_3): δ 208.1, 163.7, 156.3, 147.2, 143.6, 132.5, 129.2, 125.3, 121.9, 110.6, 103.8, 100.3, 78.2, 68.4, 60.6, 47.3, 43.8, 43.6, 37.0, 32.1, 31.7, 31.3, 30.3, 26.9, 25.9, 24.4, 21.5, 20.9, 20.0, 19.5, 15.8, 12.5, 9.9; IR (film): 2930, 1613, 1455, 1379, 1313, 1265, 1162, 1124, 1083 cm^{-1} ; HRMS (ESI) (m/z):

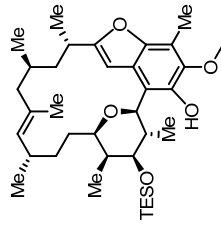
$[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{33}\text{H}_{48}\text{O}_8\text{Na}$, 563.3349 found, 563.3355; $[\alpha]_D^{20}$ -58.7 (c 0.30, CH_2Cl_2).

**Tetrahydropyran 5.**

Ketone **68** (3.0 mg, 5.5 μmol , 1.0 equiv.) was dissolved in MeOH (1 mL) and NaBH_4 (0.8 mg, 22.2 μmol , 4 equiv.) was added in one portion, whereupon the yellow solution immediately turned colorless. After 10 min, the reaction was quenched by the dropwise addition of 0.5 N HCl (200 μL) and stirring was continued at room temperature for 30 min. H_2O (10 mL) was added and the aqueous layer was extracted with CH_2Cl_2 (4 x 10 mL). The combined organic extracts were dried over MgSO_4 , filtered and evaporated to dryness. The residue was dried by azeotropic distillation with toluene (2 x 5 mL) and used without further purification.

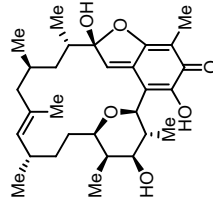
The crude product was dissolved in toluene (1 mL) and a catalytical amount of *p*-toluenesulfonic acid was added. The flask was capped and the solution was heated to 60 °C for 30 min. The mixture was allowed to cool to room temperature and was directly loaded onto a silica column and purified by flash chromatography (hexane : ethylacetate from 2 : 1 to 1 : 1) gave tetrahydropyran **5** (1.9 mg, 71%) as a white foam. All analytical data matched with those of the material described previously (confer conversion of **60** to **5**).²

² Y. Yu, H. Men, C. Lee, *J. Am. Chem. Soc.* **2004**, 126, 14720.

**TES Ether 96.**

Tetrahydropyran **5** (18 mg, 0.037 mmol, 1.0 equiv.) in CH_2Cl_2 (4 mL) was cooled to 0 °C and Et_3N (36 μL , 0.260, 7 equiv.) was added. After 5 min TESOTf (10 μL , 0.045 mmol, 1.2 equiv.) in CH_2Cl_2 (1 mL) was added to the solution *via* cannula. The mixture was allowed to warm to room temperature overnight, and was quenched with aqueous saturated NaHCO_3 (20 μL) and concentrated under reduced pressure. Purification by flash chromatography (hexane : ethylacetate from 20 : 1 to 10 : 1) afforded pure TES-ether **96** (18mg, 82%). The analytical data matched those reported by Lee.²

$^1\text{H-NMR}$ (600MHz, CDCl_3): δ 6.55 (s, 1H), 5.51 (s, 1H), 4.60 (d, J = 9.4 Hz, 1H), 4.52 (d, J = 10.2 Hz, 1H), 3.82 (s, 3H), 3.62 (d, J = 10.0, 4.7 Hz, 1H), 3.40 (d, J = 11.0, 1H), 3.10-3.02 (m, 1H), 2.48-2.40 (m, 1H), 2.45 (s, 3H), 2.26-2.18 (m, 1H), 1.87-1.79 (m, 1H), 1.78-1.73 (m, 1H), 1.62 (s, 3H), 1.61-1.54 (m, 1H), 1.47-1.39 (m, 2H), 1.37 (d, J = 6.8 Hz, 3H), 1.36-1.17 (m, 5H), 1.03 (d, J = 6.8 Hz, 3H), 0.98 (t, J = 7.9 Hz, 9H), 0.82 (d, J = 6.8 Hz, 3H), 0.81 (d, J = 6.7 Hz, 3H), 0.76 (d, J = 6.5 Hz, 3H), 0.63 (q, J = 7.9 Hz, 6H); $^{13}\text{C-NMR}$ (150MHz, CDCl_3): δ 159.6, 148.2, 141.6, 141.4, 131.5, 129.0, 122.2, 116.2, 112.3, 104.9, 77.7, 77.5, 61.4, 43.8, 41.8, 40.8, 38.7, 33.7, 32.6, 31.5, 31.3, 27.5, 21.8, 21.0, 19.6, 18.7, 13.3, 9.4, 7.0, 6.9, 5.1; IR (film): 2954, 1456, 1404, 1383, 1325, 1107, 1081 cm^{-1} ; HRMS (ESI) (m/z): $[\text{M}]^+$ calcd for $\text{C}_{36}\text{H}_{58}\text{O}_5\text{SiNa}$, 621.3951, found, 621.3943; $[\alpha]_D^{23}$ +37.2 (c 1.00, CHCl_3).

**Kendomycin (1).**

TES-ether **96** (17mg, 0.028 mmol, 1.0 equiv.) was dissolved in degassed DMF (3 mL), and IBX (24 mg, 0.085 mmol, 3 equiv.) was added at room temperature, whereupon the solution turned orange. After stirred for 24 h in the dark, TLC analysis showed complete consumption of the starting material. The purple solution was directly loaded onto a silica column and eluted (hexane : ethylacetate = 10 : 1) to afford the o-quinone as a purple-blue oil, which was dissolved in MeCN (3 mL) and treated with aqueous HF (0.1 M, 369 μL , 0.037 mmol, 1.3 equiv.). The initial blue solution turned yellow within 4 h and the reaction mixture was partitioned between ethylacetate (50 mL) and saturated aqueous NaHCO_3 (10 mL). The aqueous phase was extracted with ethylacetate (3 x 10 mL), and the organic layer was washed with brine, dried over MgSO_4 and concentrated *in vacuo*. Purification by flash chromatography (hexane : ethylacetate = 2 : 1) gave kendomycin **1** (4 mg, 29% over 2 steps) as a yellow solid. All analytical data matched with those of the material described previously (confer conversion of **71** to **1**).

Comparison of the ^1H - and ^{13}C -NMR Data of Natural and Synthetic (-)-Kendomycin 1.

| ^1H -NMR (600 MHz, CD_3COCD_3) | | ^{13}C -NMR (600 MHz, CD_3COCD_3) | |
|--|--|---|----------------------------|
| Natural | Synthetic | Natural | Synthetic |
| 8.10 (s, 4-OH) | 8.10 (s, 4-OH) | 182.1 (C-3) | 182.1 (C-3) |
| 7.19 (s, 20-H) | 7.19 (s, 20-H) | 168.6 (C-1) | 168.6 (C-1) |
| 6.54 (s, 19-OH) | 6.54 (s, 19-OH) | 146.8 (C-4) | 146.8 (C-4) |
| 4.64 (d, J = 10.2 Hz, 13-H) | 4.64 (d, J = 10.0 Hz, 13-H) | 141.3 (C-20) | 141.3 (C-20) |
| 4.36 (d, J = 10.4 Hz, 5-H) | 4.35 (d, J = 10.3 Hz, 5-H) | 132.1 (C-14) | 132.1 (C-14) |
| 3.95 (d, J = 4.5 Hz, 7-OH) | 3.95 (d, J = 4.5 Hz, 7-OH) | 130.2 (C-20a) | 130.2 (C-20a) |
| 3.56 (m, 7-H) | 3.56 (m, 7-H) | 129.9 (C-13) | 129.9 (C-13) |
| 3.53 (ddd, J = 10.8, 2.5, 1.1 Hz, 9-H) | 3.53 (ddd, J = 11.0, 2.3, 1.1 Hz, 9-H) | 119.1 (C-19) | 119.1 (C-19) |
| 2.41 (m, 18-H) | 2.42 (m, 18-H) | 111.0 (C-4a) | 111.0 (C-4a) |
| 2.36 (m, 12-H) | 2.36 (m, 12-H) | 104.2 (C-2) | 104.2 (C-2) |
| 2.12 (br d, J = 17.1 Hz, 15-H ^a) | 2.12 (br d, J = 17.0 Hz, 15-H ^a) | 78.7 (C-9) | 78.7 (C-9) |
| 1.96 (m, 16-H) | 1.96 (m, 16-H) | 77.7 (C-5) | 77.8 (C-5) |
| 1.88 (m, 8-H) | 1.88 (m, 8-H) | 76.2 (C-7) | 76.2 (C-7) |
| 1.84 (s, 2-CH ₃) | 1.84 (s, 2-CH ₃) | 46.1 (C-15) | 46.1 (C-15) |
| 1.70 (m, 6-H) | 1.71 (m, 6-H) | 41.4 (C-18) | 41.4 (C-18) |
| 1.68 (m, 15-H ^b) | 1.67 (m, 15-H ^b) | 40.8 (C-8) | 40.8 (C-8) |
| 1.63 (m, 17-H ^a) | 1.64 (m, 17-H ^a) | 39.7 (C-17) | 39.8 (C-17) |
| 1.61 (s, 14-CH ₃) | 1.61 (s, 14-CH ₃) | 38.1 (C-6) | 38.1 (C-6) |
| 1.57 (m, 10-H ^a) | 1.57 (m, 10-H ^a) | 35.8 (C-11) | 35.9 (C-11) |
| 1.45 (ddd, J = 13.1, 11.2, 3.0 Hz, 17-H ^b) | 1.45 (ddd, J = 12.9, 11.4, 2.9 Hz, 17-H ^b) | 33.4 (C-12) | 33.6 (C-12) |
| 1.32 (m, 11-H ₂) | 1.32 (m, 11-H ₂) | 33.5 (C-10) | 33.5 (C-10) |
| 1.25 (m, 10-H ^b) | 1.25 (m, 10-H ^b) | 26.5 (C-16) | 26.5 (C-16) |
| 0.95 (d, J = 7.0 Hz, 8-CH ₃) | 0.95 (d, J = 7.0 Hz, 8-CH ₃) | 22.7 (C-12) | 22.7 (C-12) |
| 0.94 (d, J = 6.5 Hz, 16-CH ₃) | 0.94 (d, J = 6.5 Hz, 16-CH ₃) | 19.9 (14-CH ₃) | 19.9 (14-CH ₃) |
| 0.89 (d, J = 6.5 Hz, 6-CH ₃) | 0.89 (d, J = 6.5 Hz, 6-CH ₃) | 19.7 (16-CH ₃) | 19.7 (16-CH ₃) |
| 0.87 (d, J = 6.5 Hz, 12-CH ₃) | 0.87 (d, J = 6.5 Hz, 12-CH ₃) | 13.3 (6-CH ₃) | 13.3 (6-CH ₃) |
| 0.71 (d, J = 7.0 Hz, 18-CH ₃) | 0.71 (d, J = 7.0 Hz, 18-CH ₃) | 12.6 (18-CH ₃) | 12.7 (18-CH ₃) |
| | | 7.6 (2-CH ₃) | 7.6 (2-CH ₃) |
| | | 7.2 (8-CH ₃) | 7.2 (8-CH ₃) |

**Ring Closing Metathesis and Photo-Fries Reaction
for the Construction of the Ansamycin Antibiotic
Kendomycin. Development of a Protecting Group
Free Oxidative Endgame.**

SUPPORTING INFORMATION (^1H and ^{13}C NMR)

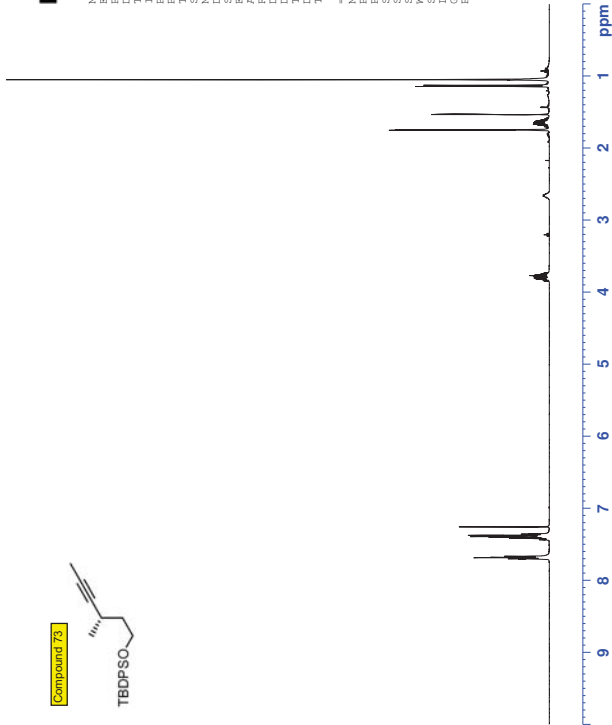
Thomas Magauer, Harry J. Martin and Johann Mulzer*

Institute of Organic Chemistry, University of Vienna
Währingerstraße 38
1090 Vienna
Austria

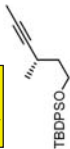


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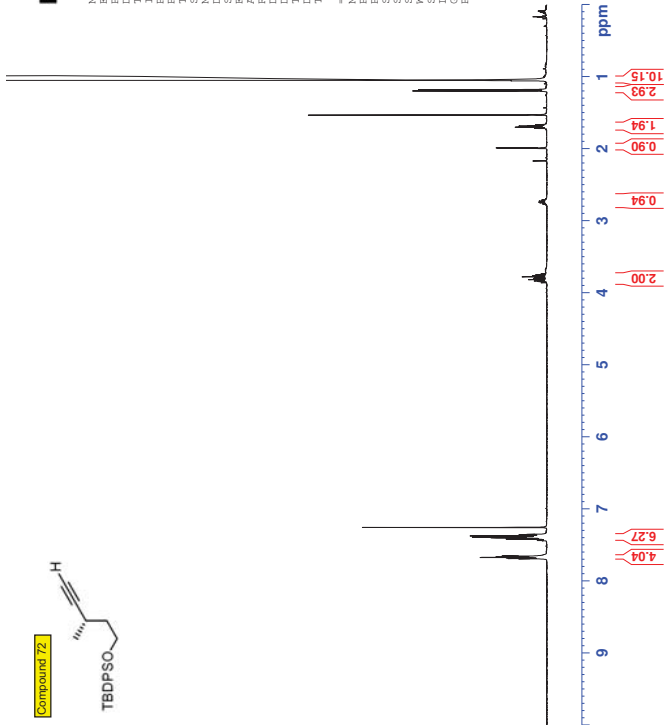


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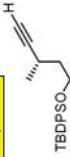


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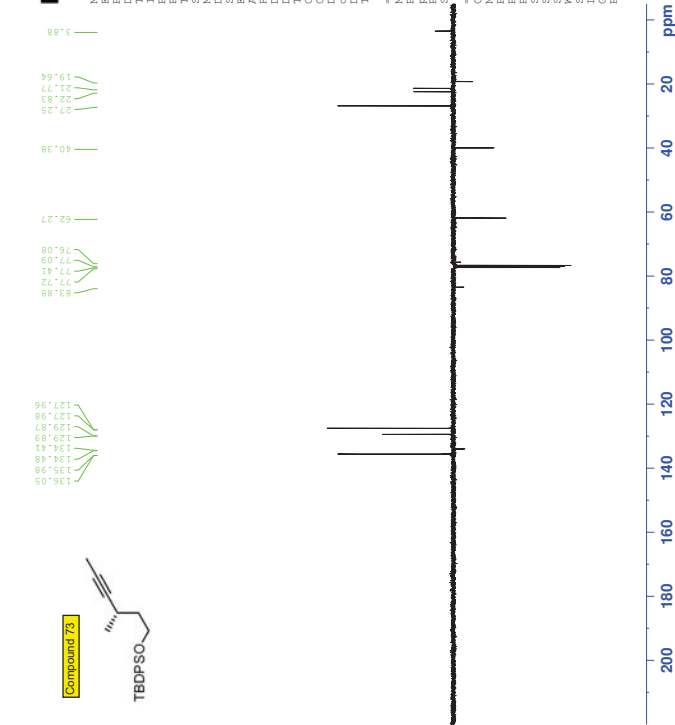
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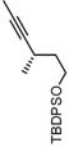
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TD0: 1



Compound 73



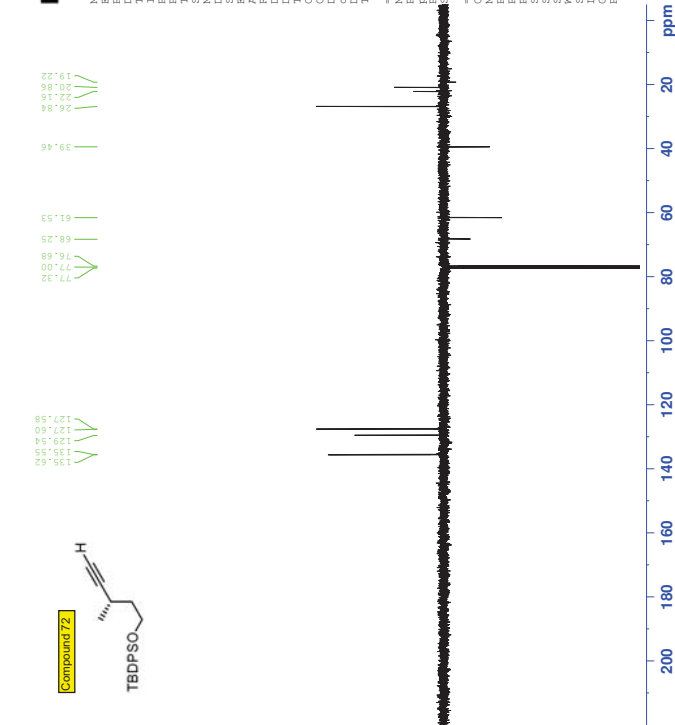
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DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.3074932 sec
RG: 103.923
DW: 19.950 usec
DE: 26.00 usec
TE: 300.2 K
D1: 1.00000000 sec
D20: 0.00689955 sec
DELTA: 0.00001311 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 13C
P1: 10.13 usec
PL1: -1.00 dB
SF01: 100.6233229 MHz

===== CHANNEL f2 =====
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NS: 1000
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.3074932 sec
RG: 103.923
DW: 19.950 usec
DE: 26.00 usec
TE: 300.2 K
D1: 1.00000000 sec
D20: 0.00689955 sec
DELTA: 0.00001311 sec
TD0: 1

===== CHANNEL f1 =====
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SF01: 100.6233229 MHz

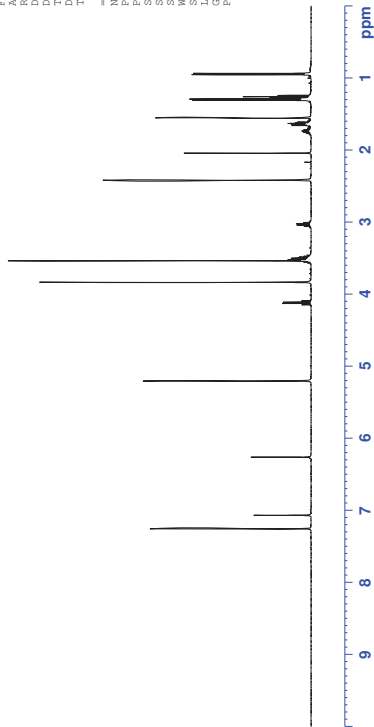
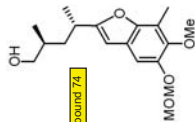
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EXPNO: 1
PROCNO: 1
Date_: 20070419
Time: 15.00
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1000
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.3074932 sec
RG: 103.923
DW: 19.950 usec
DE: 26.00 usec
TE: 300.2 K
D1: 1.00000000 sec
D20: 0.00689955 sec
DELTA: 0.00001311 sec
TD0: 1





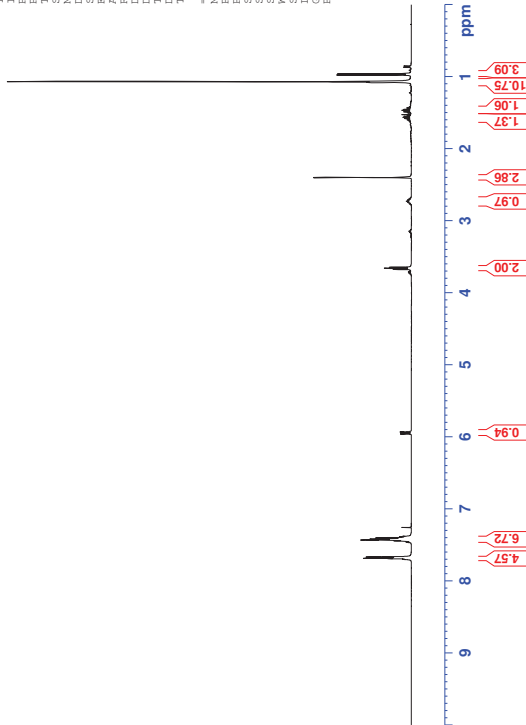
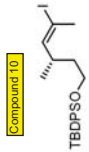
NAME: TIRIA 054
EXPNO: 7
PROCNO: 1
Date_: 20070412
Time: 22:21
INSTRUM: avance400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 1H
P1: 8.75 usec
PL1: -2.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: HANN
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00



NAME: tirma100
EXPNO: 1
PROCNO: 1
Date_: 20070424
Time: 22:07
INSTRUM: avance400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 1H
P1: 8.75 usec
PL1: -2.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: HANN
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00



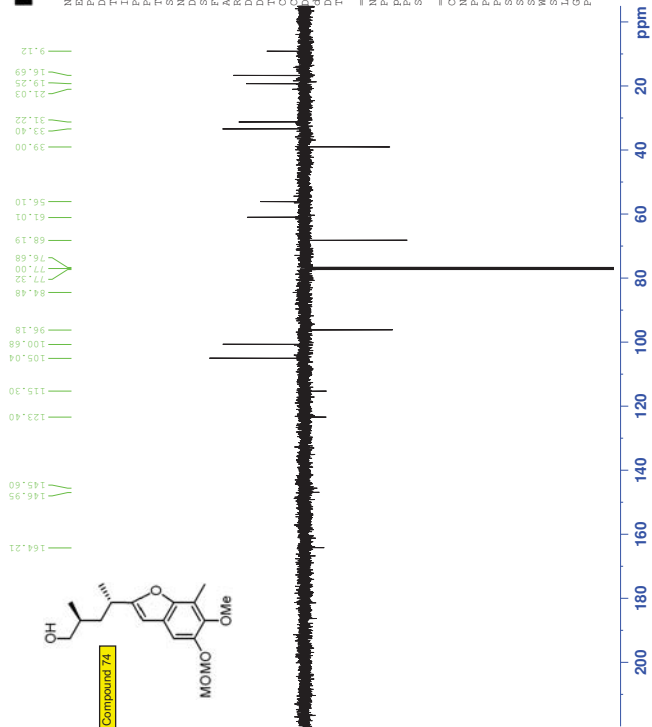
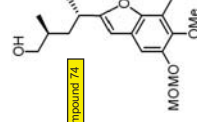
NAME: TIRIA 054
EXPNO: 7
PROCNO: 1
Date_: 20070412
Time: 22:21
INSTRUM: avance400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.0068955 sec
DELTA: 0.0001311 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 13C
P1: 13.00 usec
PL1: -1.00 dB
SF01: 100.6233329 MHz

===== CHANNEL f2 =====
NAME: waltz16
EXPNO: 1
PROCNO: 1
Date_: 20070412
Time: 22:21
INSTRUM: avance400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1000
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307972 sec
RG: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.0068955 sec
DELTA: 0.0001311 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 13C
P1: 13.00 usec
PL1: -1.00 dB
SF01: 100.6233329 MHz

===== CHANNEL f2 =====
NAME: waltz16
EXPNO: 1
PROCNO: 1
Date_: 20070412
Time: 22:21
INSTRUM: avance400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1000
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307972 sec
RG: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.0068955 sec
DELTA: 0.0001311 sec
TD0: 1



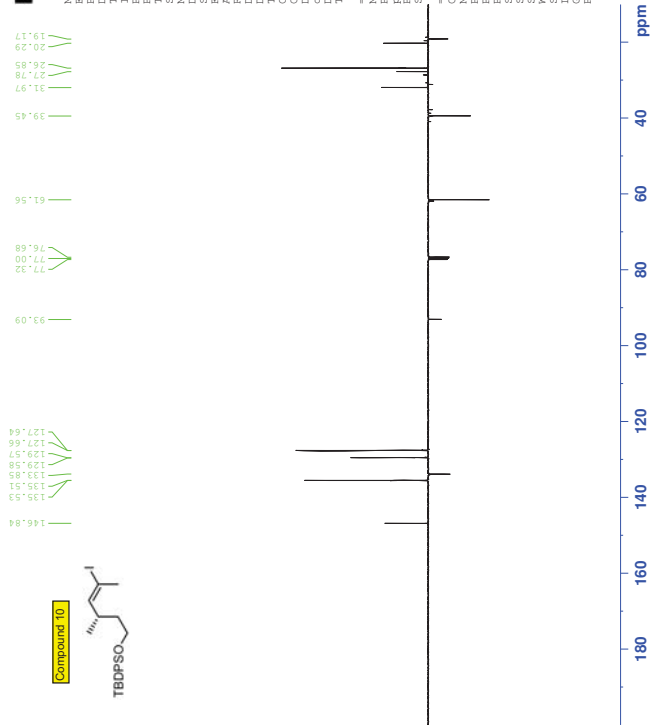
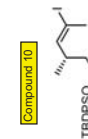
NAME: tirma100
EXPNO: 1
PROCNO: 1
Date_: 20070424
Time: 22:07
INSTRUM: avance400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1000
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307972 sec
RG: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.0068955 sec
DELTA: 0.0001311 sec
TD0: 1

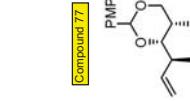
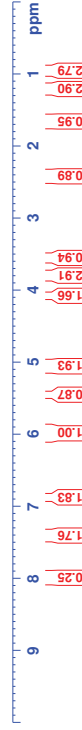
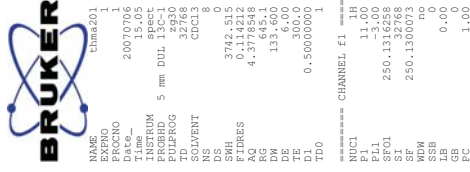
===== CHANNEL f1 =====
NUC1: 13C
P1: 13.00 usec
PL1: -1.00 dB
SF01: 100.6233329 MHz

===== CHANNEL f2 =====
NAME: waltz16
EXPNO: 1
PROCNO: 1
Date_: 20070412
Time: 22:21
INSTRUM: avance400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1000
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307972 sec
RG: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.0068955 sec
DELTA: 0.0001311 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 13C
P1: 13.00 usec
PL1: -1.00 dB
SF01: 100.6233329 MHz

===== CHANNEL f2 =====
NAME: waltz16
EXPNO: 1
PROCNO: 1
Date_: 20070412
Time: 22:21
INSTRUM: avance400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1000
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307972 sec
RG: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.0068955 sec
DELTA: 0.0001311 sec
TD0: 1





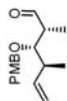


NAME: thma073
PROCNO: 31
Date_: 20070706
Time: 2:53
INSTRUM: avnc600
PROBHD: 5 mm PAQNP Sx1
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 32
DS: 4
SWH: 12376.237 Hz
FIDRES: 0.188846 Hz
AQ: 2.6477449 sec
RG: 655.2
DW: 40.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 1H
P1: 13.85 usec
PL1: 0.00 dB
SF01: 600.1337060 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.00 Hz
GB: 0
PC: 1.00



Compound 12



NAME: thma202
PROCNO: 1
Date_: 20070710
Time: 10:06
INSTRUM: 5 mm DUL 13C-1
PROBHD: zgpg30
SOLVENT: CDCl3
NS: 8
DS: 0
SWH: 3742.915 Hz
FIDRES: 0.114212 Hz
AQ: 4.377848 sec
RG: 655.2
DW: 133.600 usec
DE: 6.00 usec
TE: 300.2 K
D1: 0.5000000 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 13C
P1: 11.00 usec
PL1: 0.00 dB
SF01: 250.1316258 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.00 Hz
GB: 0
PC: 1.00



Compound 78



NAME: thma073
PROCNO: 31
Date_: 20070706
Time: 2:53
INSTRUM: avnc600
PROBHD: 5 mm PAQNP Sx1
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1000
DS: 4
SWH: 35971.223 Hz
FIDRES: 0.548877 Hz
AQ: 0.915743 sec
RG: 13.900 usec
DW: 13.900 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.00689655 sec
DELTA: 0.0001019 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 13C
P1: 16.00 usec
PL1: 0.00 dB
SF01: 150.9179988 MHz

===== CHANNEL f2 =====
NAME: waitz16
PROCNO: 1
Date_: 20070706
Time: 10:06
INSTRUM: 5 mm BBO BB-1H
PROBHD: zgpg30
SOLVENT: CDCl3
NS: 1000
DS: 4
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307452 sec
RG: 19.950 usec
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.00689655 sec
DELTA: 0.0001019 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 13C
P1: 16.00 usec
PL1: 0.00 dB
SF01: 150.9179988 MHz

===== CHANNEL f2 =====
NAME: waitz16
PROCNO: 1
Date_: 20070706
Time: 10:06
INSTRUM: 5 mm BBO BB-1H
PROBHD: zgpg30
SOLVENT: CDCl3
NS: 1000
DS: 4
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307452 sec
RG: 19.950 usec
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.00689655 sec
DELTA: 0.0001019 sec
TD0: 1



Compound 12



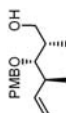
NAME: thma202
PROCNO: 1
Date_: 20070710
Time: 10:06
INSTRUM: avnc600
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1000
DS: 4
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307452 sec
RG: 19.950 usec
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.00689655 sec
DELTA: 0.0001019 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 13C
P1: 16.00 usec
PL1: 0.00 dB
SF01: 150.9179988 MHz

===== CHANNEL f2 =====
NAME: waitz16
PROCNO: 1
Date_: 20070706
Time: 10:06
INSTRUM: 5 mm BBO BB-1H
PROBHD: zgpg30
SOLVENT: CDCl3
NS: 1000
DS: 4
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307452 sec
RG: 19.950 usec
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.00689655 sec
DELTA: 0.0001019 sec
TD0: 1



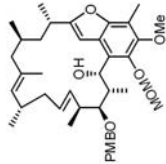
Compound 78



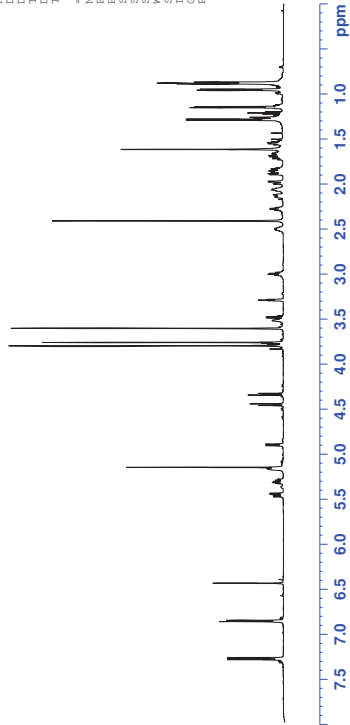


NAME: TMR1_132
EXPNO: 1
PROCNO: 1
Date_: 20070516
Time: 19:45
INSTRUM: avnnc600
PROBHD: 5 mm TBI 1H/13
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 32
DS: 4
SWH: 4807.692 Hz
FIDRES: 0.293438 Hz
AQ: 1.7045900 sec
RG: 327.68
DW: 104.000 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

CHANNEL f1 1H
NUC1: 1H
P1: 8.50 usec
PL1: -2.00 dB
SF01: 600.1324005 MHz
SI: 65536
WDW: EM
SSB: 0
LB: 0.00 Hz
GB: 0
PC: 1.00

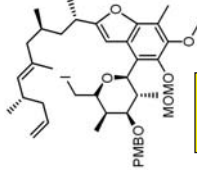


Compound 20a

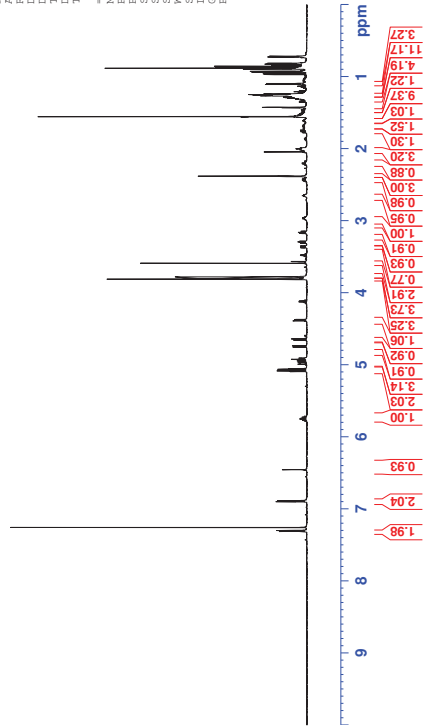


NAME: TMR1_49
EXPNO: 1
PROCNO: 1
Date_: 20070530
Time: 17:08
INSTRUM: avnnc600
PROBHD: 5 mm PAQNP SW1
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 32
DS: 4
SWH: 12376.237 Hz
FIDRES: 0.188846 Hz
AQ: 2.6477439 sec
RG: 327.68
DW: 40.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

CHANNEL f1 1H
NUC1: 1H
P1: 13.65 usec
PL1: -2.00 dB
SF01: 600.1337060 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.00 Hz
GB: 0
PC: 1.00



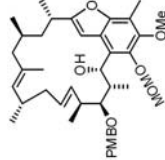
Compound 19



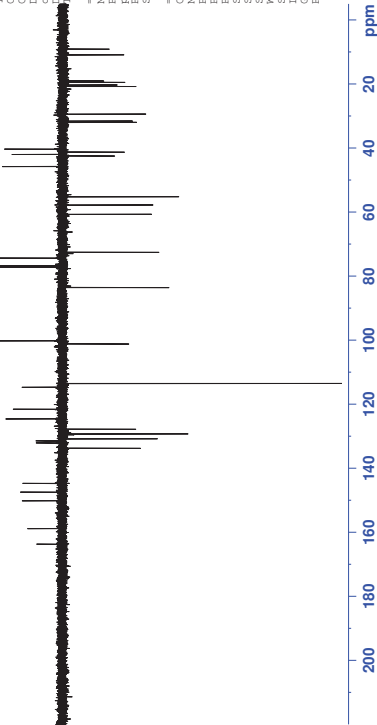
NAME: TMR1_132
EXPNO: 1
PROCNO: 1
Date_: 20070517
Time: 19:45
INSTRUM: avnnc600
PROBHD: 5 mm TBI 1H/13
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 3072
DS: 4
SWH: 36231.883 Hz
FIDRES: 0.552855 Hz
AQ: 0.9044006 sec
RG: 327.68
DW: 13.800 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.00689455 sec
DELTA: 0.00001592 sec
TD0: 1

CHANNEL f1 13C
NUC1: 13C
P1: 13.65 usec
PL1: -2.00 dB
SF01: 150.9185938 MHz

CHANNEL f2 waltz16
CPDPRG2: waltz16
NUC2: 13C
P2: 25.00 usec
PL2: -2.00 dB
SF02: 600.1324005 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.00 Hz
GB: 0
PC: 1.40



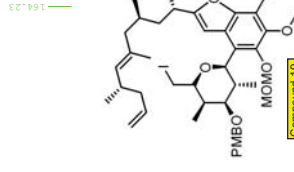
Compound 20b



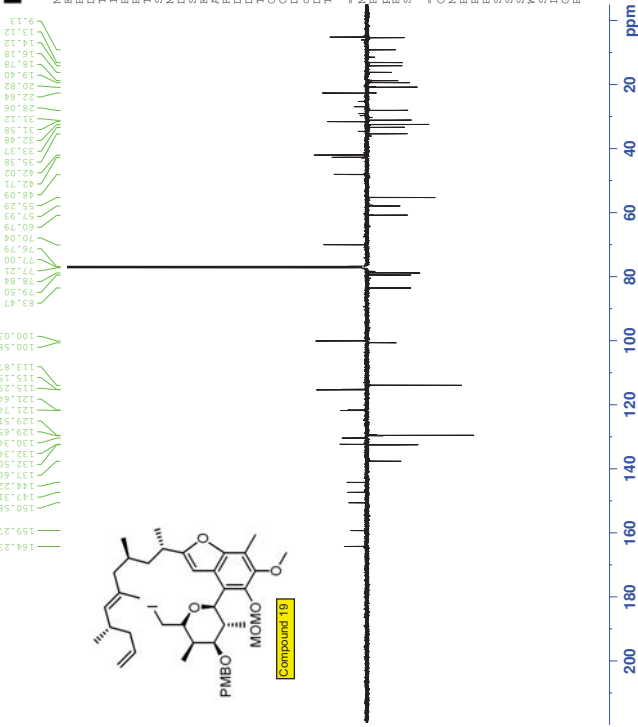
NAME: TMR1_49
EXPNO: 1
PROCNO: 1
Date_: 20070531
Time: 17:08
INSTRUM: avnnc600
PROBHD: 5 mm PAQNP SW1
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 8280
DS: 4
SWH: 35971.223 Hz
FIDRES: 0.548877 Hz
AQ: 0.9159743 sec
RG: 327.68
DW: 13.900 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.00689455 sec
DELTA: 0.00001019 sec
TD0: 1

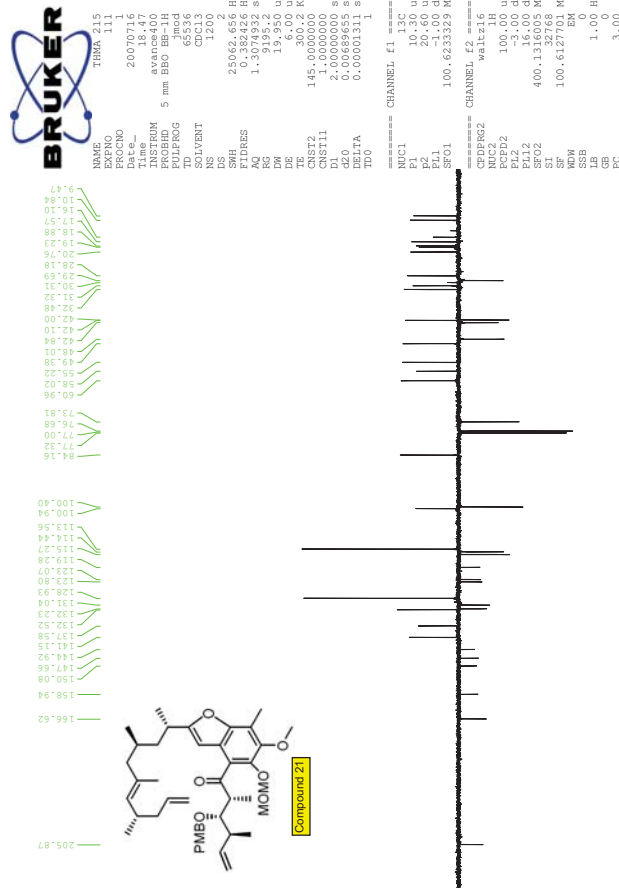
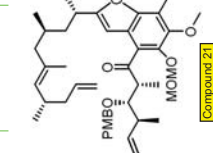
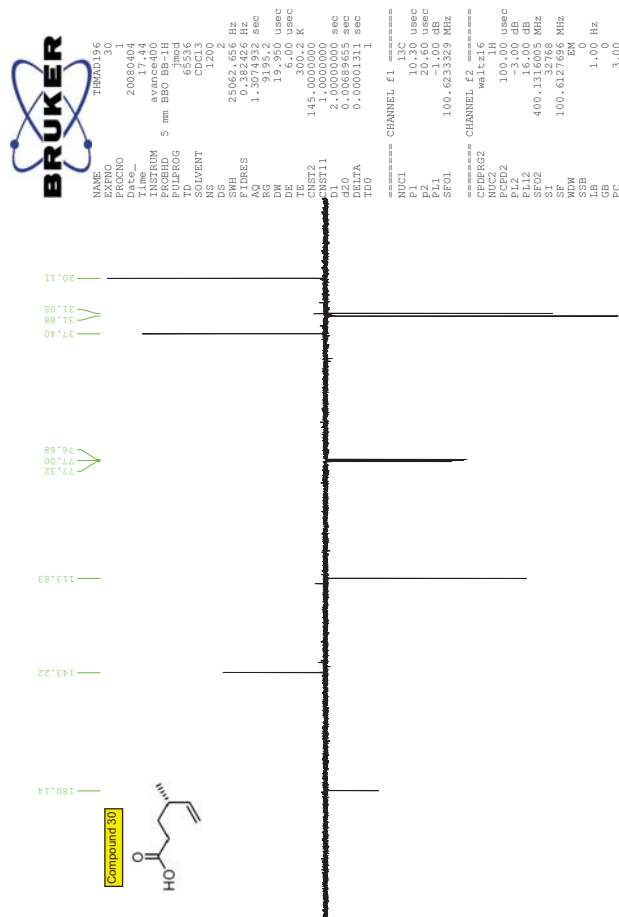
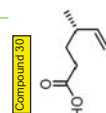
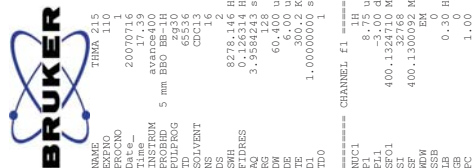
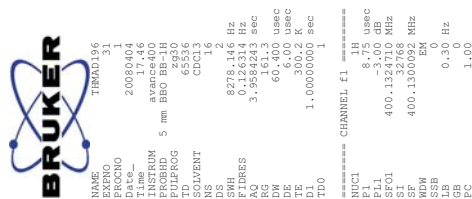
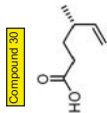
CHANNEL f1 13C
NUC1: 13C
P1: 13.65 usec
PL1: -2.00 dB
SF01: 150.9178988 MHz

CHANNEL f2 waltz16
CPDPRG2: waltz16
NUC2: 13C
P2: 25.00 usec
PL2: -2.00 dB
SF02: 600.1324005 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.00 Hz
GB: 0
PC: 4.00



Compound 19



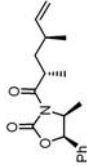




NAME: tlmad203
EXPNO: 4
PROCNO: 1
Date_: 20080409
Time: 11.04
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.998423 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 1H
P1: 8.75 usec
PL1: -2.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00

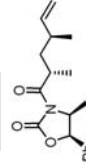
Compound 79



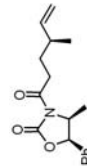
NAME: tlmad203
EXPNO: 4
PROCNO: 1
Date_: 20080409
Time: 11.04
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.998423 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 1H
P1: 8.75 usec
PL1: -2.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00

Compound 78



Compound 33



NAME: tlmad199
EXPNO: 4
PROCNO: 1
Date_: 20080407
Time: 21.04
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.998423 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

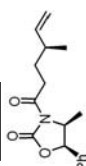
===== CHANNEL f1 =====
NUC1: 1H
P1: 8.75 usec
PL1: -2.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00



NAME: tlmad199
EXPNO: 4
PROCNO: 1
Date_: 20080407
Time: 21.04
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.998423 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 1H
P1: 8.75 usec
PL1: -2.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00

Compound 32

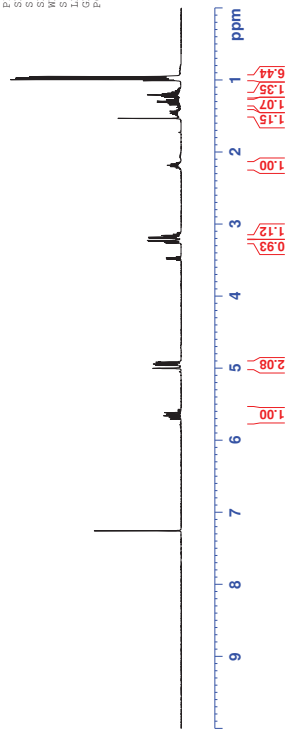




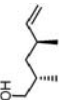
NAME: thmad098
EXPNO: 30
PROCNO: 1
Date_: 20080131
Time: 11:24
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DSH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.994243 sec
RG: 60.400 usec
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

CHANNEL f1
NUC1: 1H
P1: 8.75 usec
PL1: -2.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00

Compound 28

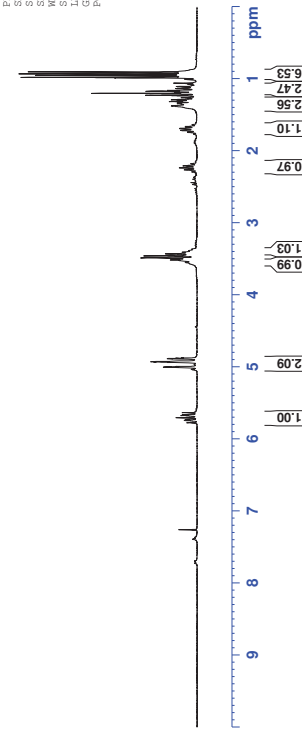


Compound 34



NAME: thmad194
EXPNO: 30
PROCNO: 1
Date_: 20080403
Time: 16:11
INSTRUM: avnnc400
PROBHD: 5 mm DUL 13C-1
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 8
DSH: 3742.915 Hz
FIDRES: 0.114212 Hz
AQ: 4.3778548 sec
RG: 133.600 usec
DW: 133.600 usec
DE: 6.00 usec
TE: 300.2 K
D1: 0.5000000 sec
TD0: 1

CHANNEL f1
NUC1: 1H
P1: 11.00 usec
PL1: -2.00 dB
SF01: 250.1316258 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.10 Hz
GB: 0
PC: 1.00



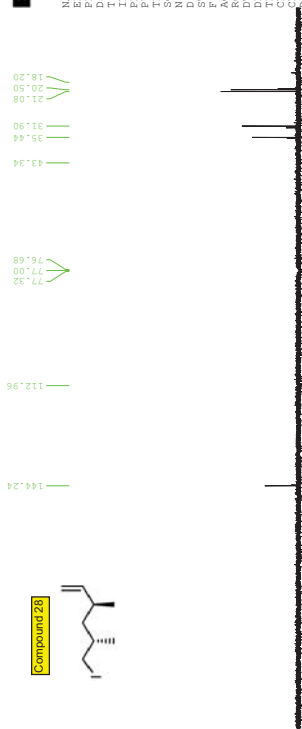
NAME: thmad098
EXPNO: 30
PROCNO: 1
Date_: 20080131
Time: 11:24
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1500
DSH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307452 sec
RG: 19.950 usec
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.0068955 sec
DELTA: 0.0000131 sec
TD0: 1

CHANNEL f1
NUC1: 13C
P1: 13.00 usec
PL1: -2.00 dB
SF01: 100.6233329 MHz

CHANNEL f2
NAME: waltz16
EXPNO: 1
PROCNO: 1
Date_: 20080131
Time: 11:24
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1500
DSH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307452 sec
RG: 19.950 usec
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.0068955 sec
DELTA: 0.0000131 sec
TD0: 1

CHANNEL f2
NAME: waltz16
EXPNO: 1
PROCNO: 1
Date_: 20080131
Time: 11:24
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1500
DSH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307452 sec
RG: 19.950 usec
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.0068955 sec
DELTA: 0.0000131 sec
TD0: 1

Compound 28



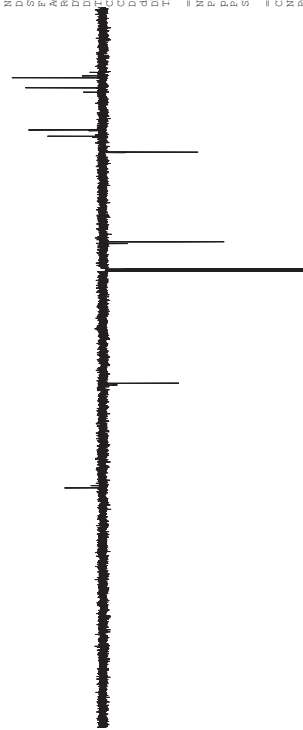
NAME: thma d095
EXPNO: 30
PROCNO: 1
Date_: 20080130
Time: 11:24
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1779
DSH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307452 sec
RG: 19.950 usec
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.0068955 sec
DELTA: 0.0000131 sec
TD0: 1

CHANNEL f1
NUC1: 13C
P1: 13.00 usec
PL1: -2.00 dB
SF01: 100.6233329 MHz

CHANNEL f2
NAME: waltz16
EXPNO: 1
PROCNO: 1
Date_: 20080130
Time: 11:24
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1779
DSH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307452 sec
RG: 19.950 usec
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.0068955 sec
DELTA: 0.0000131 sec
TD0: 1

CHANNEL f2
NAME: waltz16
EXPNO: 1
PROCNO: 1
Date_: 20080130
Time: 11:24
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1779
DSH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307452 sec
RG: 19.950 usec
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.0068955 sec
DELTA: 0.0000131 sec
TD0: 1

Compound 34

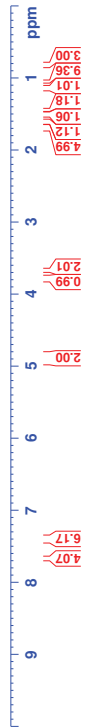
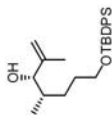




NAME: tbaad16-f3
PROCNO: 1
Date_: 20080214
Time: 11.40
INSTRUM: avnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 1H
P1: 8.75 usec
PL1: -2.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00

Compound 29a

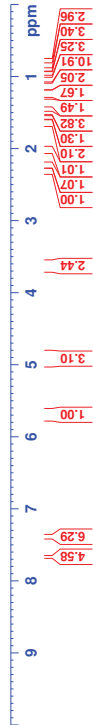


Compound 35



NAME: tbaad16-f3
PROCNO: 1
Date_: 20080214
Time: 11.40
INSTRUM: avnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

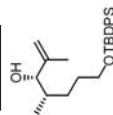
===== CHANNEL f1 =====
NUC1: 1H
P1: 8.75 usec
PL1: -2.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00



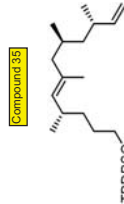
NAME: tbaad16-f3
PROCNO: 1
Date_: 20080214
Time: 11.40
INSTRUM: avnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 1H
P1: 8.75 usec
PL1: -2.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00

Compound 29a



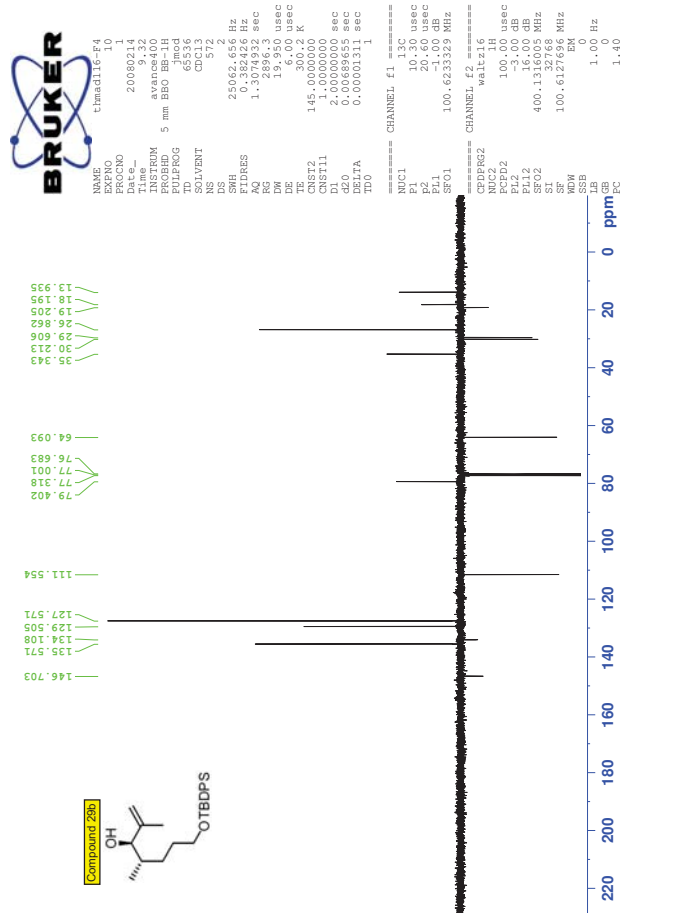
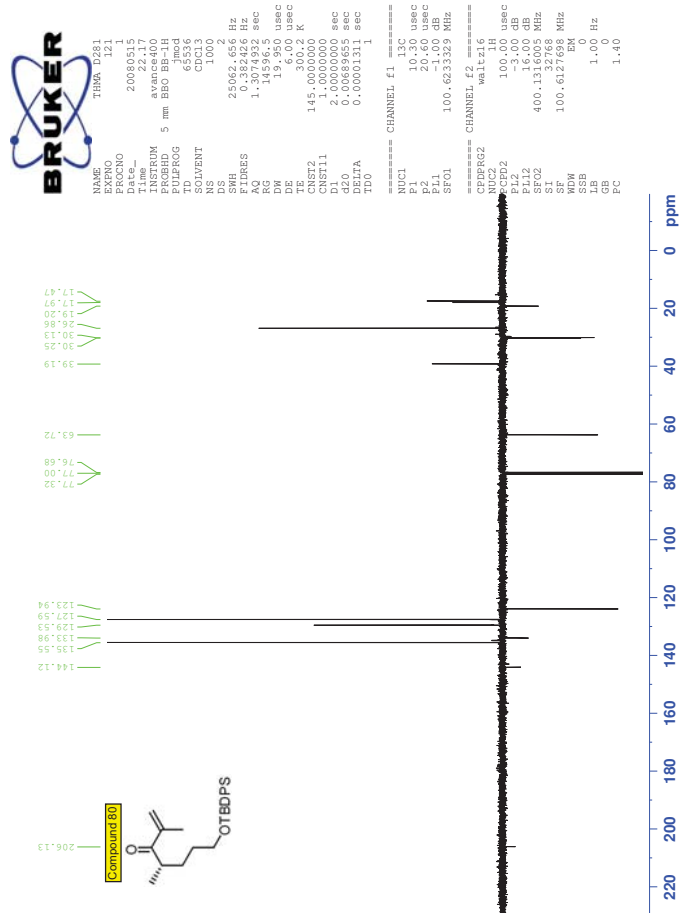
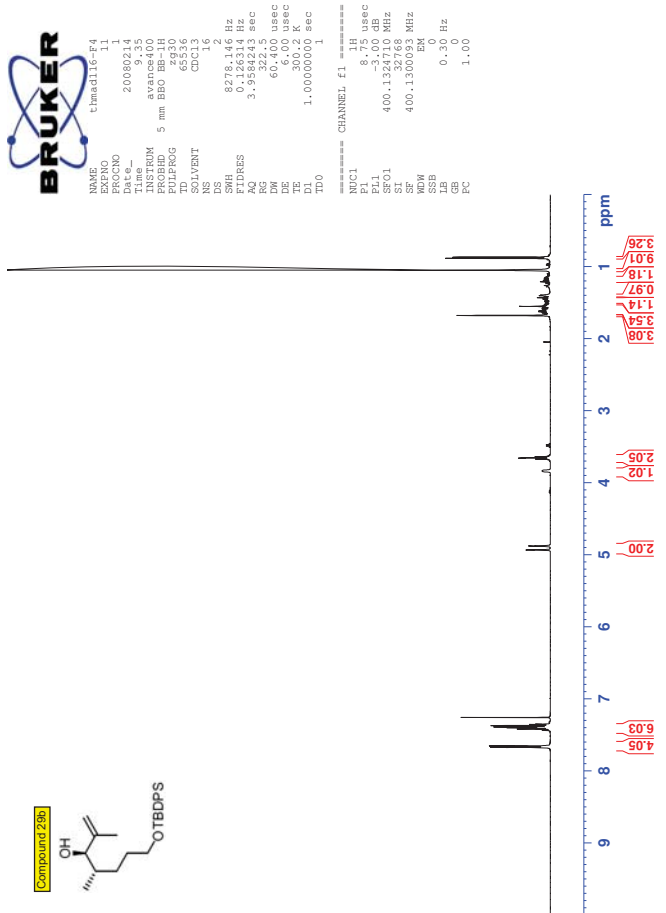
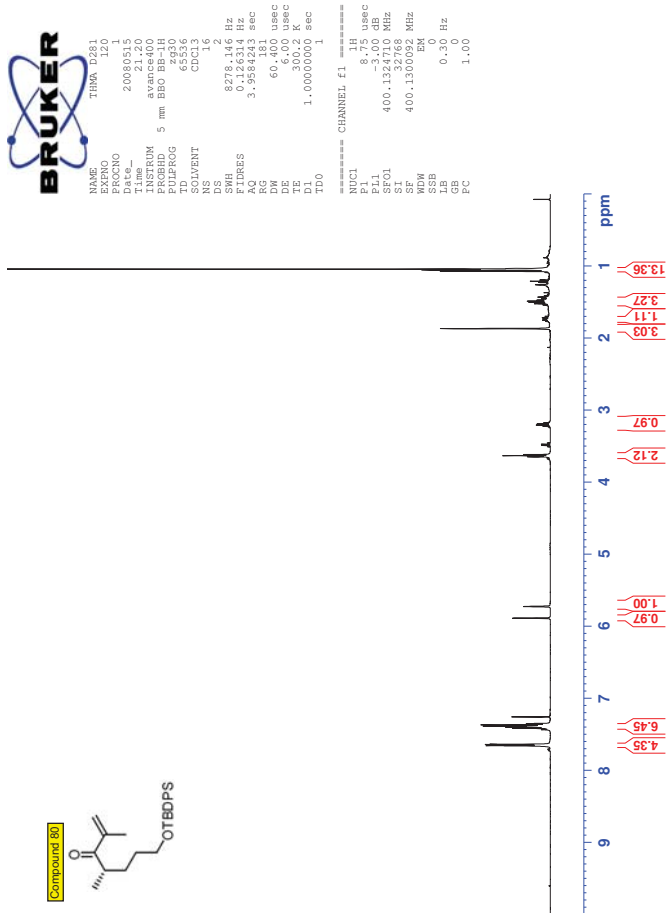
Compound 35

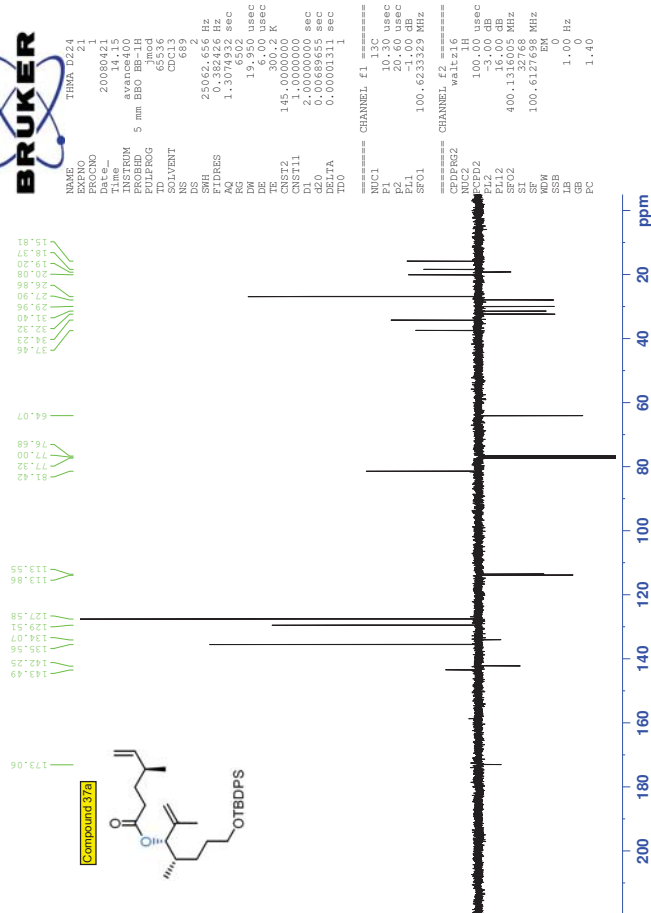
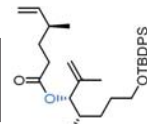
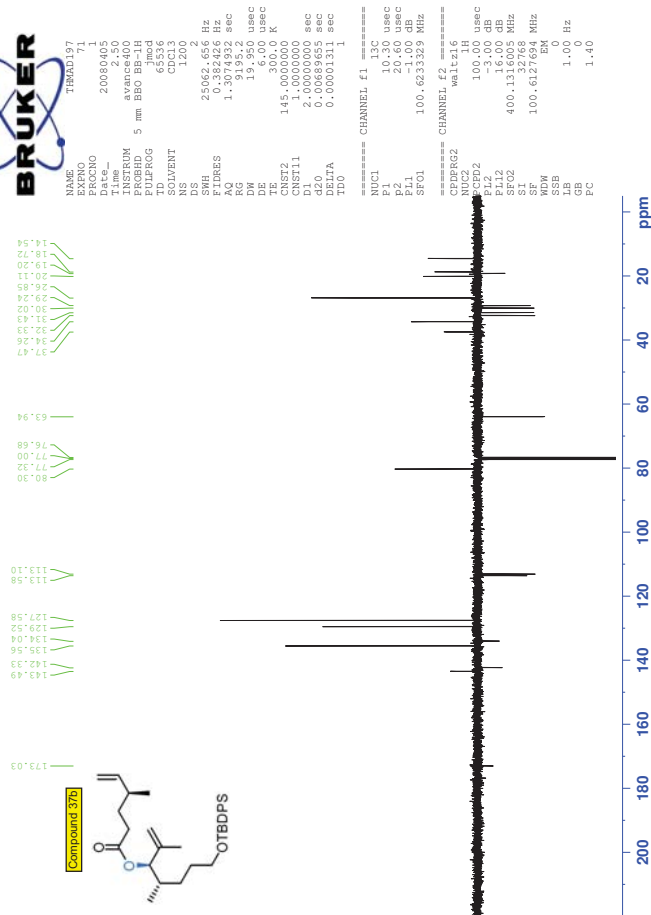
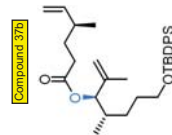
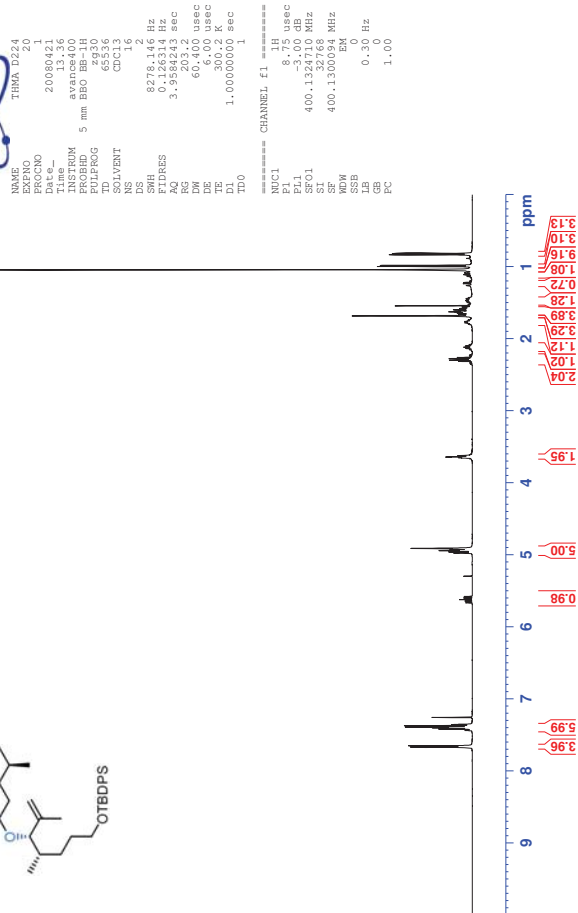
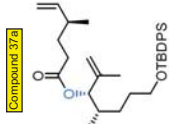
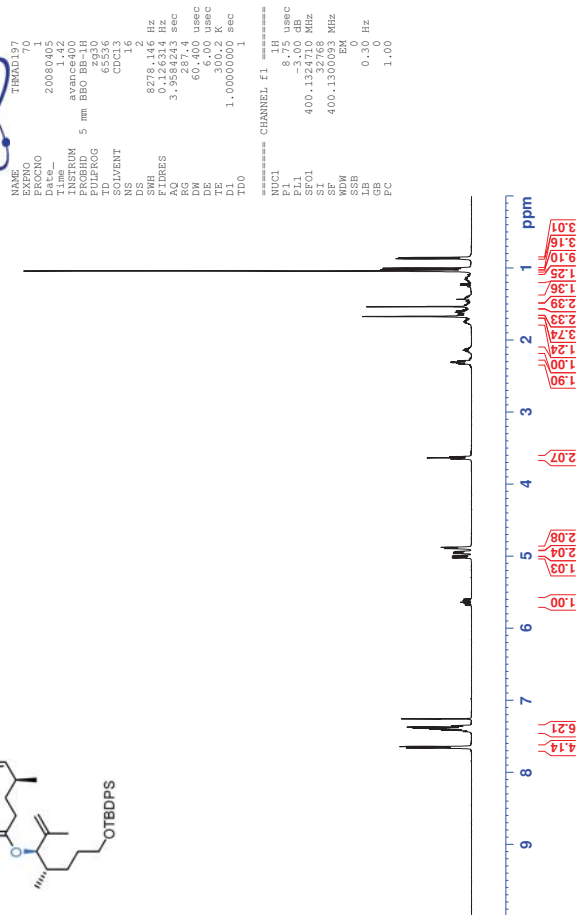
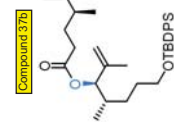


NAME: tbaad16-f3
PROCNO: 1
Date_: 20080214
Time: 11.40
INSTRUM: avnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 1H
P1: 8.75 usec
PL1: -2.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00



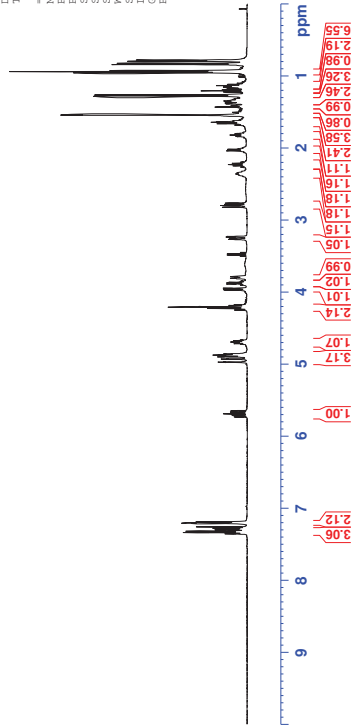
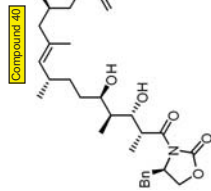






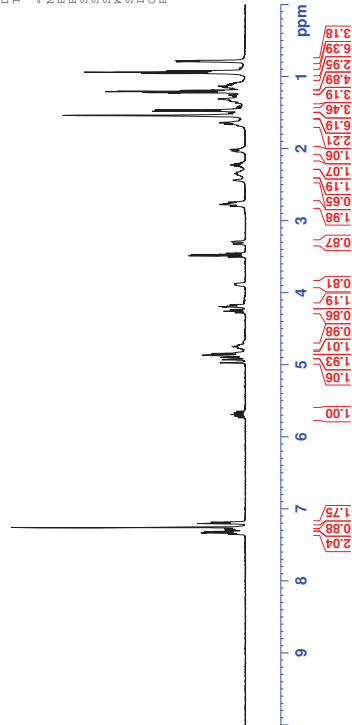
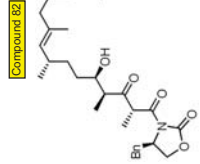
NAME: THNA D222
EXPNO: 243
PROCNO: 1
Date_: 20080419
Time: 12:08
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 1H
P1: 8.75 usec
PL1: -2.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00



NAME: THNA D221D2
EXPNO: 243
PROCNO: 1
Date_: 20080417
Time: 21:05
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

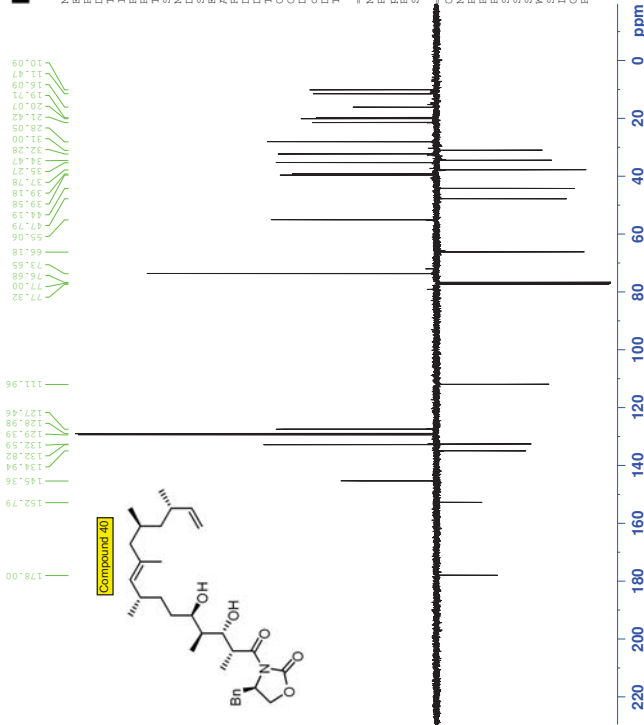
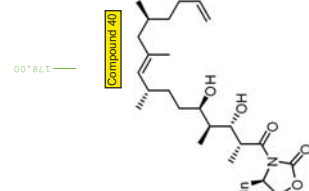
===== CHANNEL f1 =====
NUC1: 1H
P1: 8.75 usec
PL1: -2.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00

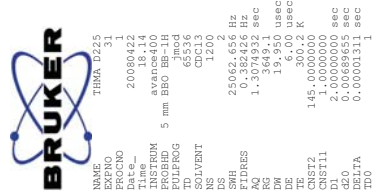
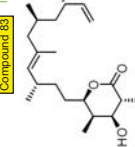
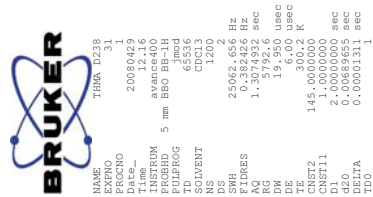
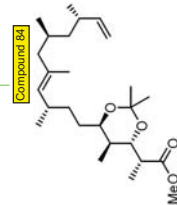
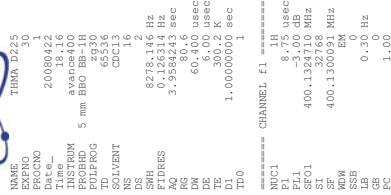
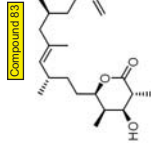
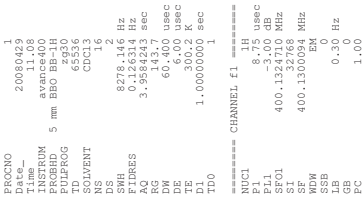
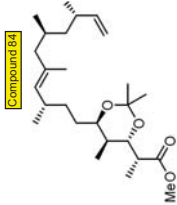


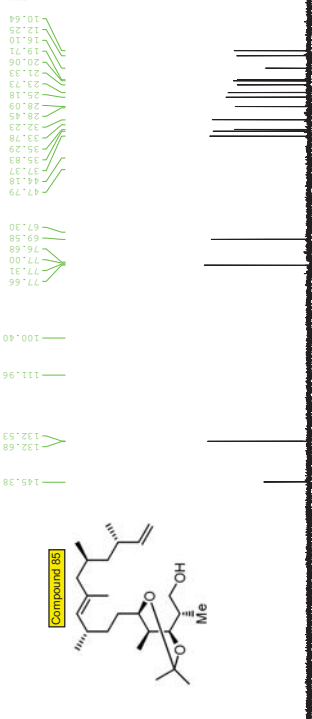
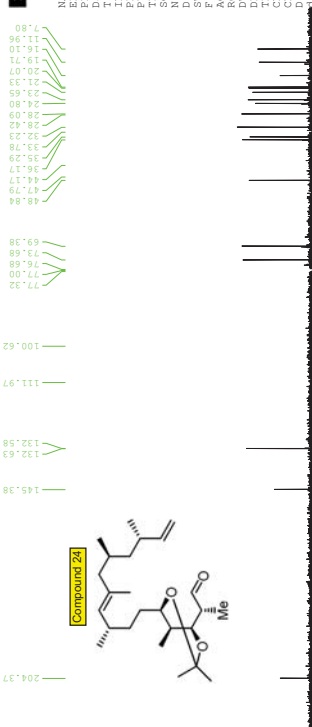
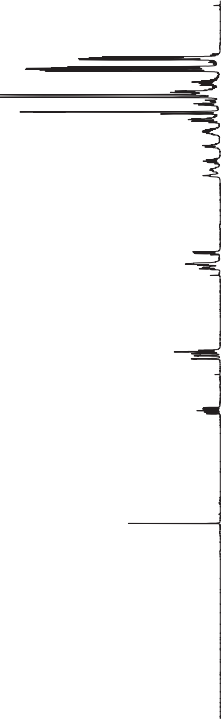
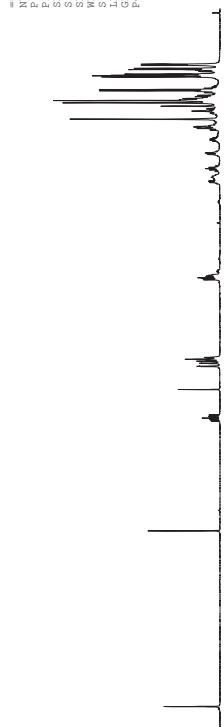
NAME: THNA D222
EXPNO: 243
PROCNO: 1
Date_: 20080419
Time: 12:08
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1200
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.3074532 sec
RG: 19.950
DW: 19.950 usec
DE: 26.00 usec
TE: 300.2 K
CNST12: 145.0000000 K
CNST11: 1.0000000
PCPD2: 100.00 usec
PL2: -3.00 dB
SF02: 400.1316005 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 1.00 Hz
GB: 0
PC: 5.00

===== CHANNEL f1 =====
NUC1: 13C
P1: 10.30 usec
PL1: -1.00 dB
SF01: 100.6233329 MHz

===== CHANNEL f2 =====
NAME: waitz16
EXPNO: 1
PROCNO: 1
Date_: 20080419
Time: 12:08
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1200
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.3074532 sec
RG: 19.950
DW: 19.950 usec
DE: 26.00 usec
TE: 300.2 K
CNST12: 145.0000000 K
CNST11: 1.0000000
PCPD2: 100.00 usec
PL2: -3.00 dB
SF02: 400.1316005 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 1.00 Hz
GB: 0
PC: 5.00







Compound 97



NAME: tlmad129
EXPNO: 101
PROCNO: 1
Date_: 20080220
Time: 0:27
INSTRUM: avnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 327.68
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 1H
P1: 8.75 usec
PL1: -2.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00



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NAME: tlmad129
EXPNO: 101
PROCNO: 1
Date_: 20080220
Time: 0:27
INSTRUM: avnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 2000
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.309532 sec
RG: 327.68
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.00689655 sec
DELTA: 0.00001311 sec
TD0: 1

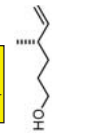
===== CHANNEL f1 =====
NUC1: 13C
P1: 13.00 usec
PL1: -1.00 dB
SF01: 100.6233329 MHz

===== CHANNEL f2 =====
NAME: waltz16
EXPNO: 101
PROCNO: 1
Date_: 20080220
Time: 0:27
INSTRUM: avnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 2000
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.309532 sec
RG: 327.68
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.00689655 sec
DELTA: 0.00001311 sec
TD0: 1

===== CHANNEL f2 =====
NAME: waltz16
EXPNO: 101
PROCNO: 1
Date_: 20080220
Time: 0:27
INSTRUM: avnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 2000
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.309532 sec
RG: 327.68
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.00689655 sec
DELTA: 0.00001311 sec
TD0: 1

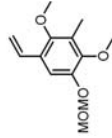


Compound 97



NAME: tlmad129
EXPNO: 101
PROCNO: 1
Date_: 20090209
Time: 16:22
INSTRUM: avnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 327.68
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 1H
P1: 8.75 usec
PL1: -2.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00



NAME: tlmad129
EXPNO: 101
PROCNO: 1
Date_: 20090209
Time: 16:22
INSTRUM: avnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1200
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.309532 sec
RG: 327.68
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.00689655 sec
DELTA: 0.00001311 sec
TD0: 1

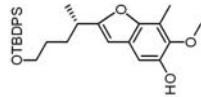
===== CHANNEL f1 =====
NUC1: 13C
P1: 13.00 usec
PL1: -1.00 dB
SF01: 100.6233329 MHz

===== CHANNEL f2 =====
NAME: waltz16
EXPNO: 101
PROCNO: 1
Date_: 20090209
Time: 16:22
INSTRUM: avnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1200
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.309532 sec
RG: 327.68
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.00689655 sec
DELTA: 0.00001311 sec
TD0: 1

===== CHANNEL f2 =====
NAME: waltz16
EXPNO: 101
PROCNO: 1
Date_: 20090209
Time: 16:22
INSTRUM: avnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1200
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.309532 sec
RG: 327.68
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D20: 0.00689655 sec
DELTA: 0.00001311 sec
TD0: 1

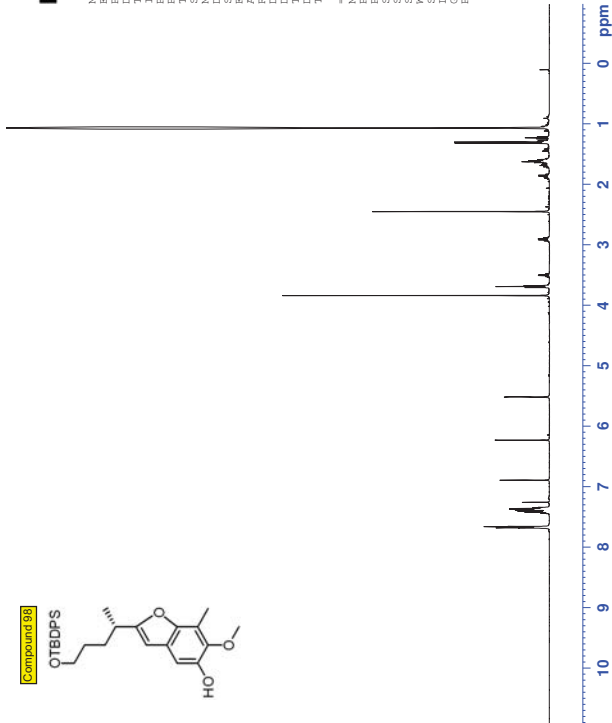


Compound 98

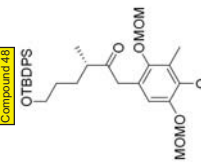


NAME THMA D356
EXPNO 6
PROCNO 1
Date_ 20080612
Time 21.00
INSTRUM avac400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 -2.00 dB
SF01 400.1324710 MHz
SI 32768
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

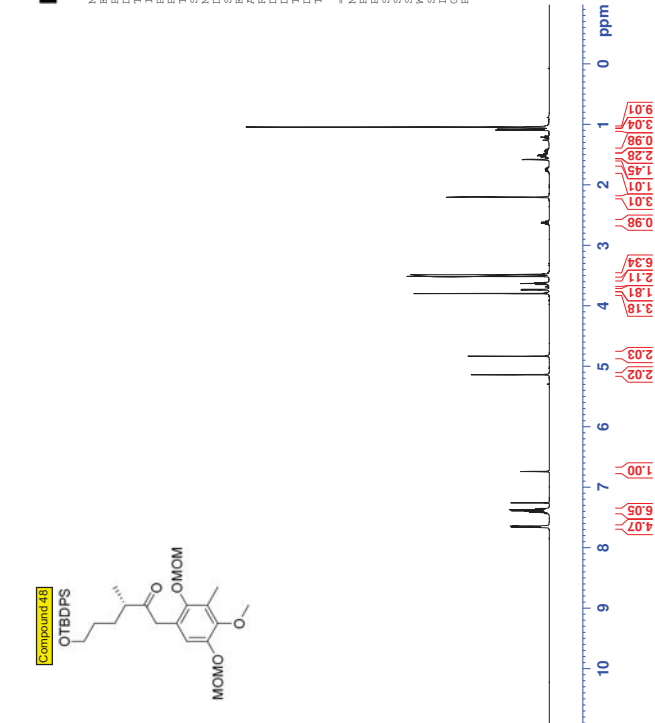


Compound 48



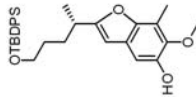
NAME THMA D413
EXPNO 6
PROCNO 1
Date_ 20080707
Time 10.00
INSTRUM avac400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 -2.00 dB
SF01 400.1324710 MHz
SI 32768
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



94

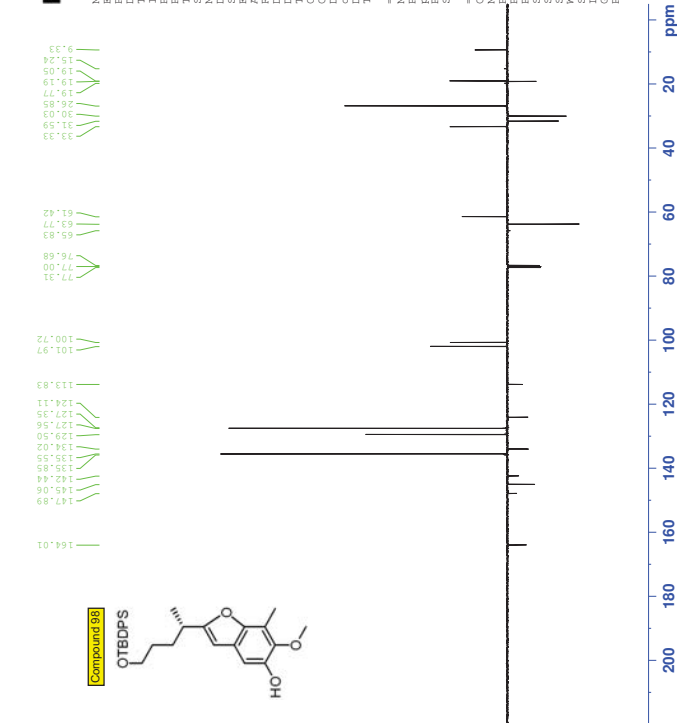
Compound 98



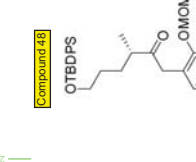
NAME THMA D356
EXPNO 6
PROCNO 1
Date_ 20080612
Time 21.00
INSTRUM avac400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
SOLVENT CDCl3
NS 1000
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3078322 sec
RG 19.950 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
D2 0.00689455 sec
DELTA 0.00001311 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -1.00 dB
SF01 100.6233329 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
PCPD2 100.00 usec
PL2 -3.00 dB
SF02 400.1316005 MHz
SI 32768
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



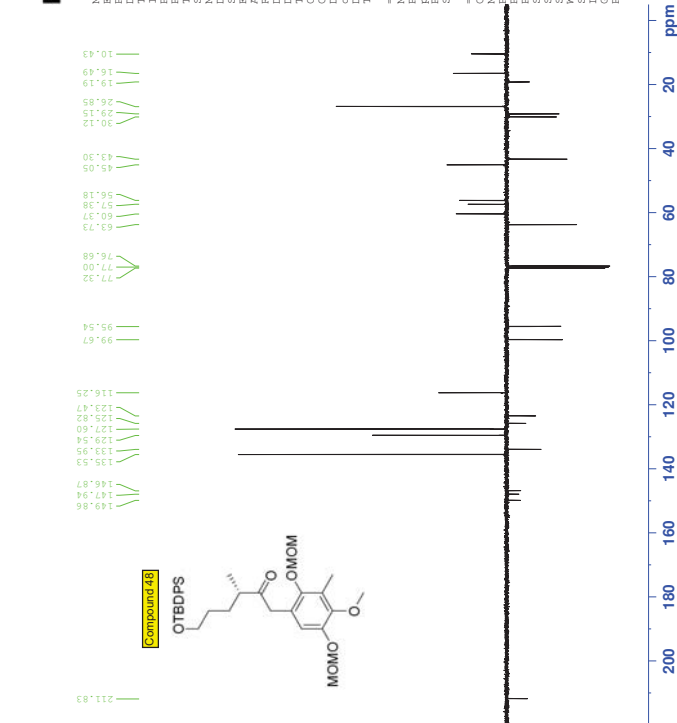
Compound 48



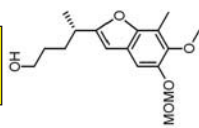
NAME THMA D413
EXPNO 6
PROCNO 1
Date_ 20080707
Time 10.00
INSTRUM avac400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
SOLVENT CDCl3
NS 1200
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3078322 sec
RG 19.950 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
D2 0.00689455 sec
DELTA 0.00001311 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -1.00 dB
SF01 100.6233329 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
PCPD2 100.00 usec
PL2 -3.00 dB
SF02 400.1316005 MHz
SI 32768
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



Compound 9b



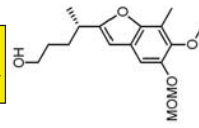
NAME THMA D410
EXPNO 7
PROCNO 1
Date_ 20080704
Time 23.09
INSTRUM avnc400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 6550
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
TD0 1

CHANNEL f1 1H
NUC1 1H
P1 8.75 usec
PL1 -2.00 dB
SF01 400.1324710 MHz
SI 32768
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



95

Compound 9b

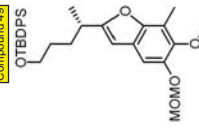


NAME THMA D410
EXPNO 7
PROCNO 1
Date_ 20080705
Time 23.09
INSTRUM avnc400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 6550
SOLVENT CDCl3
NS 1200
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3076532 sec
RG 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
D2 0.00689655 sec
DELTA 0.00001311 sec
TD0 1

CHANNEL f1 1H
NUC1 1H
P1 8.75 usec
PL1 -2.00 dB
SF01 400.1324710 MHz
SI 32768
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Compound 49



NAME THMA D409
EXPNO 7
PROCNO 1
Date_ 20080704
Time 13.40
INSTRUM avnc400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 6550
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
TD0 1

CHANNEL f1 1H
NUC1 1H
P1 8.75 usec
PL1 -2.00 dB
SF01 400.1324710 MHz
SI 32768
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



NAME THMA D409
EXPNO 7
PROCNO 1
Date_ 20080704
Time 13.40
INSTRUM avnc400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 6550
SOLVENT CDCl3
NS 804
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3076532 sec
RG 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
D2 0.00689655 sec
DELTA 0.00001311 sec
TD0 1

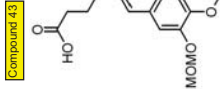
CHANNEL f1 1H
NUC1 1H
P1 8.75 usec
PL1 -2.00 dB
SF01 400.1324710 MHz
SI 32768
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.40





NAME: THMA D738
PROCNO: 1
Date: 20081215
Time: 8:27
INSTRUM: AVI1000
PROBHD: 5 mm PABBO BB-
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8223.685 Hz
FIDRES: 0.125483 Hz
AQ: 3.9846387 sec
RG: 655.000
DW: 60.800 usec
DE: 6.50 usec
TE: 293.2 K
D1: 2.000000 sec
TD0: 1

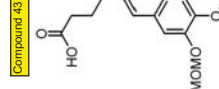
===== CHANNEL f1 =====
NUC1: 1H
P1: 13.50 usec
PL1: 0.00 dB
F1F2: 400.132410 MHz
SF01: 400.2724718 MHz
SF: 400.2701015 MHz
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00



NAME: THMA D412
PROCNO: 1
Date: 20080705
Time: 15:00
INSTRUM: avance400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1200
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307432 sec
RG: 19.950
DW: 19.950 usec
DE: 26.00 usec
TE: 300.2 K
CNST2: 145.000000
CNST11: 1.0000000
D1: 2.0000000 sec
d20: 0.0068965 sec
DELTA: 0.00001311 sec
TD0: 1

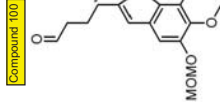
===== CHANNEL f1 =====
NUC1: 13C
P1: 13.00 usec
PL1: 20.40 usec
PL2: -1.00 dB
SF01: 100.6233329 MHz

===== CHANNEL f2 =====
NAME: waitz16
PROCNO: 1
Date: 20080705
Time: 15:00
INSTRUM: avance400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1200
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307432 sec
RG: 19.950
DW: 19.950 usec
DE: 26.00 usec
TE: 300.2 K
CNST2: 145.000000
CNST11: 1.0000000
D1: 2.0000000 sec
d20: 0.0068965 sec
DELTA: 0.00001311 sec
TD0: 1



NAME: THMA D411
PROCNO: 1
Date: 20080705
Time: 15:00
INSTRUM: avance400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.000
DW: 60.400 usec
DE: 6.00 usec
TE: 293.2 K
D1: 2.000000 sec
TD0: 1

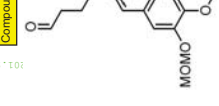
===== CHANNEL f1 =====
NUC1: 1H
P1: 8.75 usec
PL1: 0.00 dB
F1F2: 400.132410 MHz
SF01: 400.132410 MHz
SF: 400.132410 MHz
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00



NAME: THMA D411
PROCNO: 1
Date: 20080705
Time: 15:00
INSTRUM: avance400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1200
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307432 sec
RG: 19.950
DW: 19.950 usec
DE: 26.00 usec
TE: 300.2 K
CNST2: 145.000000
CNST11: 1.0000000
D1: 2.0000000 sec
d20: 0.0068965 sec
DELTA: 0.00001311 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 13C
P1: 13.00 usec
PL1: 20.40 usec
PL2: -1.00 dB
SF01: 100.6233329 MHz

===== CHANNEL f2 =====
NAME: waitz16
PROCNO: 1
Date: 20080705
Time: 15:00
INSTRUM: avance400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1200
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.307432 sec
RG: 19.950
DW: 19.950 usec
DE: 26.00 usec
TE: 300.2 K
CNST2: 145.000000
CNST11: 1.0000000
D1: 2.0000000 sec
d20: 0.0068965 sec
DELTA: 0.00001311 sec
TD0: 1

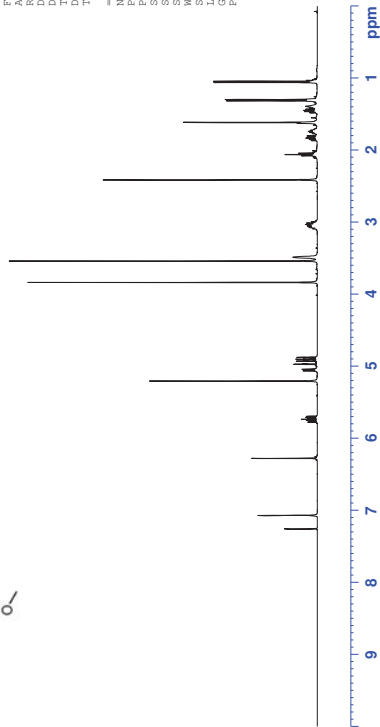
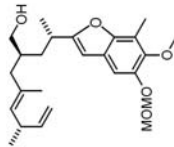




NAME: THMA D579
EXPNO: 41
PROCNO: 1
Date_: 20080915
Time: 14.51
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

CHANNEL f1
NUC1: 1H
P1: 8.75 usec
PL1: -1.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00

Compound S51

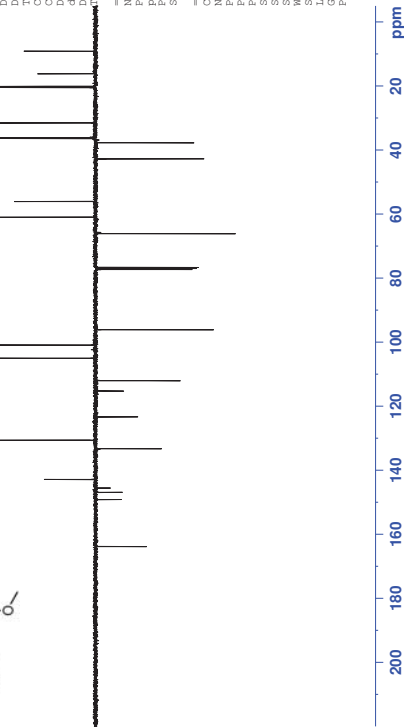
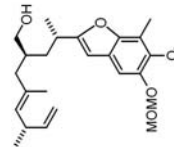


NAME: THMA D579
EXPNO: 41
PROCNO: 1
Date_: 20080915
Time: 14.51
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

CHANNEL f1
NUC1: 13C
P1: 10.00 usec
PL1: -1.00 dB
SF01: 100.6233329 MHz

CHANNEL f2
NAME: waitz16
PROCNO: 1
Date_: 20080915
Time: 14.51
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

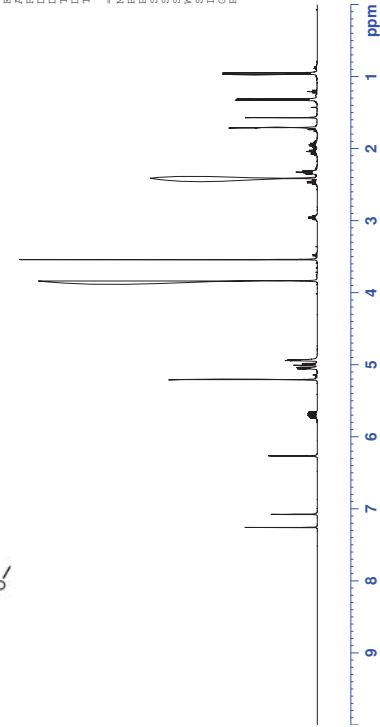
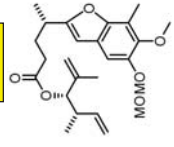
Compound S51



NAME: THMA D595
EXPNO: 1
PROCNO: 1
Date_: 20081009
Time: 20.24
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

CHANNEL f1
NUC1: 1H
P1: 8.75 usec
PL1: -1.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00

Compound 50

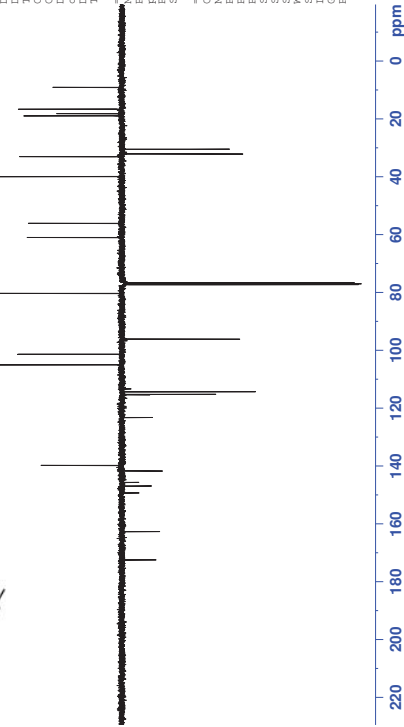
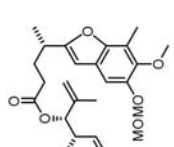


NAME: THMA D595
EXPNO: 1
PROCNO: 1
Date_: 20081009
Time: 20.24
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

CHANNEL f1
NUC1: 13C
P1: 10.00 usec
PL1: -1.00 dB
SF01: 100.6233329 MHz

CHANNEL f2
NAME: waitz16
PROCNO: 1
Date_: 20081009
Time: 20.24
INSTRUM: avnnc400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

Compound 50

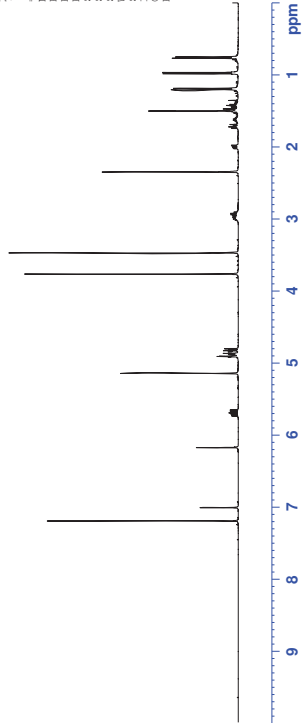
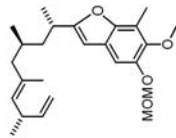




NAME: THMA D748
EXPNO: 60
PROCNO: 1
Date_: 20090102
Time: 17:49
INSTRUM: AVANCE400
PROBHD: 5 mm PABBO BB-
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8223.685 Hz
FIDRES: 0.125483 Hz
AQ: 3.9846387 sec
RG: 655.5
DW: 60.800 usec
DE: 6.50 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 1H
P1: 13.50 usec
PL1: 0.00 dB
FLLW: 15.28361320 W
SF01: 400.2724718 MHz
SI: 32768
WDW: EM
SSB: 0
GB: 0
PC: 1.00

Compound 92

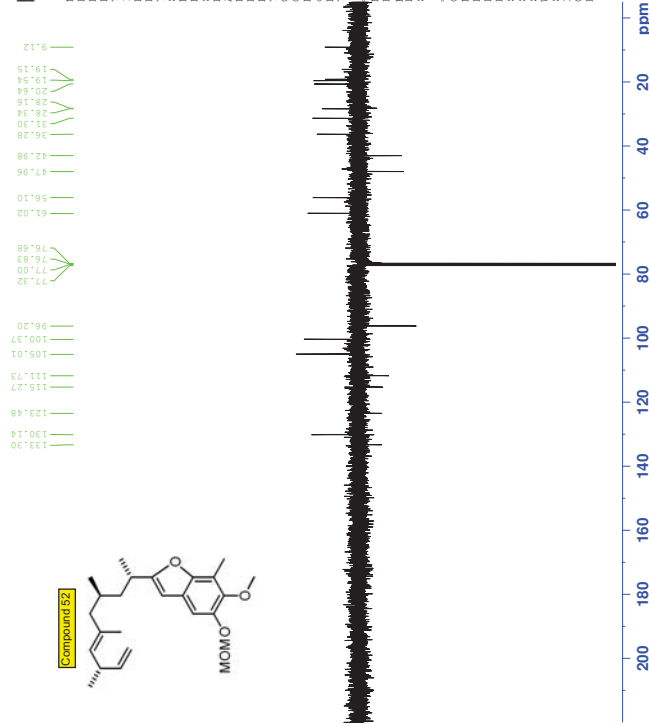
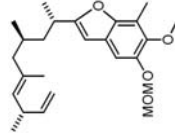


NAME: THMA D603
EXPNO: 21
PROCNO: 1
Date_: 20081015
Time: 12:50
INSTRUM: AVANCE400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1200
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.3074052 sec
RG: 655.5
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D2: 0.0068955 sec
DELTA: 0.00001311 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 13C
P1: 10.30 usec
PL1: 0.00 dB
PL2: 20.40 usec
PL3: -1.00 dB
SF01: 100.623329 MHz

===== CHANNEL f2 =====
CPDPRG2: waltz16
NUC2: 1H
PCPD2: 100.00 usec
PL2: -3.00 dB
PL3: 20.40 usec
SF02: 400.1316005 MHz
SI: 32768
WDW: EM
SSB: 0
GB: 0
PC: 1.10

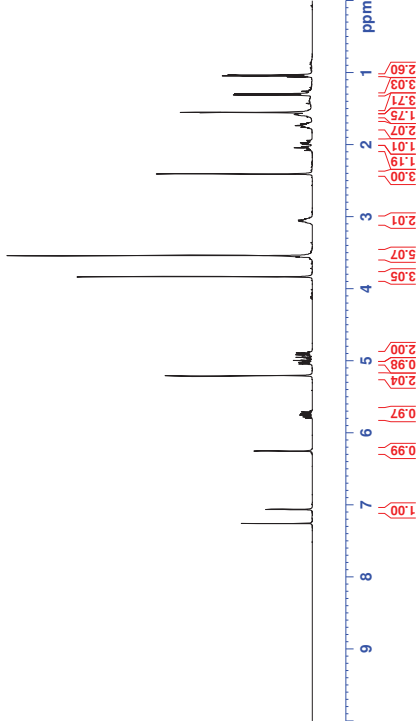
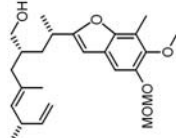
Compound 92



NAME: THMA D598
EXPNO: 0
PROCNO: 1
Date_: 20081014
Time: 21:00
INSTRUM: AVANCE400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 1H
P1: 13.50 usec
PL1: 0.00 dB
FLLW: 15.28361320 W
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
GB: 0
PC: 1.00

Compound Ref1

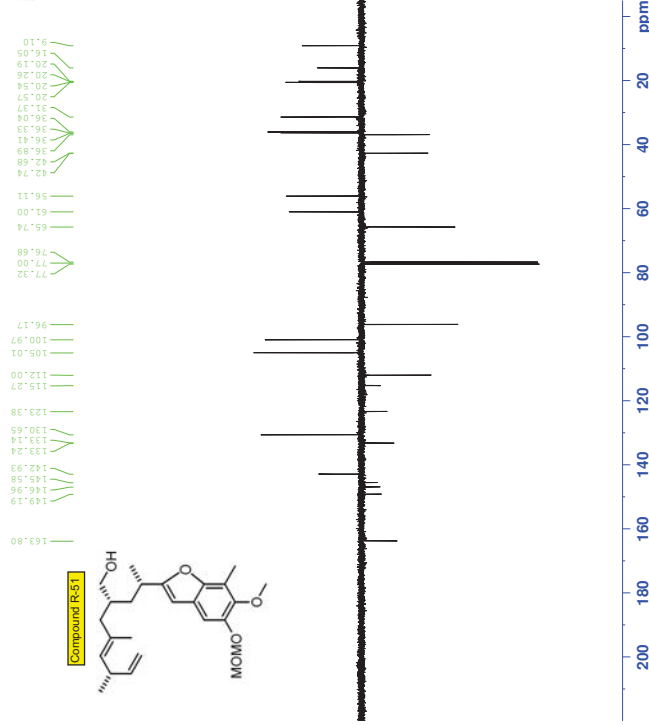
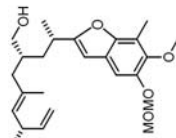


NAME: THMA D598
EXPNO: 1
PROCNO: 1
Date_: 20081014
Time: 21:00
INSTRUM: AVANCE400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1200
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.3074052 sec
RG: 655.5
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.0000000 sec
D2: 0.0068955 sec
DELTA: 0.00001311 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 13C
P1: 10.30 usec
PL1: 0.00 dB
PL2: 20.40 usec
PL3: -1.00 dB
SF01: 100.623329 MHz

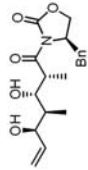
===== CHANNEL f2 =====
CPDPRG2: waltz16
NUC2: 1H
PCPD2: 100.00 usec
PL2: -3.00 dB
PL3: 20.40 usec
SF02: 400.1316005 MHz
SI: 32768
WDW: EM
SSB: 0
GB: 0
PC: 1.40

Compound Ref1



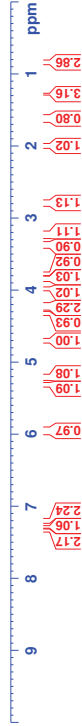


Compound 101

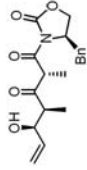


NAME THMA D497
EXPNO 40
PROCNO 1
Date_ 20080813
Time 18:09
INSTRUM avance600
PROBHD 5 mm PAQNP Sx1
PULPROG zgpg30
TD0 6550
SOLVENT CDCl3
NS 16
DS 2
SWH 12376.237 Hz
FIDRES 0.188846 Hz
AQ 2.6777449 sec
RG 327.68
WDW 600.1337060 MHz
SSB 0
LB 0.00 Hz
GB 0
PC 1.00
TD0 1

CHANNEL f1 1H
NUC1 1H
P1 13.85 usec
PL1 0.00 dB
SF01 600.1337060 MHz
SI 32768
WDW 600.1330174 MHz
SSB 0
LB 0.00 Hz
GB 0
PC 1.00
TD0 1



Compound 83

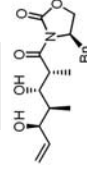


NAME THMA D495
EXPNO 10
PROCNO 1
Date_ 20080813
Time 5:42
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD0 6550
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 327.68
WDW 400.1299584 MHz
SSB 0
LB 0.30 Hz
GB 0
PC 1.00
TD0 1

CHANNEL f1 1H
NUC1 1H
P1 8.75 usec
PL1 0.00 dB
SF01 400.1324710 MHz
SI 32768
WDW 400.1299584 MHz
SSB 0
LB 0.30 Hz
GB 0
PC 1.00
TD0 1



Compound 101

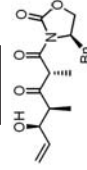


NAME THMA D497
EXPNO 40
PROCNO 1
Date_ 20080813
Time 18:09
INSTRUM avance600
PROBHD 5 mm PAQNP Sx1
PULPROG zgpg30
TD0 6550
SOLVENT CDCl3
NS 16
DS 2
SWH 35971.223 Hz
FIDRES 0.548877 Hz
AQ 2.6777449 sec
RG 327.68
WDW 600.1337060 MHz
SSB 0
LB 0.00 Hz
GB 0
PC 1.00
TD0 1

CHANNEL f1 1H
NUC1 1H
P1 13.85 usec
PL1 0.00 dB
SF01 600.1337060 MHz
SI 32768
WDW 600.1330174 MHz
SSB 0
LB 0.00 Hz
GB 0
PC 1.00
TD0 1



Compound 83



NAME THMA D495
EXPNO 10
PROCNO 1
Date_ 20080813
Time 5:42
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD0 6550
SOLVENT CDCl3
NS 16
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 3.9584243 sec
RG 327.68
WDW 400.1299584 MHz
SSB 0
LB 0.30 Hz
GB 0
PC 1.00
TD0 1

CHANNEL f1 1H
NUC1 1H
P1 8.75 usec
PL1 0.00 dB
SF01 400.1324710 MHz
SI 32768
WDW 400.1299584 MHz
SSB 0
LB 0.30 Hz
GB 0
PC 1.00
TD0 1

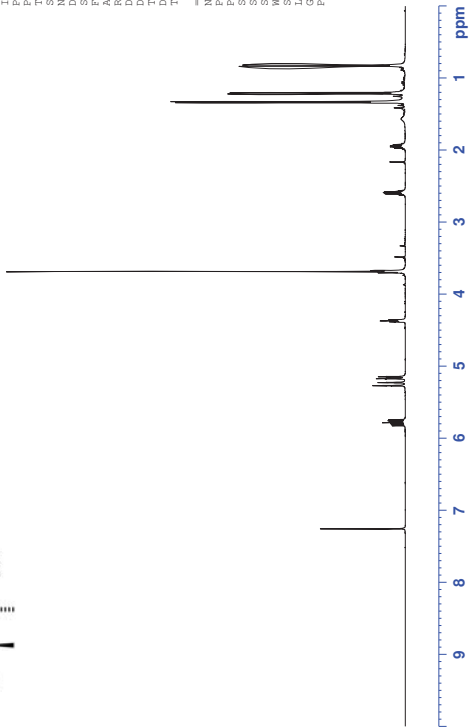
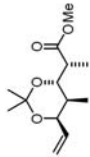




NAME: THMA D500
EXPNO: 80
PROCNO: 1
Date_: 20080818
Time: 19:54
INSTRUM: avance400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.00000000 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 1H
P1: 8.75 usec
PL1: -1.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00

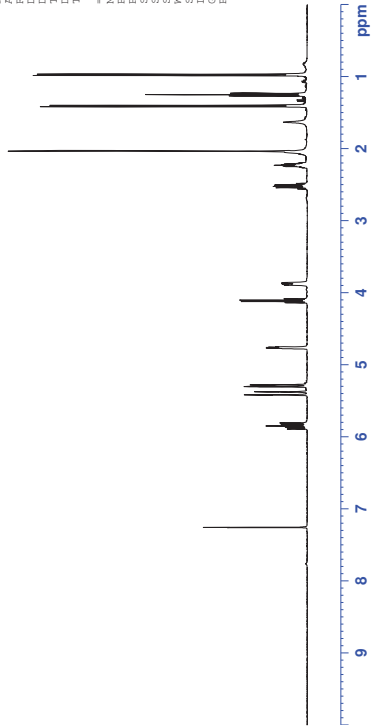
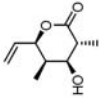
Compound 55



NAME: THMA D499
EXPNO: 231
PROCNO: 1
Date_: 20080815
Time: 8:20
INSTRUM: avance400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.126314 Hz
AQ: 3.9584243 sec
RG: 655.5
DW: 60.400 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.00000000 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 1H
P1: 8.75 usec
PL1: -1.00 dB
SF01: 400.1324710 MHz
SI: 32768
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00

Compound 54

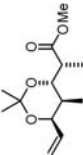


NAME: THMA D500
EXPNO: 80
PROCNO: 1
Date_: 20080818
Time: 19:54
INSTRUM: avance400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1000
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.3074052 sec
RG: 655.5
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.00000000 sec
D20: 0.00689355 sec
DELTA: 0.00001311 sec
TD0: 1

===== CHANNEL f1 =====
NUC1: 13C
P1: 10.00 usec
PL1: -1.00 dB
SF01: 100.6233329 MHz

===== CHANNEL f2 =====
CPDPRG2: waltz16
NUC2: 1H
P2: 20.60 usec
PL2: -3.00 dB
PL12: 100.00 usec
SI: 400.1316005 MHz
WDW: EM
SSB: 0
LB: 1.00 Hz
GB: 0
PC: 1.40

Compound 55

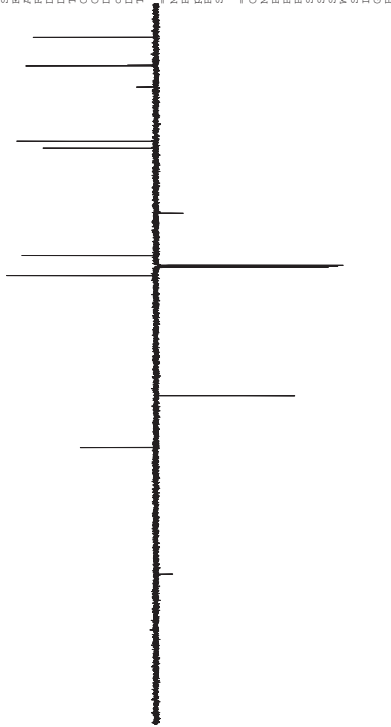
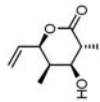


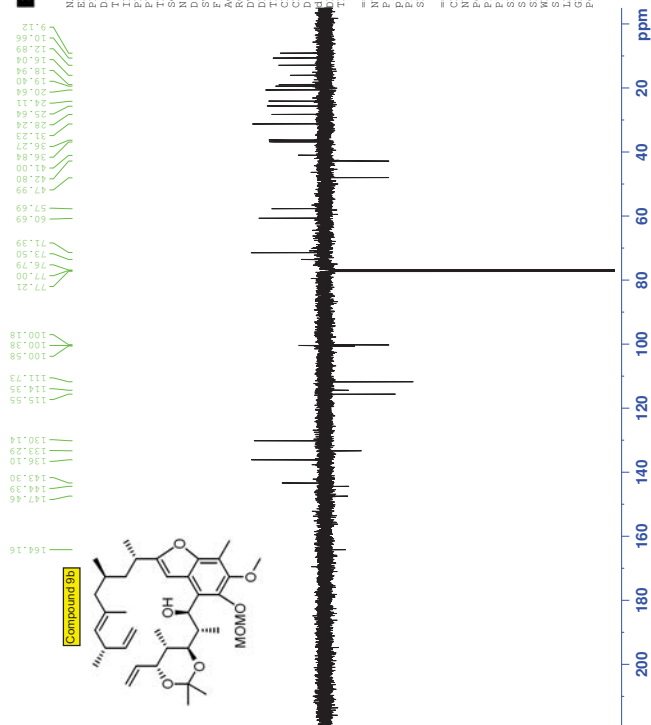
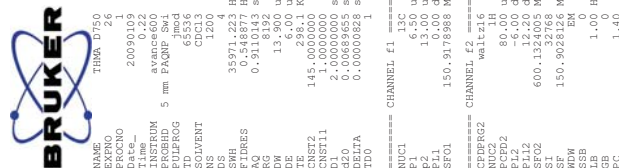
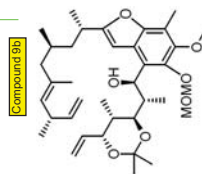
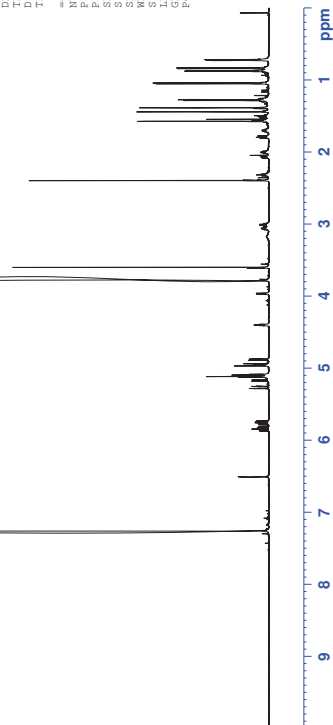
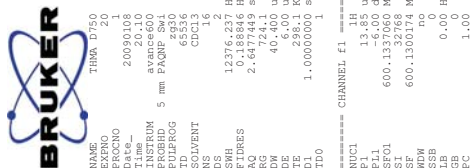
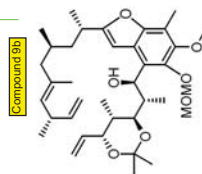
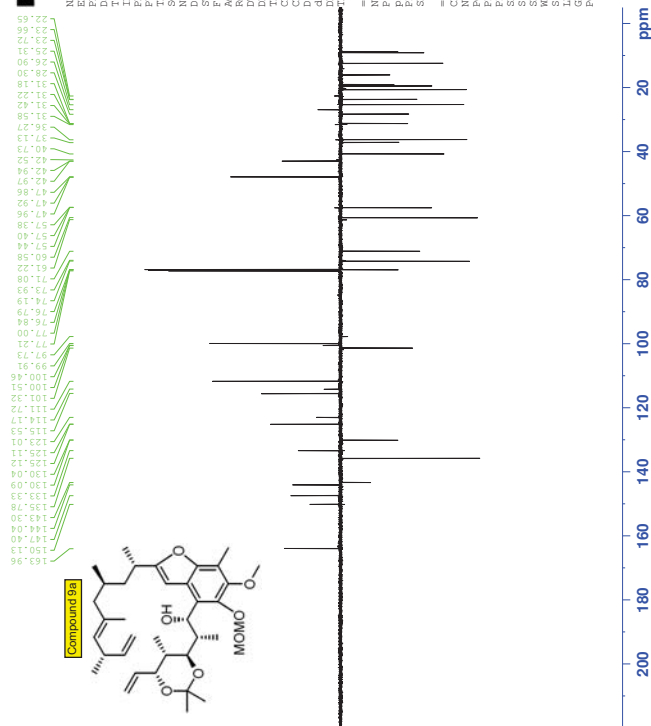
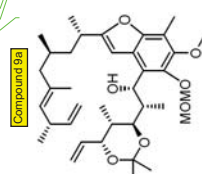
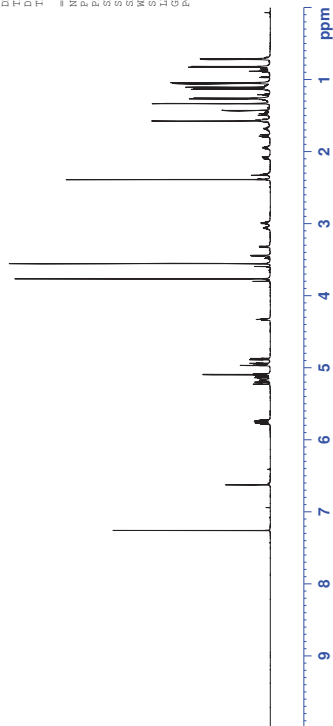
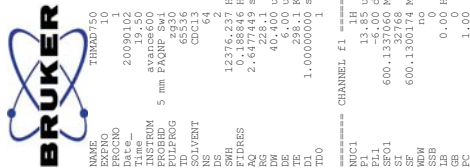
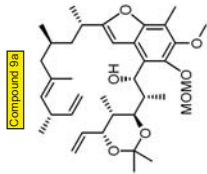
NAME: THMA D499
EXPNO: 230
PROCNO: 1
Date_: 20080815
Time: 8:20
INSTRUM: avance400
PROBHD: 5 mm BBO BB-1H
PULPROG: zgpg30
SOLVENT: CDCl3
NS: 1500
DS: 2
SWH: 25062.656 Hz
FIDRES: 0.382426 Hz
AQ: 1.3074052 sec
RG: 655.5
DW: 19.950 usec
DE: 6.00 usec
TE: 300.2 K
D1: 1.00000000 sec
D20: 0.00689355 sec
DELTA: 0.00001311 sec
TD0: 1

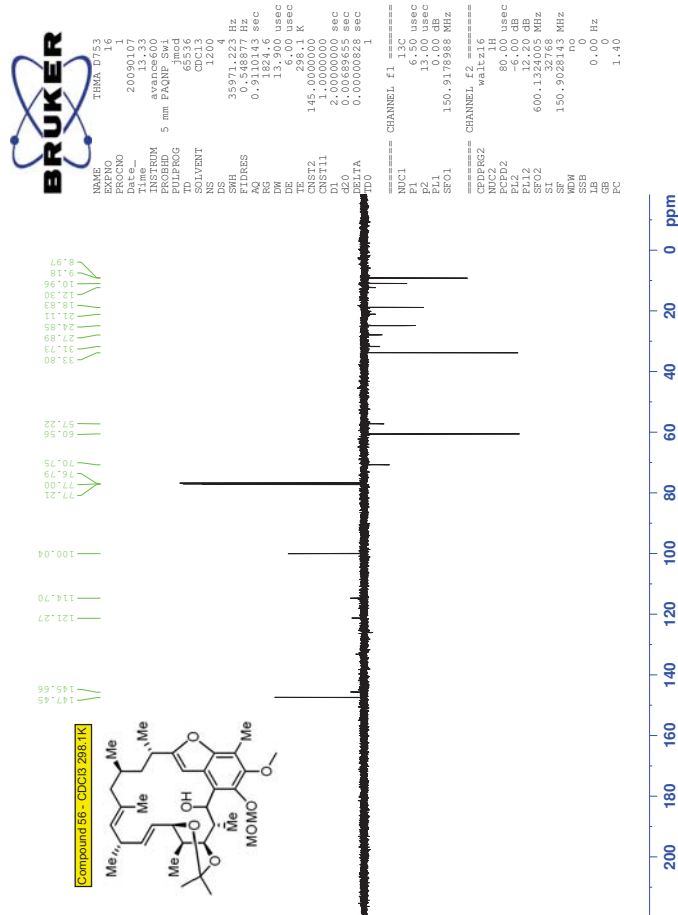
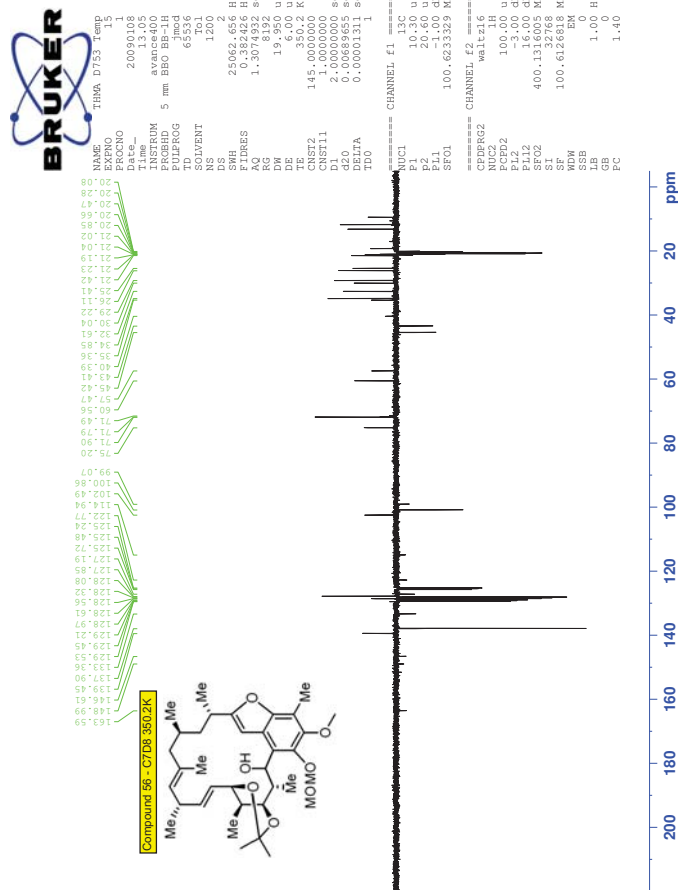
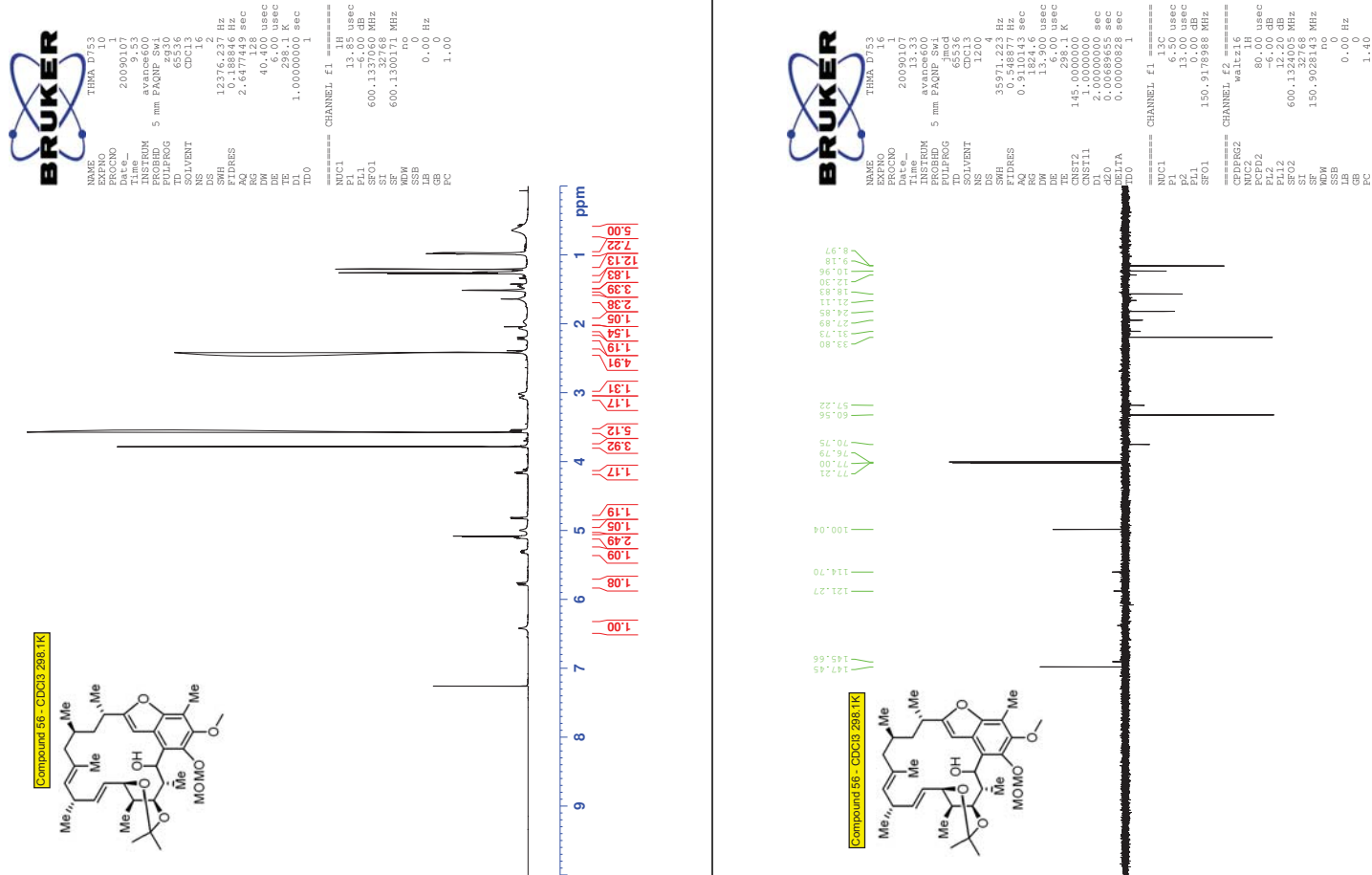
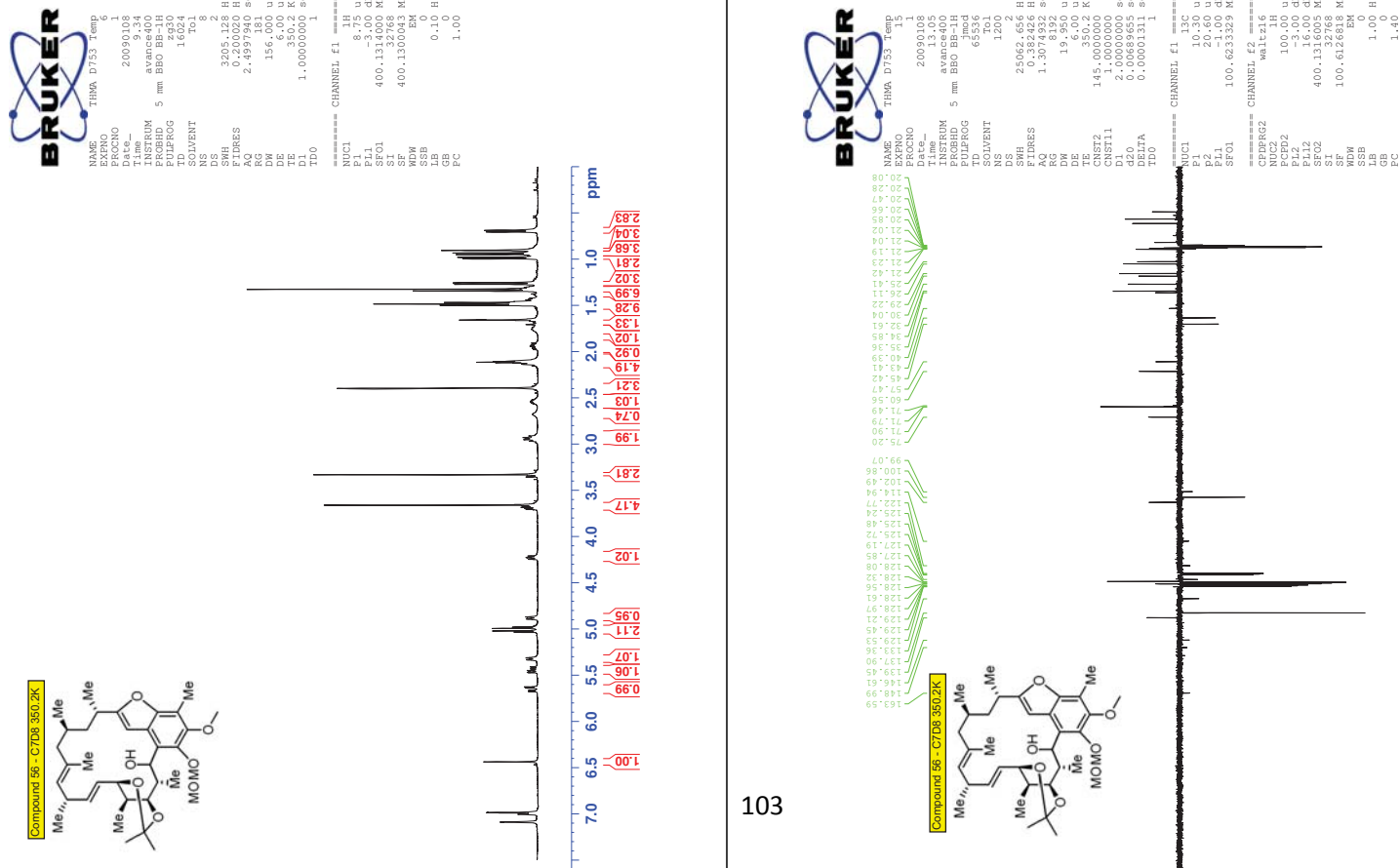
===== CHANNEL f1 =====
NUC1: 13C
P1: 10.00 usec
PL1: -1.00 dB
SF01: 100.6233329 MHz

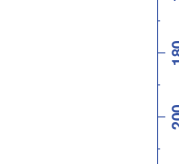
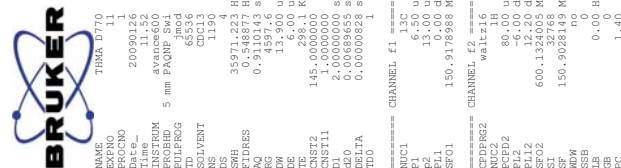
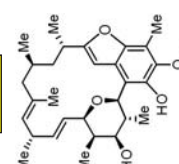
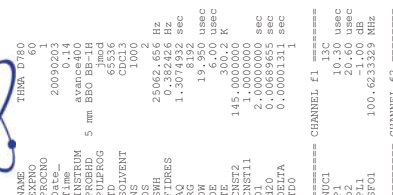
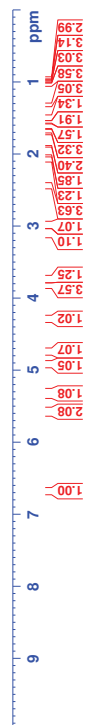
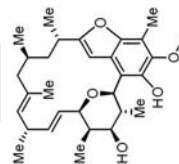
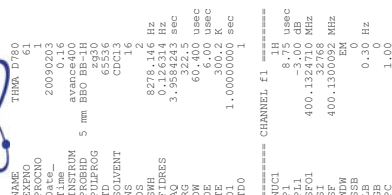
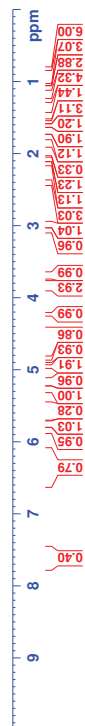
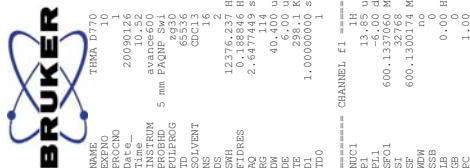
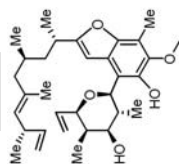
===== CHANNEL f2 =====
CPDPRG2: waltz16
NUC2: 1H
P2: 20.60 usec
PL2: -3.00 dB
PL12: 100.00 usec
SI: 400.1316005 MHz
WDW: EM
SSB: 0
LB: 1.00 Hz
GB: 0
PC: 1.40

Compound 54

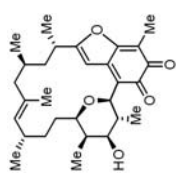






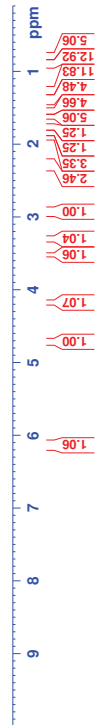


Compound 7i

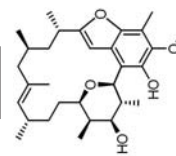


NAME THMA D781
EXPNO 260
PROCNO 1
Date_ 20090202
Time 15:07
INSTRUM AVI1107
PROBHD 5 mm PA100 BB-
PULPROG zgpg30
TD 6550
SOLVENT CDCl3
NS 4
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 320.000
DW 60.800 usec
DE 2.50 usec
TE 300.2 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.50 usec
PL1 0.00 dB
F1W 15.28561320 W
SFO1 400.2724718 MHz
SF 400.2724718 MHz
WDW EM
SSB 0
GB 0
PC 1.00

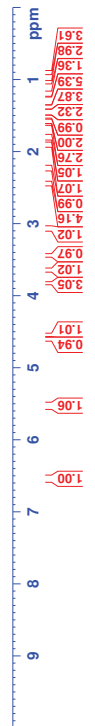


Compound 3

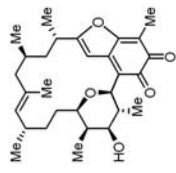


NAME THMA D725
EXPNO 1
PROCNO 1
Date_ 20081209
Time 9:43
INSTRUM avnc400
PROBHD 5 mm PAQNP SW1
PULPROG zgpg30
TD 2930
SOLVENT CDCl3
NS 16
DS 2
SWH 12376.237 Hz
FIDRES 0.188846 Hz
AQ 2.6477449 sec
RG 320.000
DW 40.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.65 usec
PL1 0.00 dB
F1W 600.1337060 MHz
SFO1 600.1337060 MHz
SF 600.1337060 MHz
WDW EM
SSB 0
GB 0
PC 1.00



Compound 7i



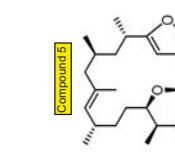
NAME THMA D778
EXPNO 106
PROCNO 1
Date_ 20090130
Time 15:25
INSTRUM avnc400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 6550
SOLVENT CDCl3
NS 5000
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.372982 sec
RG 320.000
DW 19.950 usec
DE 2.50 usec
TE 300.2 K
CNST2 145.0000000 K
CNST11 1.0000000
DELTA 0.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 13.00 usec
PL1 0.00 dB
F1W 100.623329 MHz
SFO1 100.623329 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
P2 13.00 usec
PL2 -3.00 dB
F2W 400.1316005 MHz
SFO2 400.1316005 MHz
SF 400.1316005 MHz
WDW EM
SSB 0
GB 0
PC 1.10



Compound 3

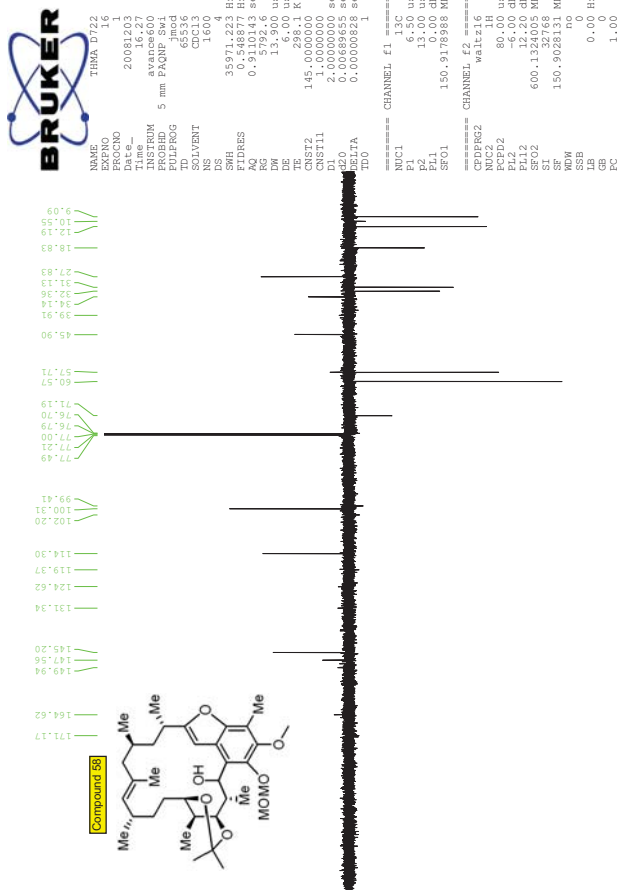
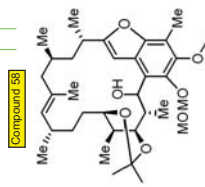
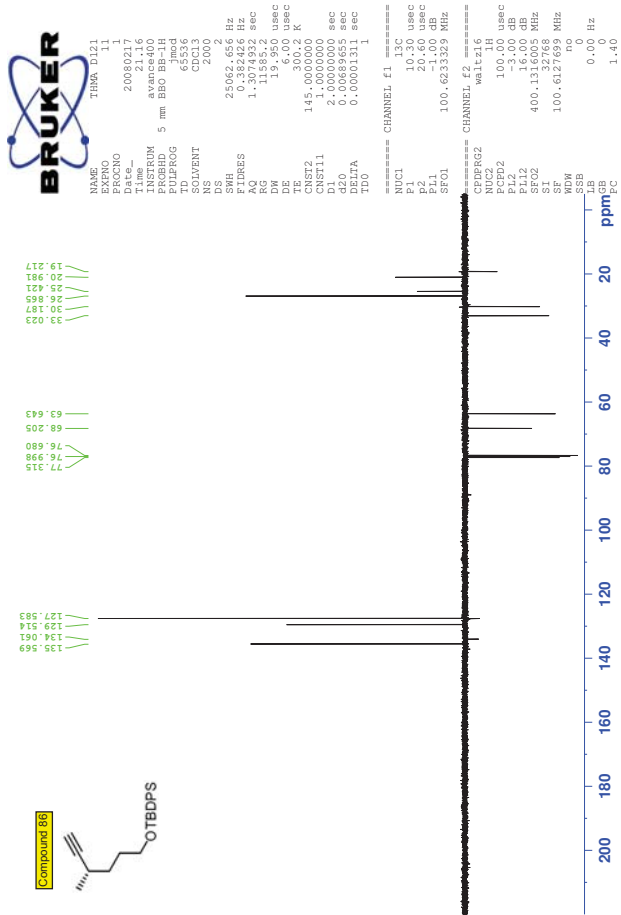
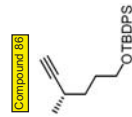
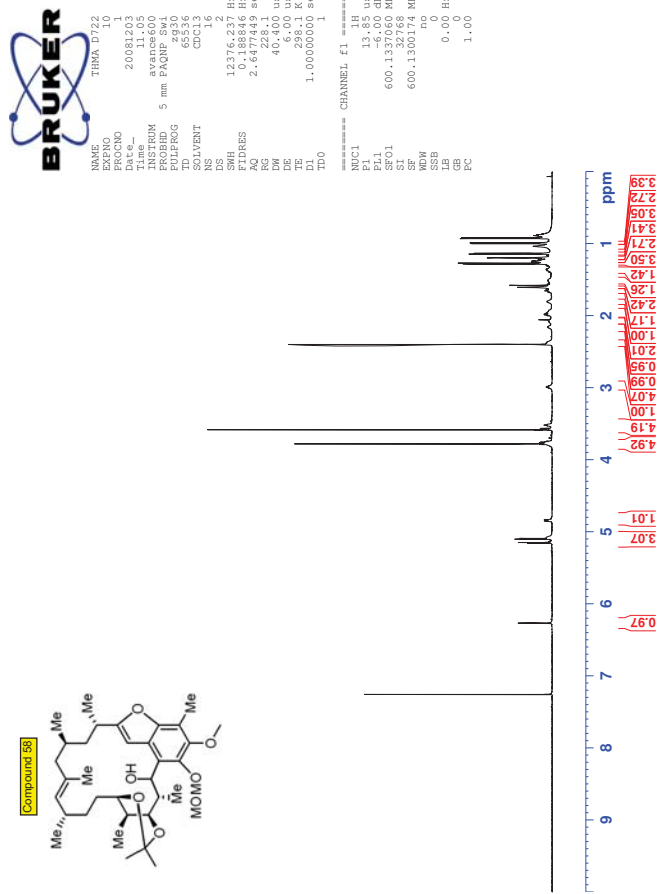
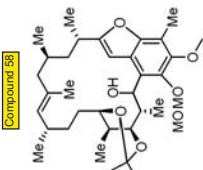
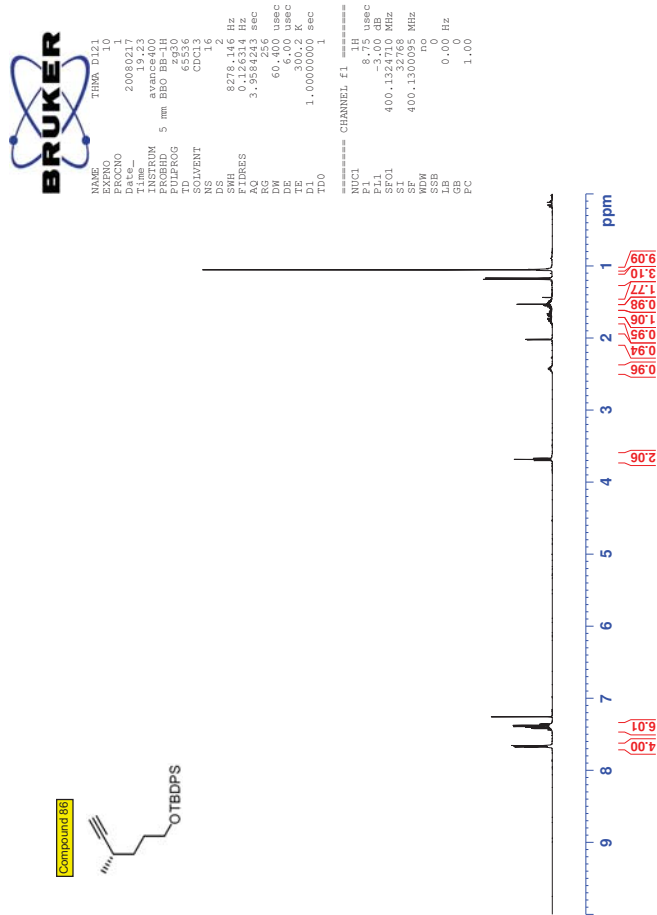
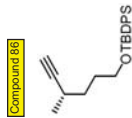


NAME THMA D725
EXPNO 1
PROCNO 1
Date_ 20081209
Time 9:43
INSTRUM avnc400
PROBHD 5 mm PAQNP SW1
PULPROG zgpg30
TD 2930
SOLVENT CDCl3
NS 2000
DS 2
SWH 35971.223 Hz
FIDRES 0.548877 Hz
AQ 0.919443 sec
RG 320.000
DW 13.900 usec
DE 6.00 usec
TE 300.2 K
CNST2 145.0000000 K
CNST11 1.0000000
DELTA 0.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 13.00 usec
PL1 0.00 dB
F1W 130.9178988 MHz
SFO1 130.9178988 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
P2 13.00 usec
PL2 -6.00 dB
F2W 600.1324005 MHz
SFO2 600.1324005 MHz
SF 600.1324005 MHz
WDW EM
SSB 0
GB 0
PC 1.40



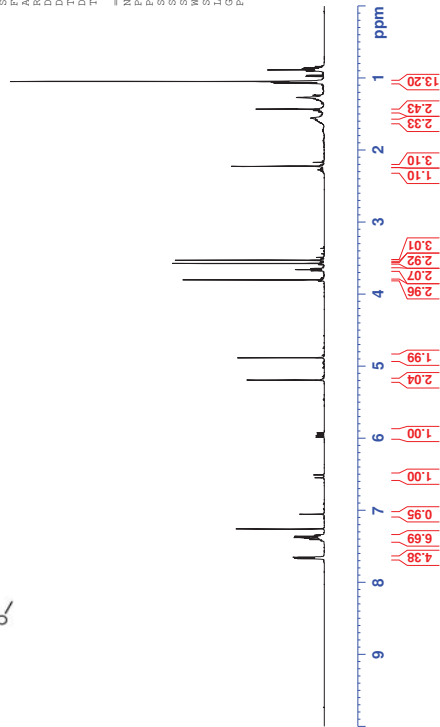
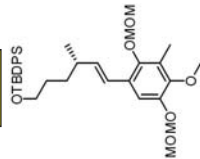




NAME THMA D370
EXPNO 1
PROCNO 1
Date_ 20080620
Time 13.44
INSTRUM avac400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD0 6520
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 327.68
DW 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 -2.00 dB
SF01 400.1324710 MHz
SI 32768
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

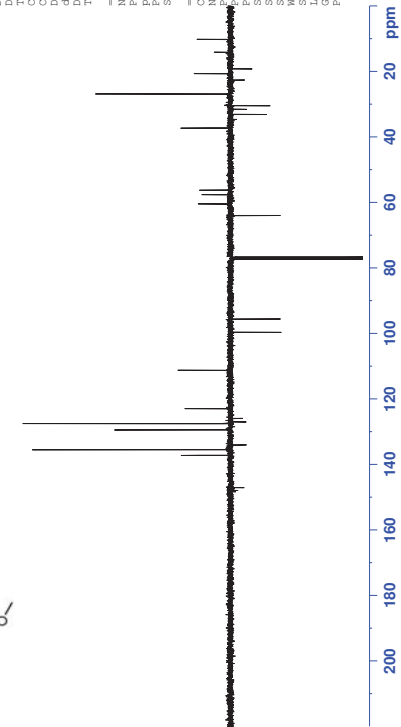
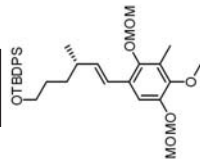
Compound 62



NAME THMA D370
EXPNO 1
PROCNO 1
Date_ 20080620
Time 13.44
INSTRUM avac400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD0 6520
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 327.68
DW 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 -2.00 dB
SF01 400.1324710 MHz
SI 32768
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

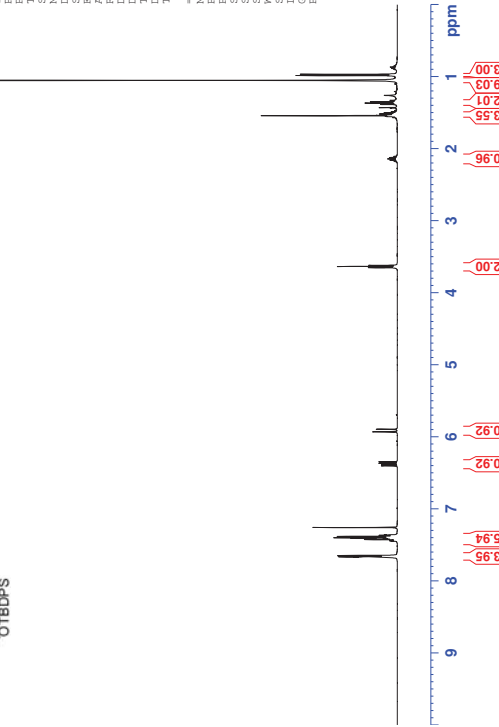
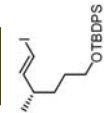
Compound 62



NAME THMA D436
EXPNO 1
PROCNO 1
Date_ 20080722
Time 19.47
INSTRUM avac400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD0 6520
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 327.68
DW 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 -2.00 dB
SF01 400.1324710 MHz
SI 32768
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

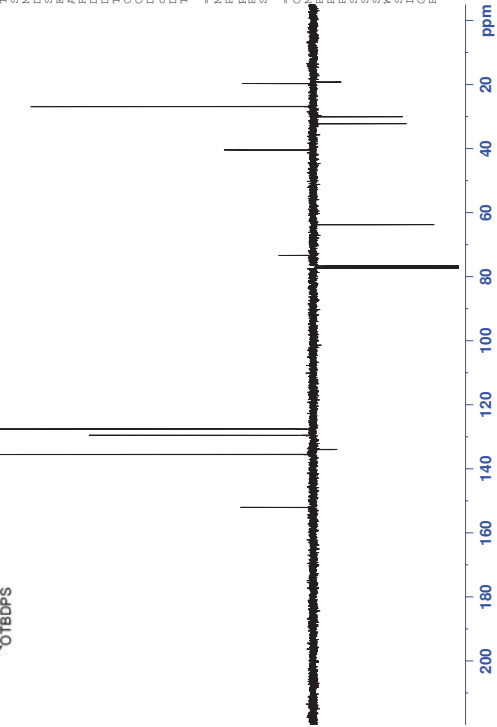
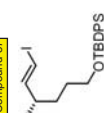
Compound 67

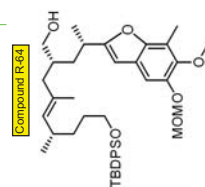
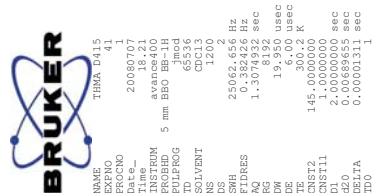
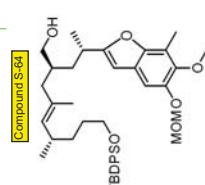
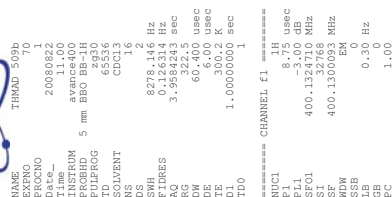
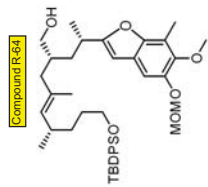
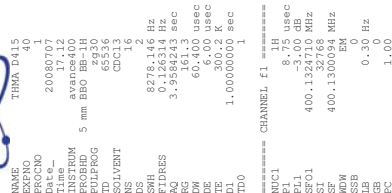
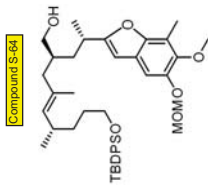


NAME THMA D436
EXPNO 1
PROCNO 1
Date_ 20080722
Time 19.47
INSTRUM avac400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD0 6520
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 327.68
DW 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 -2.00 dB
SF01 400.1324710 MHz
SI 32768
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

Compound 67

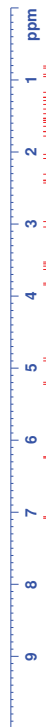
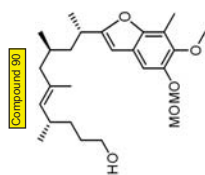






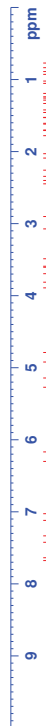
NAME THMA D491
PROCNO 51
Date_ 20080812
Time 12:03
INSTRUM avn600
PROBHD 5 mm PAQNP Sx1
PULPROG zgpg30
SOLVENT CDCl3
NS 16
DS 2
SWH 12376.237 Hz
FIDRES 0.188846 Hz
AQ 2.477749 sec
RG 640.000
DW 40.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.85 usec
PL1 0.00 dB
SF01 600.1337060 MHz
SI 32768
WDW 600.1301174 MHz
SSB 0
LB 0
GB 0
PC 1.00



NAME THMA D419
PROCNO 1
Date_ 20080708
Time 17:21
INSTRUM avn600
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 640.000
DW 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 0.00 dB
SF01 400.1324710 MHz
SI 32768
WDW 400.1300053 MHz
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

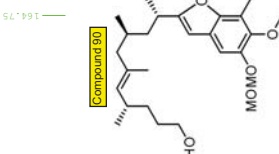


NAME THMA D491
PROCNO 51
Date_ 20080812
Time 12:03
INSTRUM avn600
PROBHD 5 mm PAQNP Sx1
PULPROG zgpg30
SOLVENT CDCl3
NS 1200
DS 2
SWH 35971.223 Hz
FIDRES 0.548877 Hz
AQ 0.921048 sec
RG 640.000
DW 13.900 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
D20 0.00689455 sec
DELTA 0.0000028 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 13.00 usec
PL1 0.00 dB
SF01 150.9179988 MHz

===== CHANNEL f2 =====
NAME waltz16
PROCNO 51
Date_ 20080812
Time 12:03
INSTRUM avn600
PROBHD 5 mm PAQNP Sx1
PULPROG zgpg30
SOLVENT CDCl3
NS 1200
DS 2
SWH 35971.223 Hz
FIDRES 0.548877 Hz
AQ 0.921048 sec
RG 640.000
DW 13.900 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
D20 0.00689455 sec
DELTA 0.0000028 sec
TD0 1

===== CHANNEL f2 =====
NAME waltz16
PROCNO 51
Date_ 20080812
Time 12:03
INSTRUM avn600
PROBHD 5 mm PAQNP Sx1
PULPROG zgpg30
SOLVENT CDCl3
NS 1200
DS 2
SWH 35971.223 Hz
FIDRES 0.548877 Hz
AQ 0.921048 sec
RG 640.000
DW 13.900 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
D20 0.00689455 sec
DELTA 0.0000028 sec
TD0 1



NAME THMA D419
PROCNO 1
Date_ 20080708
Time 17:21
INSTRUM avn600
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
SOLVENT CDCl3
NS 1200
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3058932 sec
RG 640.000
DW 19.950 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
D20 0.00689455 sec
DELTA 0.0000131 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 13.00 usec
PL1 0.00 dB
SF01 100.6233329 MHz

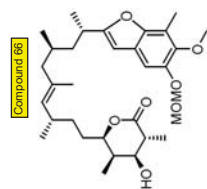
===== CHANNEL f2 =====
NAME waltz16
PROCNO 1
Date_ 20080708
Time 17:21
INSTRUM avn600
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
SOLVENT CDCl3
NS 1200
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3058932 sec
RG 640.000
DW 19.950 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
D20 0.00689455 sec
DELTA 0.0000131 sec
TD0 1

===== CHANNEL f2 =====
NAME waltz16
PROCNO 1
Date_ 20080708
Time 17:21
INSTRUM avn600
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
SOLVENT CDCl3
NS 1200
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3058932 sec
RG 640.000
DW 19.950 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
D20 0.00689455 sec
DELTA 0.0000131 sec
TD0 1



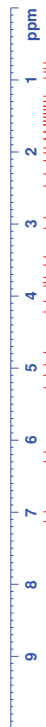


Compound 66

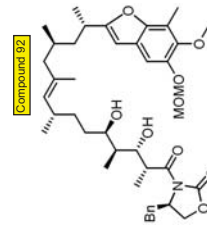


NAME THMA D597
PROCNO 31
Date_ 20081010
Time_ 12.45
INSTRUM avac400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
SOLVENT CDCl3
NS 32
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
TD0 1

CHANNEL f1 1H
NUC1 1H
P1 8.75 usec
PL1 -1.00 dB
SFO1 400.1324710 MHz
SI 32768
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

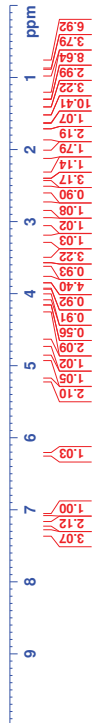


Compound 92

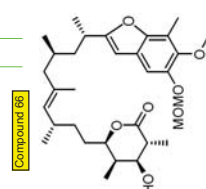


NAME THMA D593
PROCNO 1
Date_ 20081009
Time_ 10.43
INSTRUM avac400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
TD0 1

CHANNEL f1 1H
NUC1 1H
P1 8.75 usec
PL1 -1.00 dB
SFO1 400.1324710 MHz
SI 32768
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Compound 66

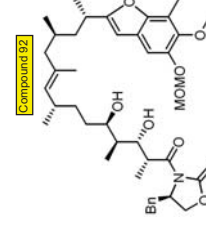


NAME THMA D597
PROCNO 31
Date_ 20081010
Time_ 12.45
INSTRUM avac400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
SOLVENT CDCl3
NS 1493
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3079322 sec
RG 19.950 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
D2 0.00689355 sec
DELTA 0.00001311 sec
TD0 1

CHANNEL f1 1H
NUC1 1H
P1 8.75 usec
PL1 -1.00 dB
SFO1 400.1324710 MHz
SI 32768
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Compound 92



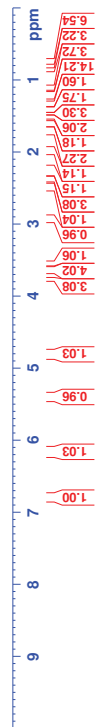
NAME THMA D593
PROCNO 1
Date_ 20081009
Time_ 10.43
INSTRUM avac400
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
SOLVENT CDCl3
NS 2155
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3079322 sec
RG 19.950 usec
DE 6.00 usec
TE 300.2 K
D1 1.0000000 sec
D2 0.00689355 sec
DELTA 0.00001311 sec
TD0 1

CHANNEL f1 1H
NUC1 1H
P1 8.75 usec
PL1 -1.00 dB
SFO1 400.1324710 MHz
SI 32768
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

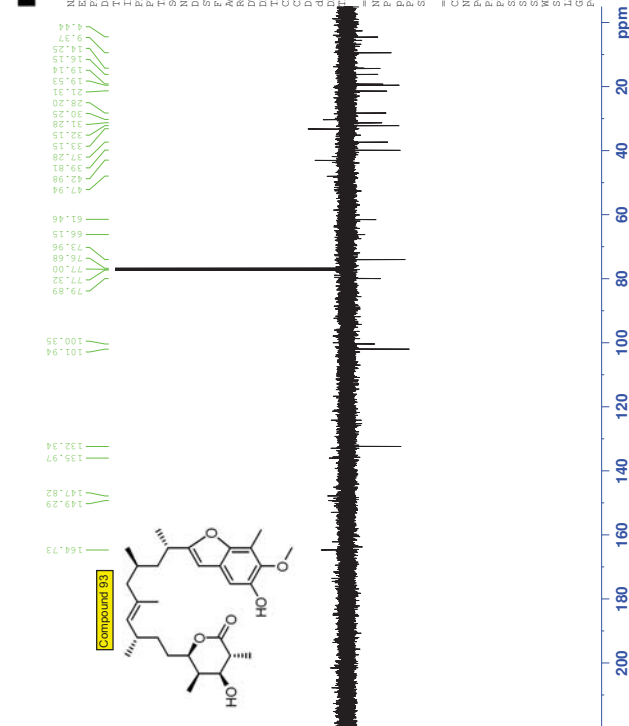




| NUC1 | CHANNEL f1 | IH |
|-------|-------------|------|
| P1 | 13.50 | usec |
| PL1 | -1.80 | dB |
| PL1W | 15.28361320 | W |
| SSFO1 | 400.2724718 | MHz |
| SI | 32768 | |
| SF | 400.2700325 | MHz |
| WDW | EM | |
| SSB | 0 | |
| LB | 0.30 | Hz |
| GB | 0 | |
| PC | 1.00 | |

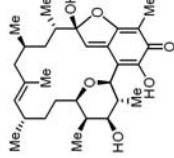
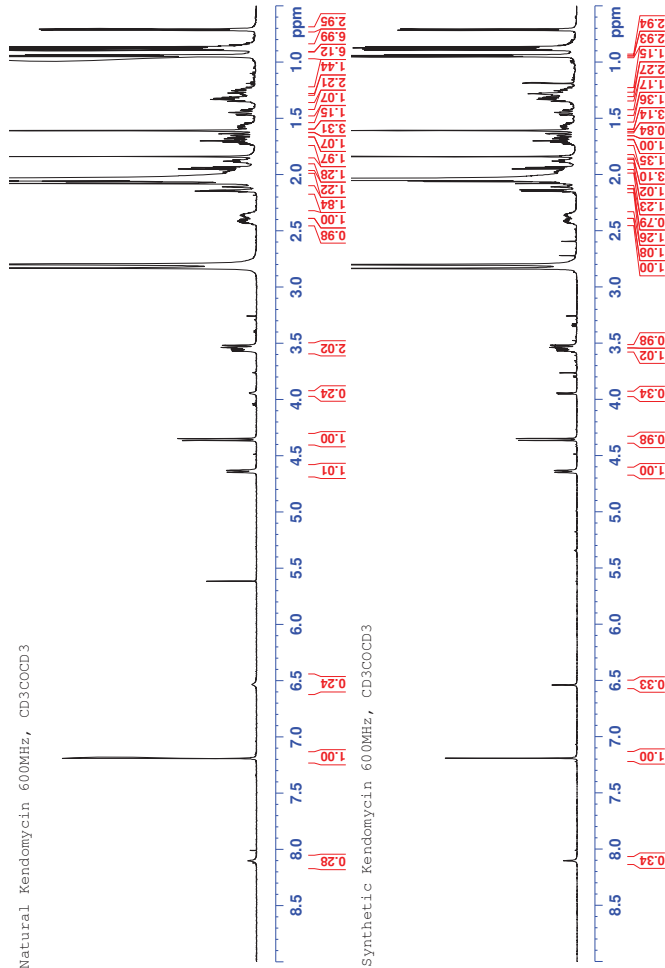


```
===== CHANNEL f1 =====
NUC1      13C
P1        10.30 usec
p2        20.60 usec
PL1       -1.00 dB
SF01      100.6233329 MHz
```

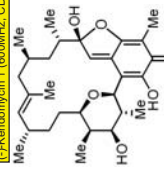
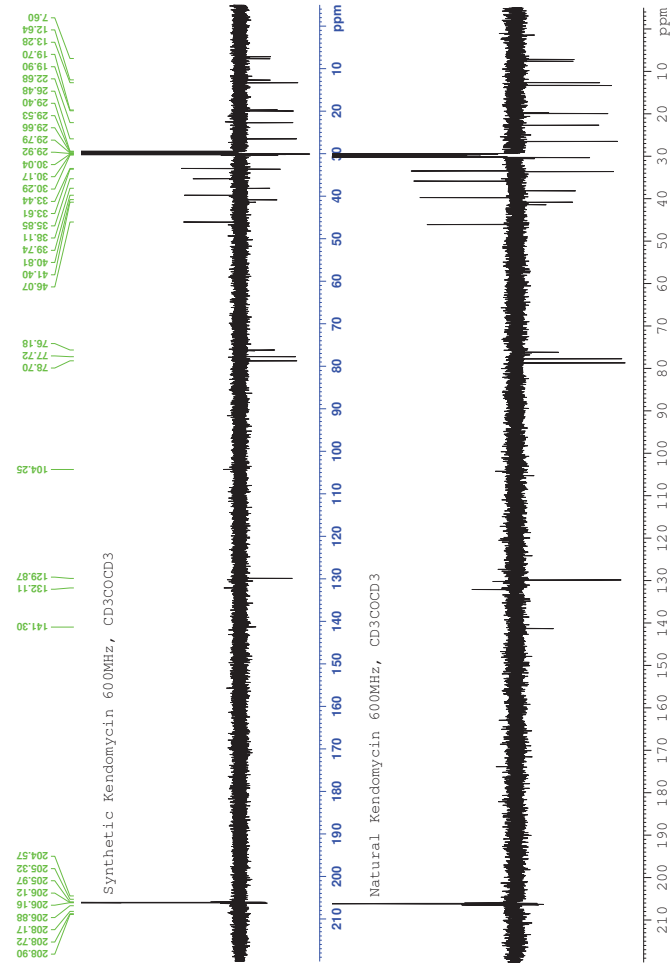
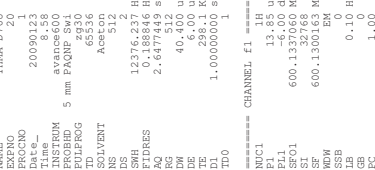
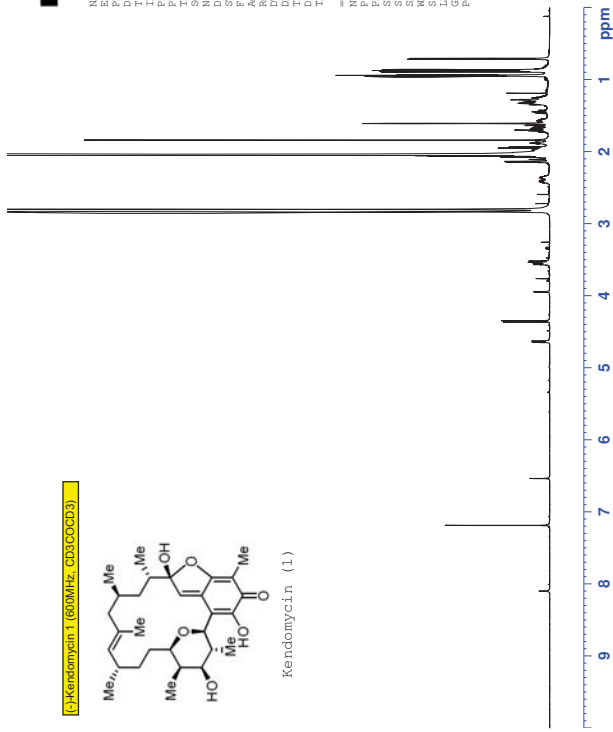


| | | |
|------|-----------------|-----------|
| | CHANNEL f1 | 1H |
| NUC1 | | 8.75 usec |
| P1 | | -3.00 dB |
| PL1 | | |
| SFO1 | 400.1324710 MHz | |
| S1 | 32768 | |
| SF | 400.1300094 MHz | |
| MW | n | |
| SSB | o | |
| LB | 0.00 Hz | |
| GB | 0 | |
| PC | 1.00 | |

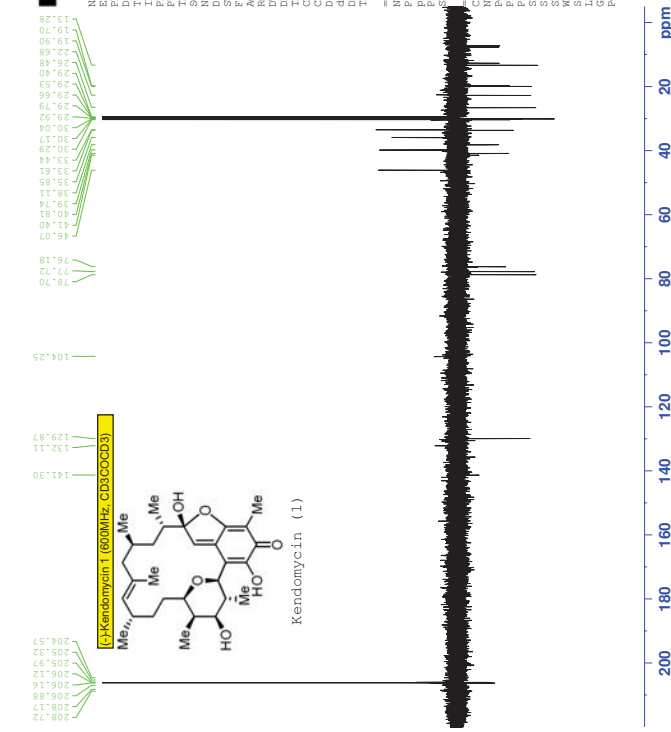




Kendomycin (1)



Kendomycin (1)



4. Summary

In summary, we have accomplished two convergent enantioselective syntheses of the *ansa*-macrolide kendomycin. The routes developed during this dissertation start from common and inexpensive starting materials such as β -(+)-citronellene, 2,6-dimethoxytoluene and (meth)acrolein. The outstanding key step for both, the photo-Fries and the RCM approach, are the highly complex Ireland-Claisen rearrangements. High efficiency and diastereoselectivity were achieved after some optimization work. In this manner, the installation of the north-eastern domain, with concomitant formation of the (*E*)-double bond, and the desired C16 stereocenter was achieved in one step. For the preparation of the eastern benzofuran moiety two successful strategies, a Pd(0)-mediated Negishi coupling and an epoxide opening reaction, were pursued. The incorporation of the underestimated photo-Fries reaction in total synthesis demonstrates its power and efficiency. The drawback of the latter approach is the high number of linear steps (32 steps from 2,6-dimethoxytoluene) and an overall yield of 0.4%, which is, however, comparable to that of Smith's total synthesis (0.5%).

In order to overcome the above drawbacks we focused on an alternative synthesis in parallel. Primarily efforts to close the macrocycle at various positions via RCM were meager. The finally developed RCM-approach, which aims at the C10-C11 disconnection, diminishes the number of steps to 24 and increases the overall yield to 1.0%. This optimized sequence also would allow for a rapid and modular synthesis of well-designed derivatives.

In contrast to Rychnovsky, we were able to reproduce Lee's final steps and disclosed the fourth total synthesis of kendomycin. The development of a protecting group free endgame for the preparation of the quinone-lactol unit saved another two steps.

PART B: Total Syntheses of (+)-Echinopine A and B

5. Introduction and Background

5.1. Overview

Terpenes belong to the most abundant natural products and contain more than 20.000 members of great diversity.²⁴ The word “terpene”, which was originally applied to hydrocarbons found in turpentine is often used in place of “terpenoid (isoprenoid)”. In a strict sense, the term “terpenoids” also include the rearranged and oxidized compounds. Terpenoids are important constituents in perfume (e.g. patchoulol, cineol) and flavouring (e.g. menthol) industry and medicine (e.g. paclitaxel).²⁵ Apart from that, terpenoids frequently serve as excellent chiral building blocks for organic synthesis²⁶ and are used for the design of catalyst ligands.

In 1959, Ruzicka²⁷ reemphasized the famous isoprene rule, which was already proposed by Wallach in 1887: All terpenoids are built up by multiple isoprenoid (2-methyl-1,3-butadiene) units. According to this, the primary classification of terpenoids is done by the number of isoprene units: C₅ hemiterpenes, C₁₀ monoterpenes, C₁₅ sesquiterpenes, C₂₀ diterpenes, C₂₅ sesterpenes, C₃₀ triterpenes, C₄₀ tetraterpenes and (C₅)_n polyterpenes. Further classification, for instance, is done with regard to the terpenoid family, which is often named after the first or most common member (e.g. guaiane-type terpenoids).

Figure 5 shows a personal selection of terpenoids, whereas the most versatile and complex natural products are derived from the C₁₅– to C₃₀–members.

²⁴ C. S. Sell, *A Fragrant Introduction to Terpenoid Chemistry* (Kindle Edition), Royal Society of Chemistry, 1st edition **2003**.

²⁵ E. Breitmaier, *Terpenes: Flavors, Fragrances, Pharmaca, Pheromones*, Wiley-VCH **2006**.

²⁶ a) T.-L. Ho, *Enantioselective Synthesis: Natural Products from Chiral Terpenes*, John Wiley & Sons, Inc., New York **1992**; b) D. J. Ager, *Handbook of Chiral Chemicals*, Taylor & Francis Group, LLC, 2nd edition **2006**, 59–72.

²⁷ L. Ruzicka, *Proc. Chem. Soc. (London)* **1959**, 341.

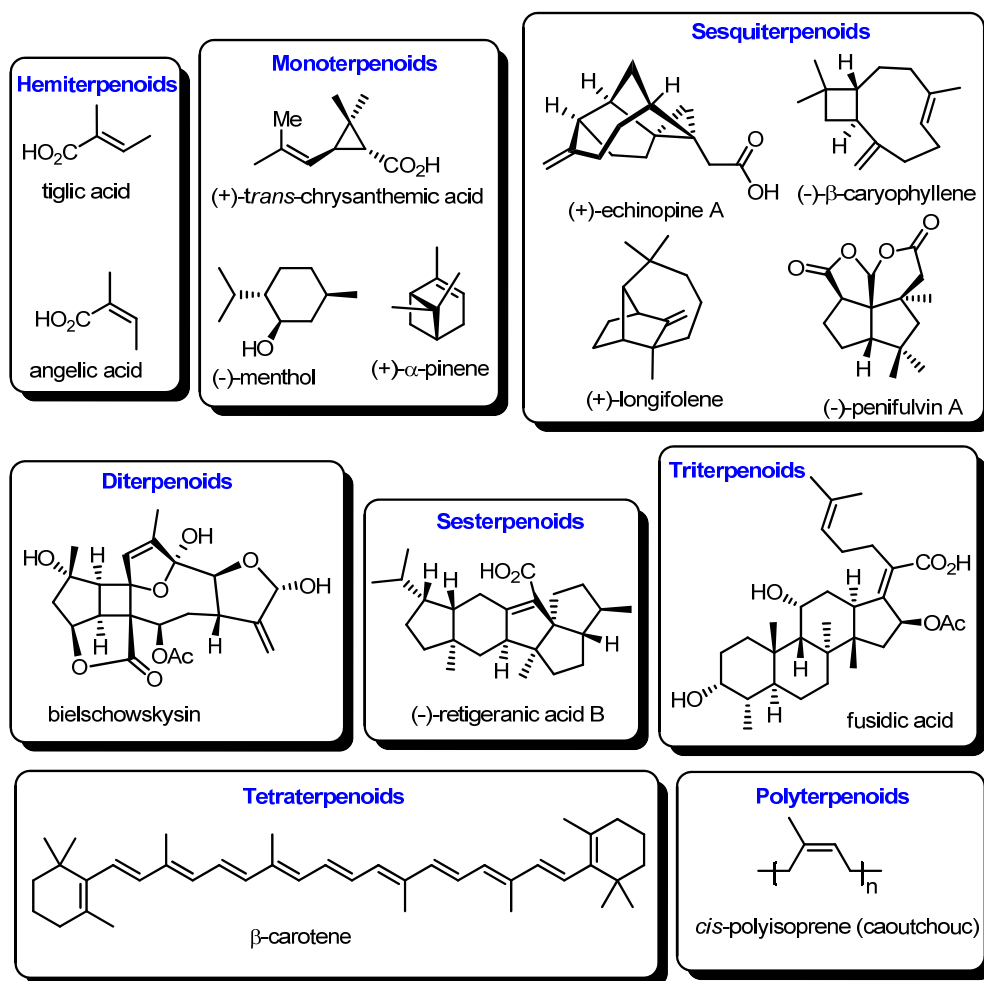


Figure 5. A selection of naturally occurring terpenoids.

5.2. Isolation, Biological Activity and Structure of Echinopine A and B

(+)-Echinopine A (**109**) and B (**110**), two novel tetracyclic sesquiterpenoids were isolated in 2008 from the roots of *Echinops spinosus* (Morocco).²⁸ Repeated solvent extraction (MeOH, CH₂Cl₂ and EtOAc), column chromatography and separation with RP-HPLC afforded 2 mg of (+)-**109** and 1.6 mg of (+)-**110** respectively. Because of the scarcity of natural product source, determination of the relative stereochemistry could only be performed by 2D-NMR and HRMS.

The X-ray structure of synthetic **109** is shown in Figure 6 and represents (-)-**109**, since structural refinements, complicated due to low intensity of reflections, were more accurate.

²⁸ M. Dong, B. Cong, S. H. Yu, F. Sauriol, C.-H. Huo, Q.-W. Shi, Y.-C. Gu, L. O. Zamir, H. Kiyota, *Org. Lett.*, **2008**, *10*, 701–704.

The densely packed 3,5,5,7-tetracycle features two vicinal quaternary centers, 5 stereocenters in total, a 1,1-disubstituted olefin unit and a carboxyl functionality.

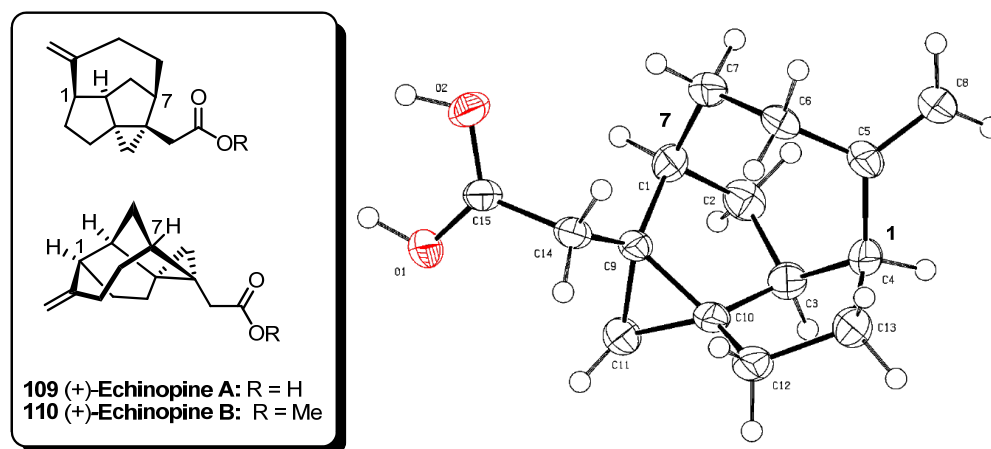


Figure 6. 2D-Views of (+)-echinopine A (**109**) and B (**110**). Confirmation of the relative structure by X-ray structure analysis of synthetic **109**.

Due to the low amounts of isolated echinopine A and B, only an *in vitro* biological test against the human breast cancer MCF-7 line could be performed, whereas both proved to be inactive. However, proper biological screening assays of the title compounds are currently pursued at the University of Hannover in corporation with Prof. Kalesse.

5.3. Biosynthesis

5.3.1. Biosynthesis of Terpenes

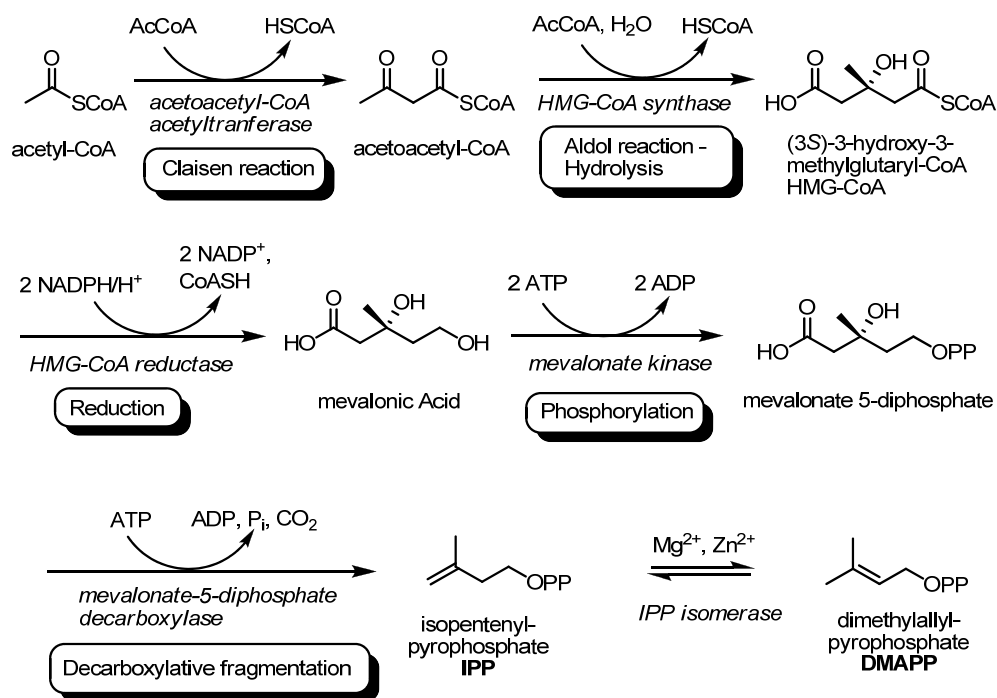
The biosynthetic origin of all terpenes is isopentenyl diphosphate (IPP), which, after isomerization to dimethylallyl diphosphate (DMAPP), is sequentially elongated by prenyltransferases to geranyl diphosphate (GPP, C₁₀), farnesyl diphosphate (FPP, C₁₅), and geranyl-geranyl diphosphate (GGP, C₂₀).²⁹ For the biosynthesis of IPP two different pathways are considered: The mevalonate (MVA)³⁰ and the Rohmer (1-deoxyxylose-5-phosphate, DXP)³¹ pathway. The former is considered for the synthesis of sesquiterpenes and triterpenes in animals and plants, the latter for monoterpenes, diterpenes, and tetraterpenes. Both pathways work in bacteria.

²⁹ F. J. Leeper, J. C. Vederas, Eds., *Biosynthesis: Aromatic Polyketides, Isoprenoids, Alkaloids* (Topics in Current Chemistry), Springer, 1st edition **2000**, 54–92.

³⁰ W. R. Nes, M. L. McKean, *Biochemistry of steroids and other isopentanooids*, University Park Press, Baltimore **1977**.

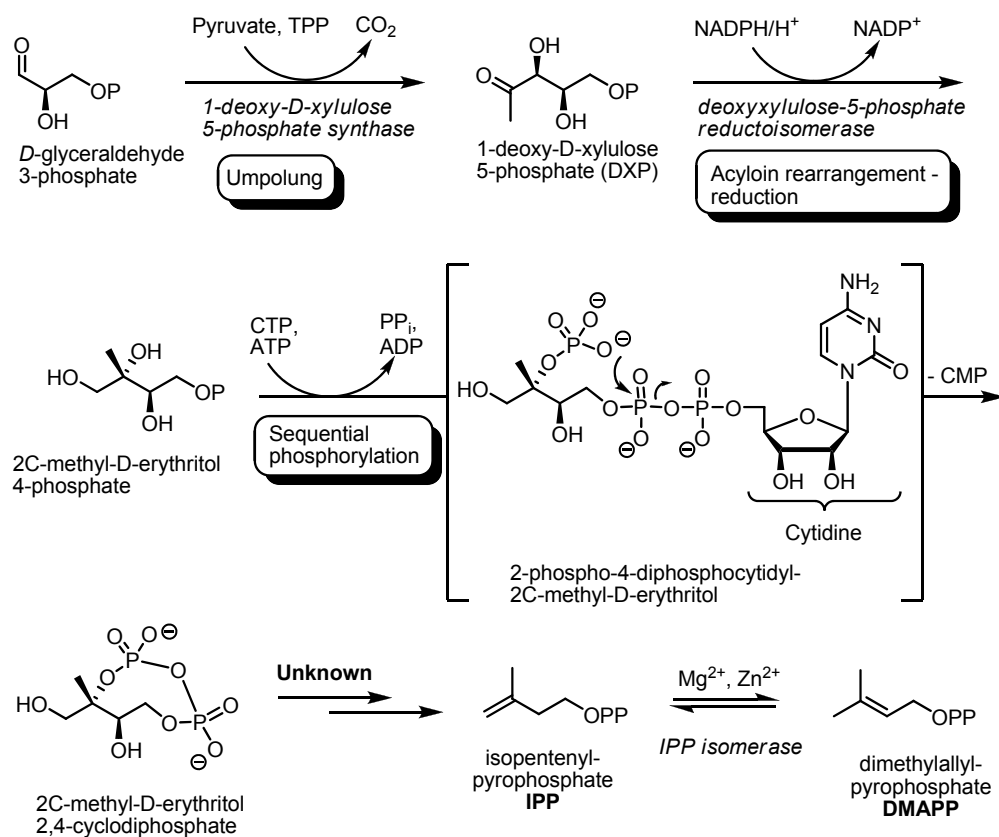
³¹ a) M. Rohmer *Nat. Prod. Rep.* **1999**, *16*, 565–574; b) M. Rohmer, M. Knani, P. Simonin, B. Sutter, H. Sahm, *Biochem J.* **1993**, *295*, 517–524.

The mevalonic pathway: The MVA pathway (Scheme 20) starts with the condensation of three molecules of acetyl CoA, first to give acetoacetyl CoA via a Claisen condensation and second to afford HMG-CoA via an aldol-like reaction. The six-carbon thioester intermediate is reduced to mevalonic acid (MVA) by NADPH/H⁺, which in turn is phosphorylated with the consumption of three molecules of ATP and converted to IPP by decarboxylative fragmentation. The next step, before elongation can take place, is the isomerization of IPP to DMAPP by IPP isomerase.



Scheme 20. The mevalonate (MVA) pathway.

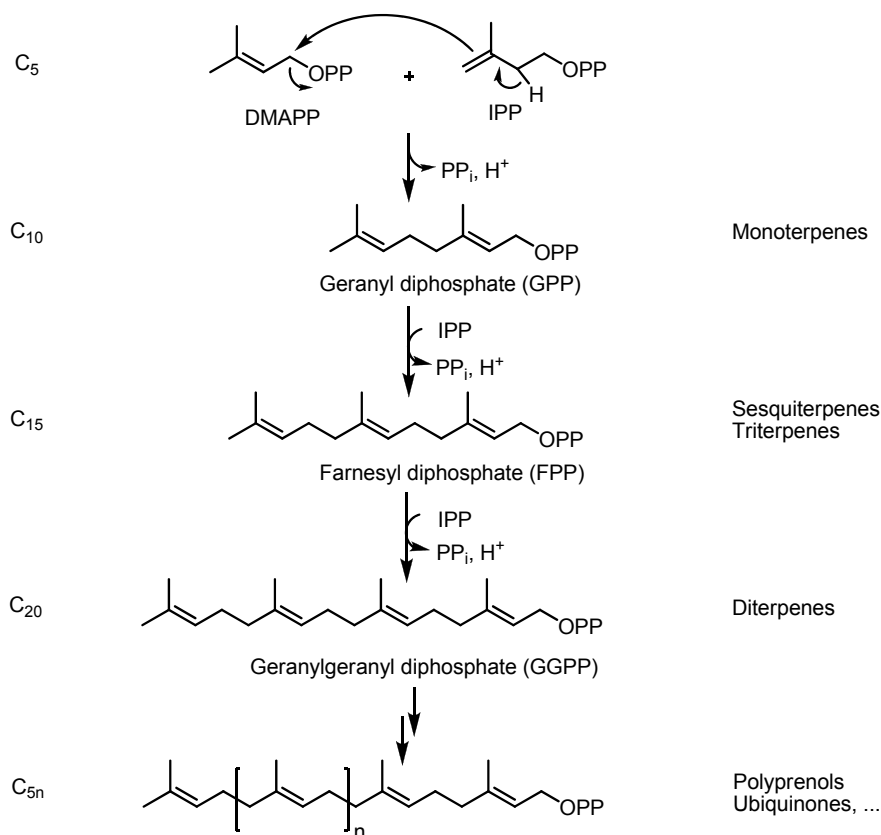
The Rohmer – DXP pathway: The DXP pathway (Scheme 21), which was discovered in 1993, is still not fully understood. The synthesis of IPP is initiated by a thiamin catalyzed “Umpolung”-reaction of pyruvate, which then attacks glyceraldehydes 3-phosphate with concomitant loss of CO₂. The so produced DXP is converted to 2C-methyl-D-erythritol 4-phosphate by an acyloin type rearrangement, to give an intermediate aldehyde, which is reduced by one equivalent of NADPH/H⁺. Sequential phosphorylation by CTP (cytidine triphosphate) and ATP results in the formation of 2C-methyl-D-erythritol 2,4-cyclodiphosphate. Little is known about the further conversion to IPP.



Scheme 21. The Rhomer – deoxyxylulose (DXP) pathway.

Scheme 22 shows the biosynthesis of higher terpenoids. After isomerization of IPP to DMAPP, Mg (II) mediated head-to-tail condensation of IPP with DMAPP gives geranyl diphosphate (GPP, C₁₀). Nucleophilic Alder-ene-type attack³² of IPP at the primary position of GPP creates the C₁₅ terpenoid farnesyl diphosphate (FPP).

³² K. Alder, F. Pascher, A. Schmitz, *Chem. Ber.* **1943**, 76, 27–53.



Scheme 22. Biosynthesis of higher terpenoids.

Tail-to-tail dimerization of FPP gives triterpenes and reductive coupling of two FPP units affords squalene, the biosynthetic precursor of steroids. Geranylgeranyl diphosphate (GGPP) and higher terpenes are derived from FPP by sequential addition of IPP molecules.

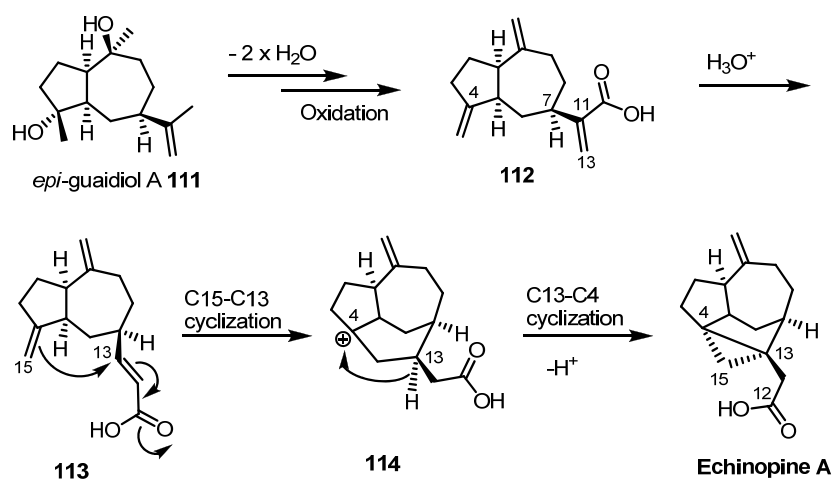
The great diversity of natural occurring terpenoids, in turn, arises from enzymatic catalyzed (terpene cyclases) cascade reactions such as cyclization of carbocation intermediates, Wagner–Meerwein rearrangements, hydride shifts, attack of nucleophiles (H₂O) and deprotonation.

5.3.2. Proposed Biosynthesis of Echinopine A and B

Kiyota and Shi²⁸ also speculated about the possible biosynthetic origin of echinopine A (**109**) and B (**110**). Although their biosynthesis is plausible and reasonable, there is no evidence for this kind of pathway (Scheme 23). However, proposed **112** could be derived from recently isolated *epi*-guaidiol A **111**.³³ Hydration of the C11–C13 double bond, followed by acid induced migration of the C7–C11 bond gives α,β -unsaturated acid **113**. Attack of the C14–C15 *exo*-methylene group at the C13 double bond of **113**

³³ Y. Xu, H.-W. Zhang, X.-C. Wan, Z.-M. Zou, *Magn. Reson. Chem.* **2009**, *47*, 527–531.

and additional cyclization of **114** with loss of a proton leads to echinopine A. Subsequent esterification affords echinopine B.



Scheme 23. Proposed biosynthesis of echinopine A.

6. Total Syntheses of Selected Terpenoids

The following total syntheses of sesquiterpenoids were chosen particularly with regard to echinopine A and B structurally related natural products. The ambition was to find structurally closely related targets such as 5,5-, 5,7- and 3,5,7- membered compounds. Natural products possessing a *cis*-5,7-membered scaffold are not as omnipresent as the thermodynamically favored *cis*-fused 5,5-analogs. The low occurrence of 3,5,7-membered frameworks in nature is reflected in that examples of terpenoids with an inherent cyclopropane-cyclopentane bicycle are limited (e.g. shizukaol, chloranthalactone). The presented publications should provide an insight into (modern) strategies for the synthesis of the relevant structural motifs.

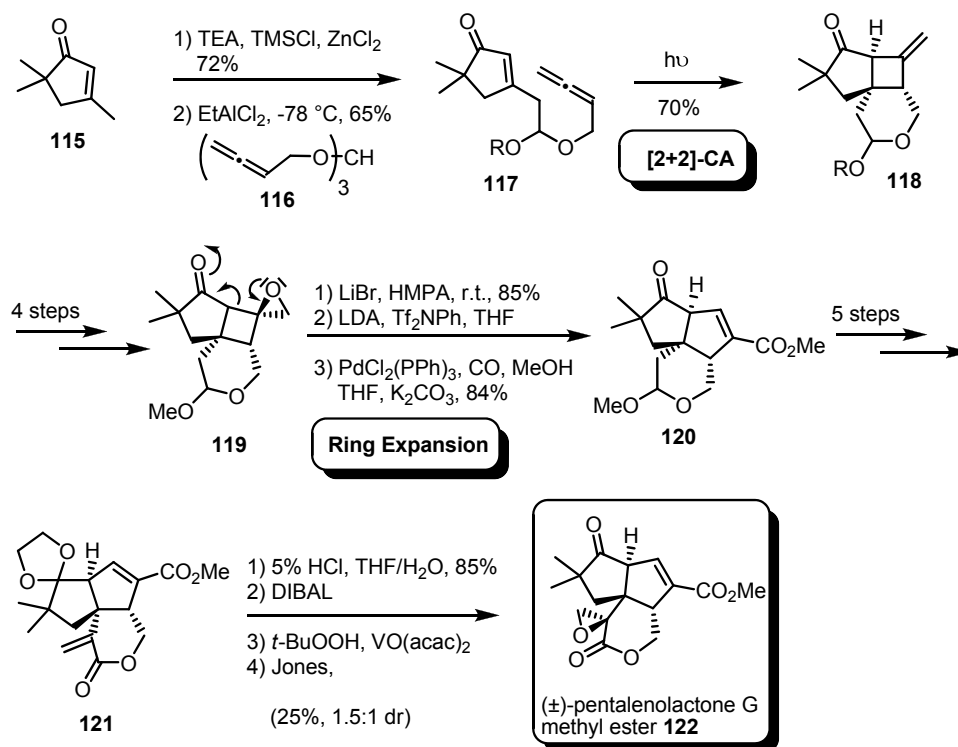
6.1. 5,5-Membered Ring Systems

6.1.1. Pirrung's Total Synthesis of (±)-Pentalenolactone G Methyl Ester

Strategy: Intramolecular [2+2]-cycloaddition – ring expansion.

In 1988, a total synthesis of (±)-pentalenolactone G methyl ester **112** (Scheme 24), bearing a 5,5-fused framework was reported by Pirrung.³⁴ Conversion of **115** to the corresponding dienylsilylether and treatment with tris((allenyl)methyl)orthoformate **116** gave cyclization precursor **117**. Irradiation induced the desired [2+2]-cycloaddition and provided *cis*-fused tricycle **118** in good yield. After four steps, rearrangement of epoxide **119**, presumably initiated by a LiBr induced retro aldol-type reaction, afforded the desired 5,5-membered ring skeleton. Palladium mediated carbonylative coupling of the vinyltriflate afforded α,β -unsaturated methylester **120**, which was converted to **121** in five steps. The remaining steps of the total synthesis were accomplished relying on Danishefsky's procedure and provided **122** in 0.5% overall yield.

³⁴ M. C. Pirrung, S. A. Thomson, *J. Org. Chem.* **1988**, *53*, 227–230.



Scheme 24. Pirrung's [2+2]-cycloaddition – ring expansion strategy.

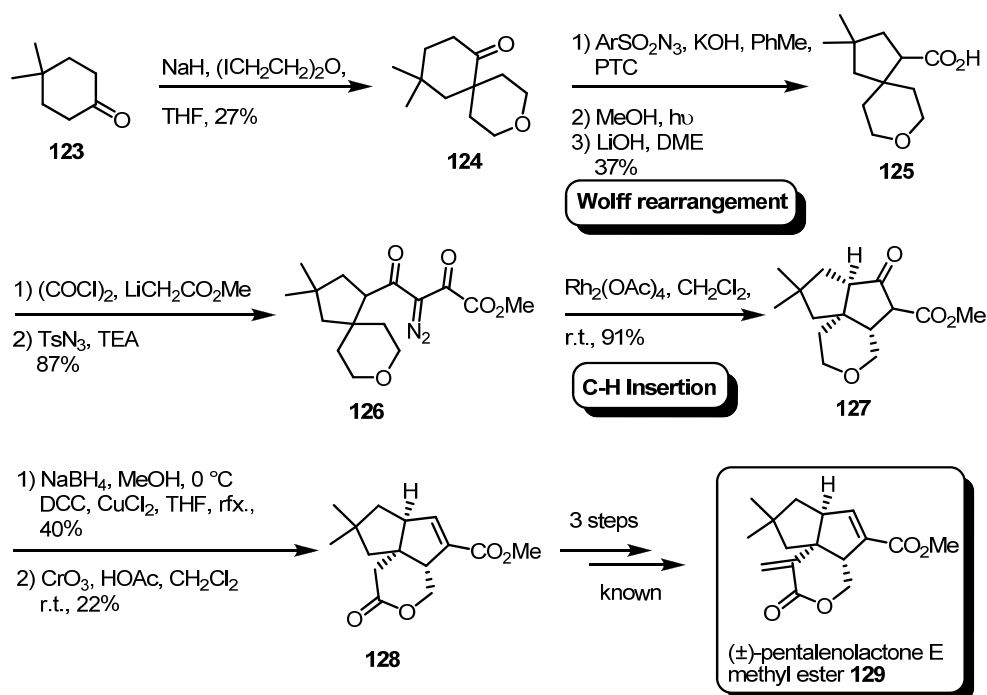
6.1.2. Taber's Formal Synthesis of (±)-Pentalenolactone E

Strategy: Rh(II) catalyzed C–H insertion.

Taber's formal synthesis of (±)-pentalenolactone E, published in 1985³⁵ and 1987³⁶, was accomplished via a Rh(II) catalyzed C–H insertion as the key step (Scheme 25). Conversion of the C₂-symmetric ketone **123** to *spiro*-compound **124** followed by a Wolff ring contraction creates the left-hand five-membered ring. Regitz' diazo transfer reaction with **125** provided intermediate **126**, which on treatment with Rh₂(OAc)₄ gave the complete skeleton of the target compound.

³⁵ D. F. Taber, J. L. Schuchardt, *J. Am. Chem. Soc.* **1985**, *107*, 5289–5290.

³⁶ D. F. Taber, J. L. Schuchardt, *Tetrahedron* **1987**, *43*, 5677.



Scheme 25. Taber's Rhodium(II) catalyzed C–H insertion strategy.

Deoxygenation and installation of the six-membered lactone afforded **128**. The completion of the synthesis could be realized by three literature known steps.

6.2. 5,7-Membered Ring Systems

The most difficult task arises from the preparation of *cis*-fused bicycles, which feature a ketone or double bond functionality next to the ring junction. The ketone intermediates are prone to *cis*–*trans* isomerization and acidic induced migration of 1,1-disubstituted double bonds³⁷ complicates the installation of *exo*-methylene groups. A practical method for the late stage installation of the latter was recently published by Vanderwal.³⁸

6.2.1. Rigby's Total Synthesis of (±)-Dehydrocostus Lactone & (±)-Estafiatin

Strategy: Intramolecular hetero Diels–Alder reaction.

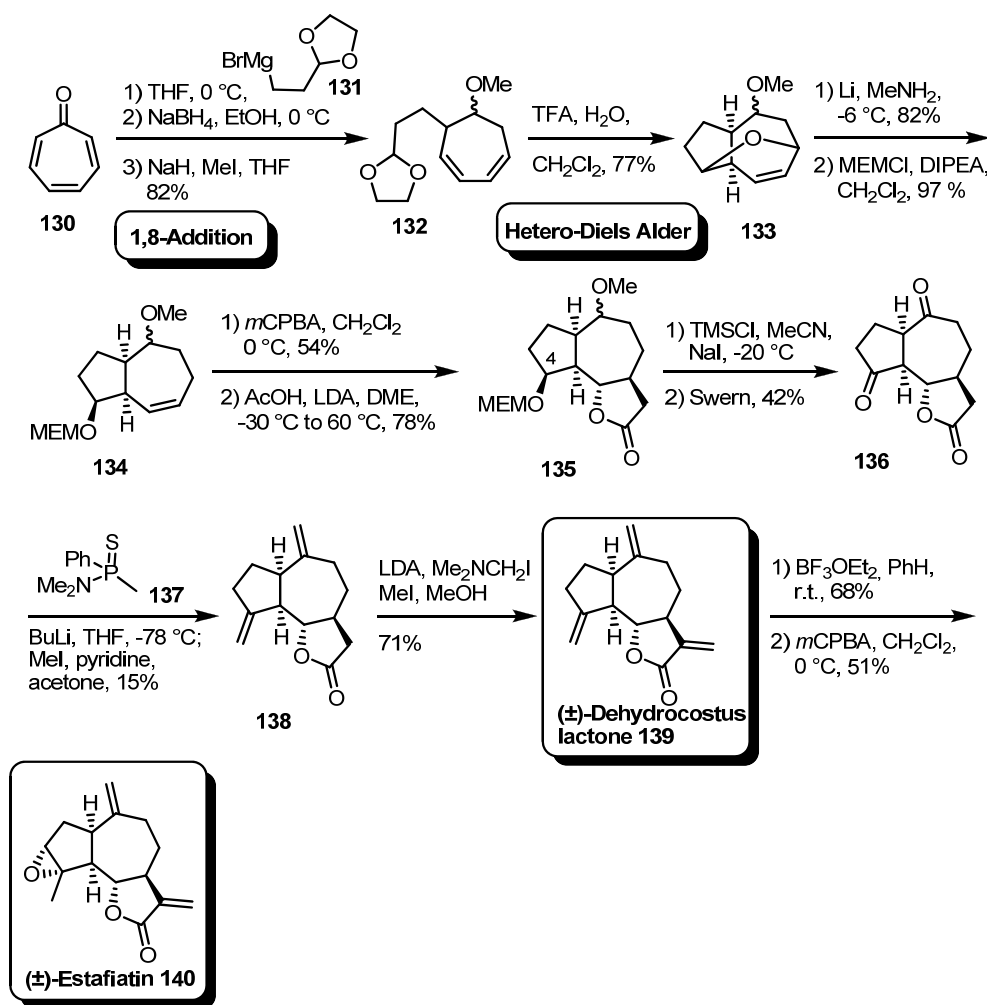
The first total syntheses of the guainolide sesquiterpenes (±)-dehydrocostus lactone **139** and (±)-estafiatin **140** were reported by Rigby (Scheme 26).³⁹ The key step was an intramolecular Diels–Alder

³⁷ G. L. Lange, C. Gottardo, *J. Org. Chem.* **1995**, *60*, 2183–2187.

³⁸ M. S. Dowling, C. D. Vanderwal, *J. Am. Chem. Soc.* **2009**, ASAP.

³⁹ J. H. Rigby, J. Z. Wilson, *J. Am. Chem. Soc.* **1984**, *106*, 8217–8224.

reaction with tropone **130**. Nucleophilic 1,8-addition of Grignard **131** to **130** followed by immediate reduction, in order to prevent [1,5]-H shifts mediated double bond isomerization, and protection gave diene **132**. One pot cleavage of the acetal and intramolecular hetero Diels–Alder reaction afforded tricyclic allyl ether **133**. Birch reduction of the allylic C–O bond and protection gave *cis*-hydroazulene **134**. Introduction of the appended lactone ring was performed via stereoselective epoxidation and subsequent epoxide opening with dilithioacetate to give **135**.



Scheme 26. Preparation of a *cis*-annulated 5,7-bicycle via a hetero Diels–Alder reaction.

After double deprotection and Swern oxidation, introduction of the exocyclic methylene groups was achieved with olefination reagent **137**, albeit in low yield. Attempts to introduce the double bonds by different methods only resulted in *cis*–*trans* epimerization, although it was known, that hydroazulenes with an appended butyrolactone often favor the *cis*-annulation. Since all efforts to cleave the methoxy

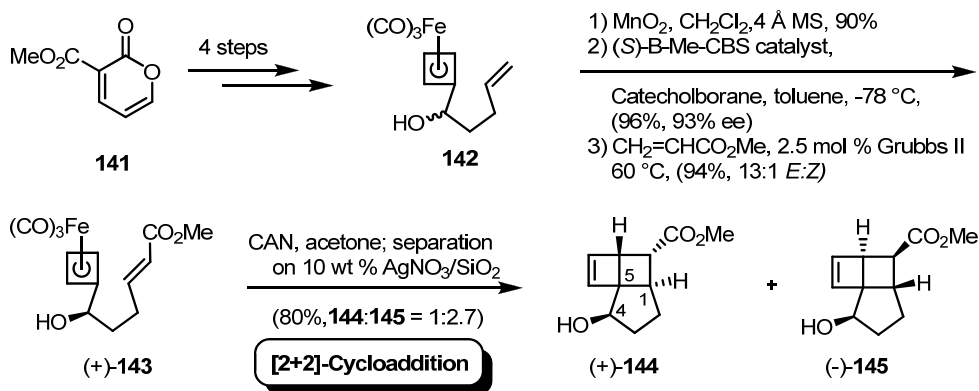
group after *mono*-olefination of **135** (C4 position) resulted in double bond migration, the simultaneous installation of the double bonds was essential.

Eschenmoser methylenation of **138** gave (\pm)-dehydrocostus lactone **139** in 1% overall yield. Conversion to (\pm)-estafiatin **140** was performed via double bond isomerization and epoxidation.

6.2.2. Snapper's Total Synthesis of *ent*-(+)-Pleocarpenene and *ent*-(-)-Pleocarpenone

Strategy: [2+2]-Cycloaddition – cyclopropanation – rearrangement.

In 2007, Snapper from Boston College reported the intramolecular cycloaddition–cyclopropanation–thermal rearrangement strategy for the total synthesis of the guaiane type sesquiterpenes pleocarpenene **148** and pleocarpenone **149**.⁴⁰ The synthesis started with α -pyrone **141**, which was converted to iron protected cyclobutadiene **142** in four steps (Scheme 27). The enantioselective route was realized by allylic MnO_2 oxidation of **142**, followed by (*S*)-B-Me-CBS reduction and final cross-metathesis to give the C_1 -elongated ester (+)-**143**. Treatment of (+)-**143** with CAN initiated the liberation of the cyclobutadiene moiety and afforded, after formal [2+2]-cycloaddition, enantioenriched diastereomers (+)-**144** and (-)-**145** as a 1:2.7 ratio in good yield.

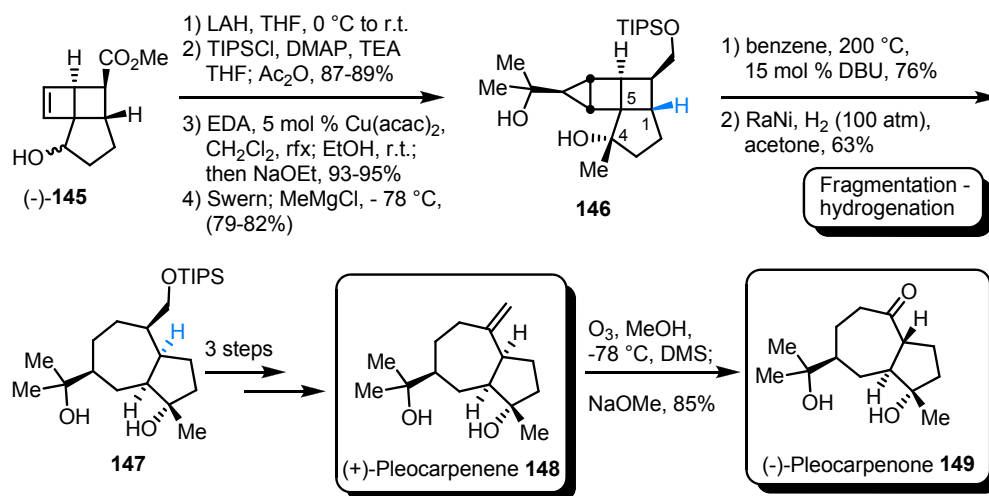


Scheme 27. Snapper's [2+2]-cycloaddition – cyclopropanation – fragmentation strategy.

Next they focused on the preparation of **146**, whereas, in principle, both diastereomers (concerning the hydroxyl group at C4) of one enantiomeric series could be used for this transformation (Scheme 28). The sequence started with reduction of (-)-**145** and sequential protection of the primary and secondary alcohol. Cu(II) mediated cyclopropanation with ethyl diazoacetate (EDA) from the less congested convex

⁴⁰ M. J. Williams, H. L. Deak, M. L. Snapper, *J. Am. Chem. Soc.* **2007**, *129*, 486–487.

face with *in situ* cleavage of the acetate, followed by oxidation and nucleophilic attack of excess MeMgCl gave diol **146**. Heating the strained tetracycle **146** to 200 °C, in the presence of catalytic amounts of DBU, caused the desired rearrangement with simultaneous inversion of stereochemistry at C1. Their mechanistic model, already published in 2001,⁴¹ suggests a stepwise fragmentation–recombination sequence. Hydrogenation provided the desired *cis*–annulation in **147**. A high yielding three step sequence was used for the conversion of **147** to pleocarpenene **148**. Ozonolysis of the *exo*–methylene group and *cis*–*trans* isomerization of the ring junction finally afforded pleocarpenone **149** in 18 steps from α –pyrone **141**.



Scheme 28. Completion of the synthesis.

The major drawback is the use of highly complex intermediates such as **146**, which are degraded to a much simpler 5,7–membered ring system in the course of the synthesis.

6.2.3. Wender's Total Synthesis of (+)-Dictamol

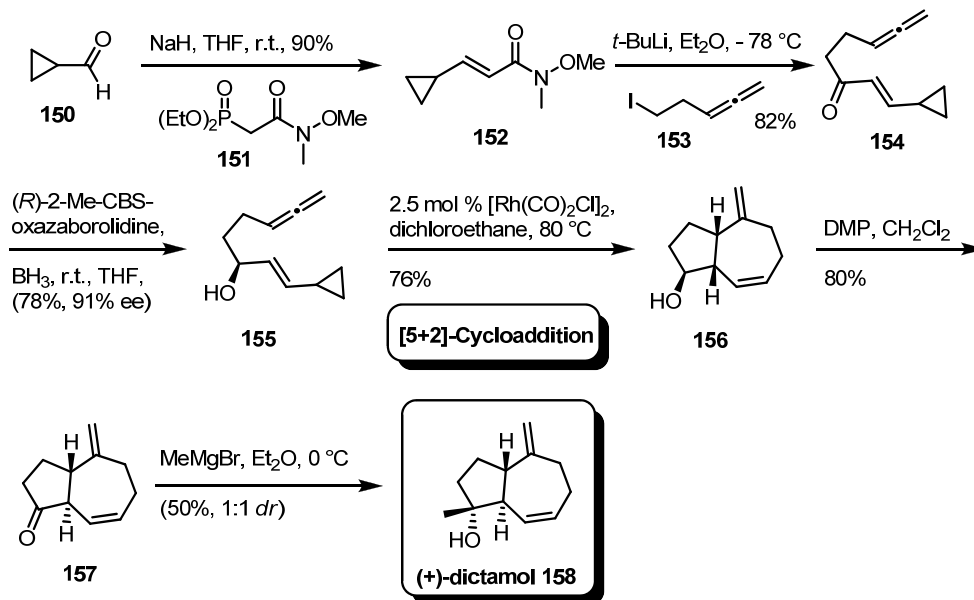
Strategy: [5+2]–Cycloaddition.

The Rh(I) catalyzed [5+2]–cycloaddition was used as the key step in Wender's nine steps total synthesis of (+)-dictamol **158**.⁴² The synthesis of the allene–cyclopropane cyclization precursor **155** started with a Horner–Wadsworth–Emmons reaction of **151** with commercially available **150** (Scheme 29). Addition of lithiated 1–iodo–3,4–butadiene **153** to **152** and subsequent CBS reduction of **154** provided (*R*)–**155**. Treatment with catalytic amounts of Rh(I) such as Rh(PPh₃)₃Cl or [Rh(CO)₂Cl]₂ delivered the desired

⁴¹ H. L. Deak, S. S. Stokes, M. L. Snapper, *J. Am. Chem. Soc.* **2001**, *123*, 5152–5153.

⁴² P. A. Wender, M. Fuji, C. O. Husfeld, J. A. Love, *Org. Lett.* **1999**, *1*, 137–139.

[5+2]–cycloadduct **156** in good yields and high diastereoselectivity. The remaining steps towards **158** were achieved via Dess–Martin oxidation, silica gel chromatography with concomitant *cis*–*trans* epimerization and methylation.



Scheme 29. Wender's total synthesis of (+)-Dictamol.

6.3. 3,5,7-Membered Ring Systems

The major challenge is again the preparation of the 5,7-membered framework, as the cyclopropane moiety is most often obtained either from chiral pool starting material or introduced at last.

6.3.1. White's Total Synthesis of (±)-Africanol

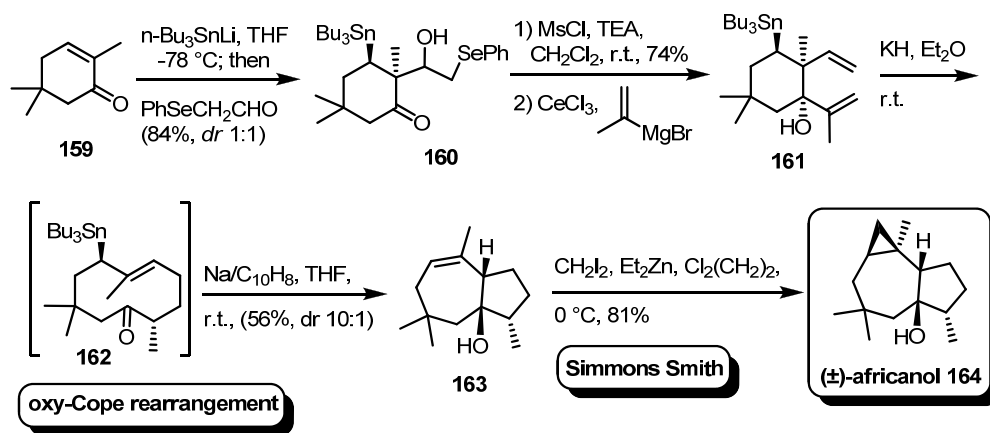
Strategy: Anionic oxy-Cope rearrangement – cyclization.

Several total syntheses of the tricyclic sesquiterpenoid africanol **164** have been reported.⁴³ White's six steps synthesis⁴⁴ starts with a conjugate addition–enolate trapping sequence to α,β –unsaturated ketone **159** (Scheme 30). After separation of the 1:1 diastereomeric mixture of **160**, the vinyl moiety was installed via elimination and the isopropenyl group via equatorial Grignard addition.

⁴³ a) H. Shirahama, K. Hayana, Y. Kanemoto, S. Misumi, T. Ohtsuka, N. Hashiba, A. Furusaki, S. Murata, R. Noyori, T. Matsumoto, *Tetrahedron Lett.* **1980**, 21, 4835–4838; b) L. A. Paquette, W. H. Ham, *Tetrahedron Lett.* **1986**, 27, 2341–2343; c) L. A. Paquette, W. H. Ham, *J. Am. Chem. Soc.* **1987**, 109, 3025–3036.

⁴⁴ W. Fan, J. B. White, *J. Org. Chem.* **1993**, 58, 3557–3562.

Anionic oxy-Cope rearrangement of **161** gave the unstable 5-cyclodecenone **162**, which, upon *in situ* treatment with sodium naphthalenide, cyclized to **163** via an intermediate ketyl radical. Careful experimentation showed that the diastereoselectivity of the cyclization step depends on the reaction conditions (TBAF, SnCl_4 , Sml_2) as well as on the stereochemical relationships of the substituents in the ten-membered intermediate **162** (*cis* vs. *trans*). A mechanistic model for this transformation is still missing. Simmons-Smith cyclopropanation from the less hindered convex face afforded (\pm)-africanol **164** in an overall yield of 11%.



Scheme 30. White's oxy-Cope rearrangement approach to (\pm)-africanol.

6.3.2. Wijnberg's & Groot's Total Synthesis of (\pm)-Alloaromadendrane-4 α , 10 α -diol

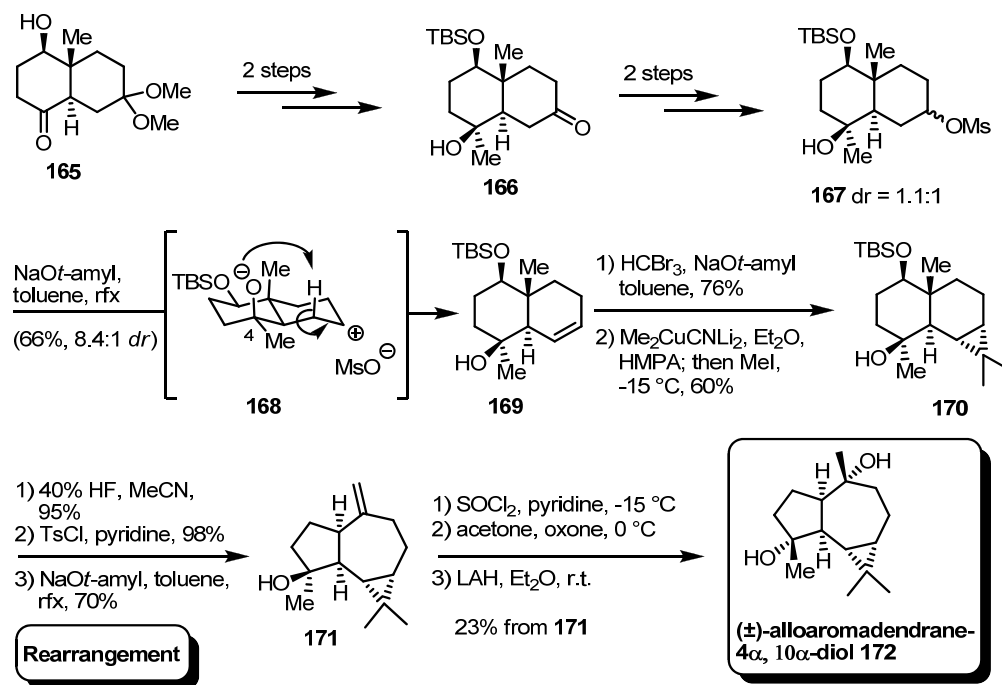
Strategy: Fragmentative rearrangement.

A total synthesis of (\pm)-alloaromadendrane-4 α , 10 α -diol **172**, bearing a *cis*-fused 5,7-membered aromadendrane skeleton, was reported from Wijnberg and Groot.⁴⁵ Starting from known **165** they constructed fragment **167** in four steps (Scheme 31). Treatment of mesylate **167** (1.1:1 diastereomeric mixture) with base in refluxing toluene predominantly afforded the desired elimination product **169**, probably via internal 1,3-diaxial deprotonation of dipolar intermediate **168**. The suggested intermediate was supported by additional experiments. Rerun of the sequence gave almost the same yield and regioselectivity for the axial α -mesylate as well as the equatorial β -mesylate within two minutes. Compounds lacking the oxygen substituent at C4 did not show any elimination or fragmentation.

The installation of the *gem*-dimethylcyclopropane moiety was realized by dibromocarbene addition to **169** and subsequent methylation. Deprotection of **170** followed by selective tosylation paved the way

⁴⁵ L. H. D. Jenniskens, J. B. P. A. Wijnberg, A. deGroot, *J. Org. Chem.* **1991**, *56*, 6585–6591.

for the key reaction. Heating a solution of the tosylate and excess sodium *t*-amylate afforded almost exclusively the rearranged 5,7-membered framework **171** in good yield. Again, internal deprotonation by the preformed axial alkoxide substituent probably favored the formation of the *exo*-double bond. The total synthesis was finished via elimination, double epoxidation with *in situ* generated DMDO and a final reduction step.



Scheme 31. Wijnberg's & Groot's total synthesis of (±)-alloaromadendrane-4 α , 10 α -diol.

7. Results

7.1. Total Synthesis of (+)-Echinopine A and B: Determination of Absolute Stereochemistry

T. Magauer, J. Mulzer, K. Tiefenbacher, *Org. Lett.* **2009**, ASAP.

Total Syntheses of (+)-Echinopine A and B: Determination of Absolute Stereochemistry

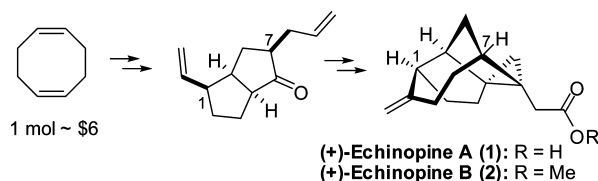
Thomas Magauer,* Johann Mulzer, and Konrad Tiefenbacher*

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Received September 30, 2009

ABSTRACT



The first total syntheses of the novel 3,5,5,7-sesquiterpenoids (+)-Echinopine A (1) and B (2) were achieved. Thereby the proposed structures were confirmed, and the absolute stereochemistry was determined. The key features are (1) the stereoselective installation of the vinyl-moiety on the concave side of the bicyclo[3.3.0]octane core via Myers' [3,3]-sigmatropic rearrangement, (2) the finding that the substituent on the C7 position next to the ketone can be epimerized to the desired concave face under basic conditions, (3) the closure of the highly strained seven-membered ring via RCM, and (4) the unusual C2-homologation of a vinyltriflate with a ketene silyl acetal.

Echinopine A (1) and B (2) were isolated in 2007 by Shi and Kiyota during their screening for biologically active compounds from *Echinops spinosus*.¹ The structures of these novel, unprecedented sesquiterpenoids were elucidated by ¹H, ¹³C, 2D-NMR and HRMS. Their molecular framework consists of a unique 3,5,5,7-membered ring skeleton (Figure 1), which is believed to arise from a guaiane-type precursor

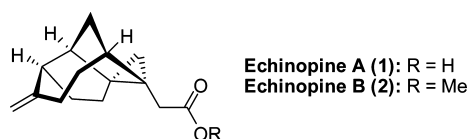


Figure 1. Structures of Echinopine A and B.

via skeletal rearrangement. The complex tetracyclic architecture features five contiguous stereocenters, two of which are quaternary.

(1) Dong, M.; Cong, B.; Yu, S.-H.; Sauriol, F.; Huo, C.-H.; Shi, Q.-W.; Gu, Y.-C.; Zamir, L. O.; Kiyota, H. *Org. Lett.* **2008**, *10*, 701–704.

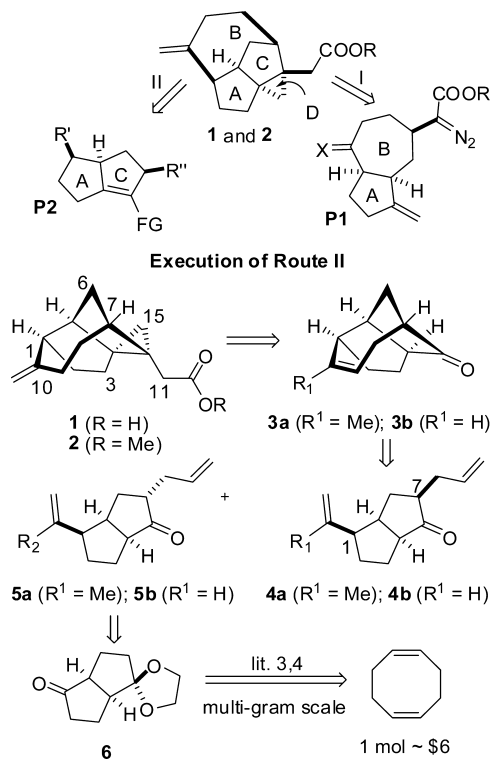
Because of the scarcity of the natural product source, the biological activity has not satisfactorily been uncovered so far. Additionally no X-ray crystallographic analysis or chemical derivatization could be performed.¹ The absolute configuration is unknown, and the assignment of the relative stereochemistry is solely based on ¹H NMR spectroscopy.

The unprecedented structure and the unknown biological profile motivated us to embark on the total synthesis of **1** and **2**.

For a retrosynthetic analysis (Scheme 1) we considered two alternatives (I and II), leading to bicyclic precursors **P1** and **P2**, respectively. **P1** is a *cis*-fused AB-guaianolide system in which both rings C and D might be generated in a one-step cyclopropanation. However, this approach would be jeopardized by a potential *cis-trans*-isomerization, if for instance X = O. Alternatively, for X = CH₂ there would be a regio problem in the cyclopropanation as well as a high risk of double bond migration. Therefore we preferred route II, which goes back to a diquinane AC precursor **P2** with two *cis*-appendages R' and R'', suitable for closing the seven-membered ring C via RCM.

Scheme 1. Retrosynthetic Analysis of **1** and **2**

Retrosynthetic Alternatives I and II



Thus, the *exo*-methylene group at C10 was retrosynthetically shifted to the *endo*-position. The cyclopropane and the acetic acid side chain were to be grafted on ketone **3**, which in turn should be elaborated from bis-alkene **4**. The synthesis of **4** was not trivial as both the vinyl/isopropenyl group at C1 (numbering identical to the final compounds) and the allyl group at C7 are located on the sterically encumbered concave face of the bicyclo[3.3.0]octane core. We therefore decided to explore a DRCM (dynamic ring closing metathesis), which has been described only once so far.² Equilibration of the kinetically allylated ketone **5** to RCM precursor **4** and in situ RCM should therefore deliver tricycle **3**. Ketone **5** in turn should be accessible from known racemic³ or optically active⁴ bicycle **6**, which can be easily prepared in multigram quantities from inexpensive 1,5-cyclooctadiene.

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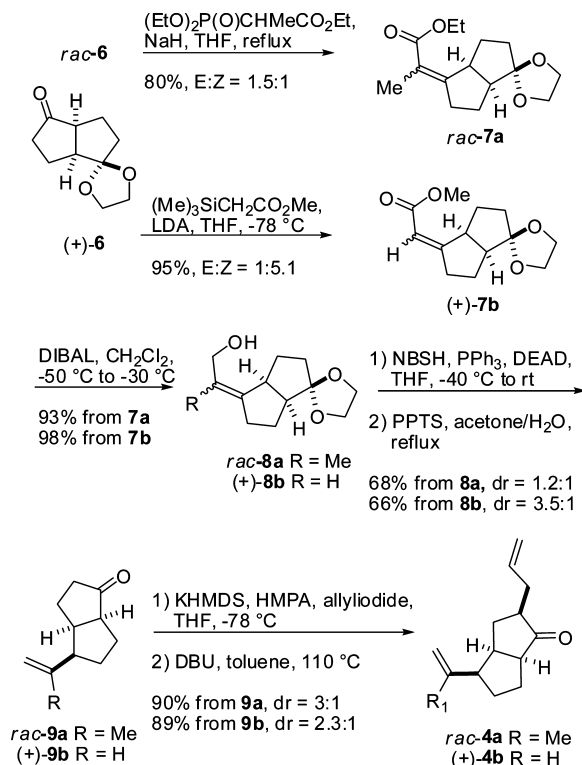
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The synthesis of the RCM precursors (Scheme 2) *rac*-**4a** and (+)-**4b** started by converting ketone **6** to unsaturated

Scheme 2. Preparation of RCM Precursors **4a/4b**



esters **7a** and **7b** via Horner–Wadsworth–Emmons⁵ and Peterson⁶ olefination. Initial efforts to force the C1 side chain on the more hindered concave face by hydrogenating either the unsaturated esters or allylic alcohols resulted in 1:1 diastereomeric mixtures. Careful experimentation, however, revealed that the allylic alcohols **8a** and **8b** could be converted to the desired vinyl appendages by Myers' [3,3]-sigmatropic rearrangement protocol.⁷ The stereochemical outcome can be rationalized by steric interactions with the adjacent cyclopentane ring (Figure 2), which is larger for the (*Z*)- than for the (*E*)-isomer. In fact, after separating the **8a** *Z/E* mixture, (*Z*)-**8a** afforded after deketalization a 10:1 ratio in favor of diastereomer **9a**, whereas the ratio dropped to 1.2:1 for (*E*)-**8a**. For compound **8b** (*Z/E* 5.1:1), the effect was less pronounced, giving **9b** as an easily separable 3.4:1 diastereomeric mixture.

Having secured gram quantities of both **9a** and **9b**, we finished the preparation of the RCM precursors. Allylation with KHMDS (only 1.05 equiv, otherwise bisallylation was observed) and excess allyliodide gave **5a** and **5b** in excellent yield and as single diastereomers.

The envisaged DRCM strategy was investigated under a variety of RCM catalysts, bases, Lewis acids, and solvents (Grubbs II, Grubbs Hoveyda II, "Grubbs III",⁸ DBU, Schwesinger's *t*-Bu-P₄-base, ZnCl₂, AlCl₃,⁹ toluene, CH₂Cl₂). After extensive experimentation, we realized that DRCM was

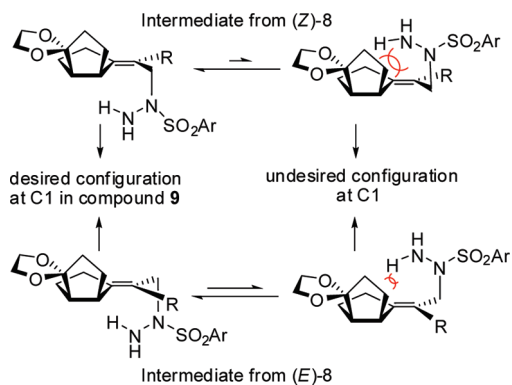


Figure 2. Models for the stereoselective [3,3]-sigmatropic rearrangement.

ineffective. However the DRCM experiments revealed that DBU (toluene, 110 °C) efficiently epimerized both **5a** and **5b** to mixtures of **4a/5a** (dr = 3:1) and **4b/5b** (dr = 2.3:1) with the desired *cis*-diastereomers in excess. The undesired diastereomers could be easily separated by HPLC and recycled.

Next RCM was applied to **4a** and **4b**. To our surprise all attempts to close the seven-membered ring from **4a** failed, under a variety of reaction conditions (Grubbs II, Grubbs Hoveyda II, “Grubbs III”, CH₂Cl₂, toluene, 2,6-dichlorobenzoquinone¹⁰). Only homodimers across the allylic units were observed. Gratifyingly the less hindered **4b** was smoothly converted to the desired core framework **3b** when exposed to Grubbs II catalyst (5 mol %) in refluxing CH₂Cl₂ (Scheme 3). As **5b** gave only polymers under these conditions, we streamlined our synthesis by submitting the **4b/5b** mixture to the RCM reaction, from which **3b** was readily isolated as the only monomeric product. In principle, the material can be carried through from **7b** to **3b** without separation as **4b** is the only diastereomer capable of RCM.

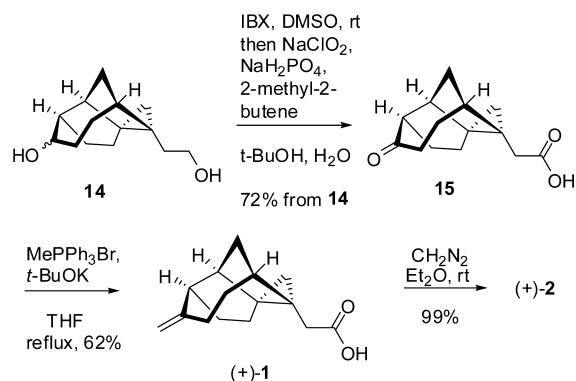
With **3b** in hand we started the installation of the remaining functionalities. We envisioned the introduction of the C2 side chain stepwise (path A) or directly (path B). In either case, the procedure commenced with the formation of the vinyltriflate with KHMDS/PhNTf₂ and a Pd(0)-mediated coupling reaction. As one-step formylation¹¹ failed, path A was focused on the well-established C1 elongation protocol with CO/MeOH in DMF,¹² which afforded the α,β -unsaturated methyl ester in excellent yield. Stereoselective Corey–Chaykovsky cyclopropanation¹³ furnished ester **10**, and primary alcohol **12** was obtained after C1 elongation. Functionalization of the disubstituted double bond of **12**

was accomplished by epoxidation with DMDO (which gave higher yields than *m*CPBA) and regioselective epoxide opening with LAH at elevated temperature to give **14** as a 16:1 diastereoisomeric mixture. Efforts to introduce appropriate functionality at C-10 in **12** via ene reactions with singlet oxygen¹⁴ or formaldehyde¹⁵ resulted only in the recovery of starting material.

In a more direct access to **14** (path B) we extended related methodology of Musco and Santi¹⁶ to the vinyltriflate derived from ketone **3a**. Indeed, reaction with (1-methoxyvinyl-oxo)trimethylsilane and Pd(PPh₃)₄ furnished methyl ester **11** in 54% yield (not optimized). Regioselective cyclopropanation with modified Furukawa–Simmons–Smith’s reagent¹⁷ afforded the tetracyclic intermediate **13** in excellent yield. Epoxidation and LAH reduction gave **14** as before.

Oxidation of **14** (Scheme 4) first with IBX and then with NaClO₂¹⁸ furnished ketoacid **15**, whose Wittig methylenation

Scheme 4. Completion of the Syntheses



in refluxing THF led to (+)-**1** ([α]_D²² = +26 (*c* 0.70, CHCl₃), lit.¹ [α]_D²² = +23 (*c* 0.11, CHCl₃)). The acid was converted to (+)-**2** ([α]_D²² = +22 (*c* 0.60, CHCl₃), lit.¹ [α]_D²² = +21 (*c* 0.14, CHCl₃)) with diazomethane. The analytical data for **1** and **2** (HRMS, ¹H and ¹³C NMR, [α]_D²²) matched those reported by Shi and Kiyota in all respects. The relative configuration of **1** was also confirmed by single crystal analysis, whereas the absolute configuration of **1** and **2** follows unambiguously from the synthesis.

In summary, the first total synthesis of Echinopine A (**1**) and B (**2**) was achieved in optically active form in 15 steps from known ketone **6** in 7% overall yield. The sequence is easy to perform and scaleable and starts from

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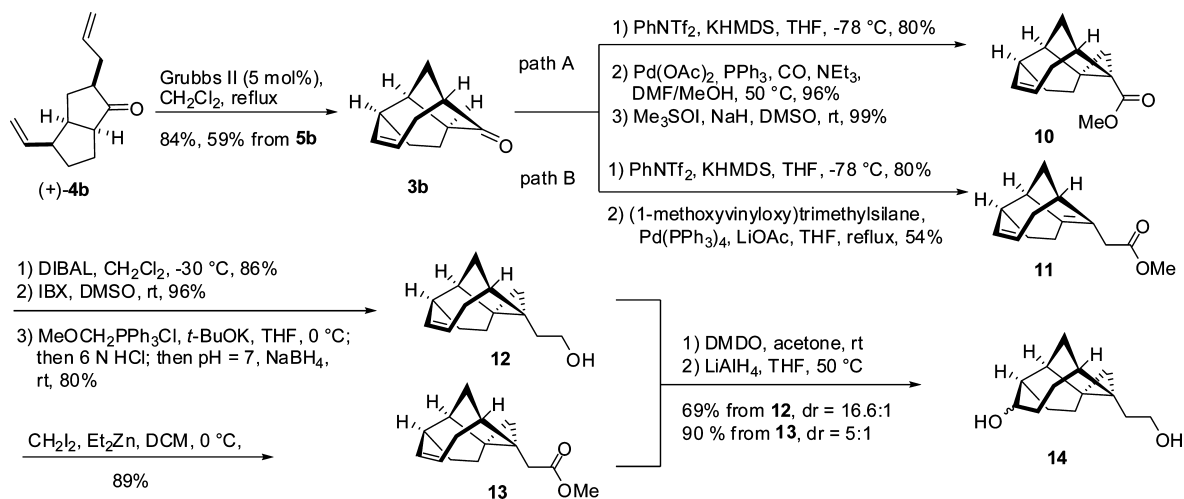
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Scheme 3. Construction of the Polycyclic [3,5,5,7] Skeleton



inexpensive material. Notable chemical features of the synthesis are (1) the stereoselective installation of the vinyl moiety on the concave side of the bicyclo[3.3.0]octane core via Myers' [3,3]-sigmatropic rearrangement, (2) the finding that the C7-substituent next to the ketone (see **4** and **5** in Scheme 1) can be epimerized to the desired concave face under basic conditions, (3) the closure of the highly strained seven-membered ring via RCM, and (4) the unusual C2-homologation of a vinyl triflate with a ketene silyl acetal.

Acknowledgment. The authors thank A. Roller and Prof. V. Arion (University of Vienna) for the crystal structure analysis of **1** and Prof. A. Fürstner (MPI Mülheim) for helpful discussions.

Supporting Information Available: Experimental procedures and analytical data for all new compounds and crystal data of **1**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

OL902263K

Total Syntheses of (+)-Echinopine A and B: Determination of Absolute Stereochemistry

SUPPORTING INFORMATION

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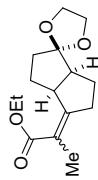
E-mail: thomas.magauer@univie.ac.at, konrad.tiefenbacher@univie.ac.at

Experimental part

General

All reactions were carried out in oven-dried glassware under an argon atmosphere using standard syringe/septa techniques, unless otherwise stated. Anhydrous toluene was distilled from sodium/benzophenone under argon. Anhydrous CH_2Cl_2 , DMF (*N,N*-dimethylformamide) and HMPA (hexamethylphosphoramide) were distilled from CaH_2 under argon or reduced pressure, respectively. Anhydrous THF (tetrahydrofuran) was purchased from Acros (99.85%, water < 50 ppm). All other solvents were HPLC grade. Solvent degassing was achieved by repeated (at least 4 cycles) freeze-pump-thaw (FPT) cycles. Reactions were magnetically stirred and monitored by thin layer chromatography (TLC) with E. Merck silica gel 60-F254 plates. Flash column chromatography was performed with Merck silica gel (0.04-0.063mm, 240-400 mesh) under pressure. Preparative HPLC was performed on a Dynamix Model SD-1 equipped with a Model UV-1 absorbance detector using a Supershere (60 Å pore size, 4 µm particle size, Ø 25 mm x 250 mm) at ambient temperature. Spots were visualized under UV light (254nm) and/or were stained with ceric ammonium molybdate (CAM), *p*-anisaldehyde or potassium permanganate stain. Yields refer to chromatographically and spectroscopically pure compounds, unless stated otherwise. NMR spectra were recorded on either Bruker Avance DRX-400 or DRX-600 at 400.13 MHz (100.61 MHz) or 600.13 MHz (150.90 MHz) spectrometer. Unless stated otherwise, all NMR spectra were measured in CDCl_3 solutions and referenced to the residual CHCl_3 signal (^1H , $\delta = 7.26$ ppm; ^{13}C , $\delta = 77.00$ ppm). All ^1H and ^{13}C shifts are given in ppm (s = singlet; d = doublet; t = triplet; q = quadruplet; qu = quintet, m = multiplet; b = broad signal). Assignments of proton resonances were confirmed, when possible, by correlated spectroscopy. IR spectra were recorded as thin films on a silicon plate on a Perkin-Elmer 1600 FT-IR spectrometer. Optical rotations were measured on a P 341 Perkin-Elmer polarimeter at 589 nm with a 100 mm path length cell at 20°C, unless otherwise stated (reported as follows: concentration (*c* in g/100mL), solvent). Mass spectra were measured on a Micro mass, trio 200 Fisions Instruments. High resolution mass spectra (HRMS) were performed with a Finnigan MAT 8230 with a resolution of 10000.

Procedures



Ester *rac*-7a. NaH (60%, 524 mg, 13.16 mmol, 1.7 equiv.) was washed with pentane (3 x 10 mL), suspended in THF (4 mL) and cooled to 0 °C. Triethyl-2-phosphonoacetate (2.86 mL, 13.16 mmol, 1.7 equiv.) was added dropwise *via* syringe and stirring was continued for 40 min at room temperature. Ketone *rac*-6 in THF (8 mL) was added and finally the solution was heated to reflux for 2 h. The reaction was cooled to ambient temperature, diluted with Et₂O (50 mL) and brine (50 mL) was added. The layers were separated, the aqueous layer was extracted with Et₂O (3 x 20 mL), the combined organic fractions were dried (MgSO₄), filtered and the solvent was evaporated. Purification of the crude residue by flash chromatography (hexane : ethylacetate = 20 : 1 to 5 : 1) yielded ester *rac*-7a (1.63 g, 80% for both diastereomers, *E:Z* = 1.5:1) as a colorless oil. Separation of the diastereomers was performed by HPLC.

(*E*)-7a: ¹H NMR (400 MHz, CDCl₃) δ = 4.16 (q, *J* = 7.1 Hz, 2H), 3.94-3.89 (m, 4H), 3.30-3.20 (m, 1H), 2.87-2.73 (m, 2H), 2.46 (q, *J* = 7.8 Hz, 1H), 2.20-2.08 (m, 1H), 1.91-1.82 (m, 4H), 1.81-1.72 (m, 3H), 1.53-1.43 (m, 1H), 1.28 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 168.7 (C), 163.9 (C), 119.0 (C), 118.8 (C), 64.9 (CH₂), 64.0 (CH₂), 59.9 (CH₂), 50.0 (CH), 47.6 (CH), 34.5 (CH₂), 33.9 (CH₂), 28.2 (CH₂), 27.1 (CH₂), 16.2 (CH₃), 14.4 (CH₃).

IR [cm⁻¹]: 2957, 1706, 1269, 1185, 1095.

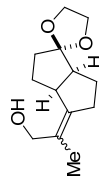
HRMS (EI) (*m/z*): [M]⁺ calcd. for C₁₅H₂₂O₄: 266.1518, found: 266.1512.

(*Z*)-7a: ¹H NMR (400 MHz, CDCl₃) δ = 4.17 (q, *J* = 7.1 Hz, 2H), 3.94-3.89 (m, 4H), 3.77-3.68 (m, 1H), 2.57-2.35 (m, 3H), 2.33-2.22 (m, 1H), 1.88-1.80 (m, 4H), 1.79-1.69 (m, 3H), 1.45-1.35 (m, 1H), 1.29 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 168.1 (C), 163.4 (C), 119.0 (C), 118.6 (C), 64.9 (CH₂), 64.0 (CH₂), 59.9 (CH₂), 51.3 (CH), 47.1 (CH), 34.8 (CH₂), 33.8 (CH₂), 29.8 (CH₂), 25.4 (CH₂), 16.4 (CH₃), 14.3 (CH₃).

IR [cm⁻¹]: 2959, 1709, 1269, 1206, 1106, 1087.

HRMS (EI) (*m/z*): [M]⁺ calcd. for C₁₅H₂₂O₄: 266.1518, found: 266.1511.



Allyl alcohol *rac*-8a. Ester *rac*-7a (1.5 g, 5.63 mmol, 1 equiv.) was dissolved in CH₂Cl₂ (19 mL) and cooled to -50 °C. DIBAL-H (1.5 M in toluene, 7.5 mL, 11.26 mmol, 2.0 equiv.) was added *via* syringe and the solution was warmed to -30 °C within 2 h. The reaction mixture was diluted with CH₂Cl₂ (100 mL), quenched by the addition of Na-K-tartrate (70 mL) and stirred over night. The aqueous layer was extracted with CH₂Cl₂ (3 x 20 mL), dried over MgSO₄ and concentrated. Purification by flash chromatography (hexane : ethylacetate = 3 : 1 to 1 : 1) gave allyl alcohol **9** (1.10 g, 93%) as a colorless oil. Purification by HPLC gave pure *rac*-(*E*)-**8a** and *rac*-(*Z*)-**8a**.

(*E*)-8a: ¹H NMR (400 MHz, CDCl₃) δ = 4.11 (d, *J* = 11.4 Hz, 1H), 4.05 (d, *J* = 11.4 Hz, 1H), 3.95-3.88 (m, 4H), 3.13 (bq, *J* = 7.8 Hz, 1H), 2.46 (dt, *J* = 8.5, 6.2 Hz, 1H), 2.40-2.32 (m, 2H), 2.12-2.01 (m, 1H), 1.85-1.63 (m, 4H), 1.74 (s, 3H), 1.49-1.39 (m, 1H), 1.24 (bs, 1H).

¹³C NMR (100 MHz, CDCl₃) δ = 146.2 (C), 124.9 (C), 118.9 (C), 65.2 (CH₂), 64.9 (CH₂), 64.0 (CH₂), 50.4 (CH), 45.0 (CH), 34.9 (CH₂), 30.1 (CH₂), 29.0 (CH₂), 27.0 (CH₂), 16.7 (CH₃).

IR [cm⁻¹]: 3426 (b), 2955, 1034, 947.

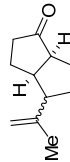
HRMS (EI) (*m/z*): [M-H₂O]⁺ calcd. for C₁₃H₁₈O₂: 206.1307, found: 206.1303.

(*Z*)-8a: ¹H NMR (400 MHz, CDCl₃) δ = 4.15 (d, *J* = 11.3 Hz, 1H), 4.08 (d, *J* = 11.4 Hz, 1H), 3.94-3.89 (m, 4H), 3.21 (bq, *J* = 7.9 Hz, 1H), 2.47 (q, *J* = 7.9 Hz, 1H), 2.42-2.23 (m, 2H), 2.10-2.00 (m, 1H), 1.90-1.82 (m, 1H), 1.78-1.66 (m, 6H), 1.45-1.36 (m, 1H), 1.26 (bs, 1H).

¹³C NMR (100 MHz, CDCl₃) δ = 146.4 (C), 124.8 (C), 118.9 (C), 65.0 (CH₂), 64.9 (CH₂), 64.0 (CH₂), 51.1 (CH), 44.3 (CH), 35.0 (CH₂), 31.1 (CH₂), 30.6 (CH₂), 26.4 (CH₂), 16.9 (CH₃).

IR [cm⁻¹]: 3393 (b), 2931, 1455, 1119.

HRMS (EI) (*m/z*): [M-H₂O]⁺ calcd. for C₁₃H₁₈O₂: 206.1307, found: 206.1304.



Ketone *rac*-9a. To a solution of PPh₃ (935 mg, 3.55 mmol, 1.6 equiv.) in 9 mL THF (0.4 M) was slowly added DEAD (620 mg, 3.55 mmol, 1.6 equiv.) at -15 °C. The orange mixture was stirred

for 15 min at -15 °C, cooled to -40 °C and allylalcohol *rac*-(*E*)-**8a** (500 mg, 2.25 mmol, 1 equiv.) was added *via* syringe (0.4 M in THF). After 5 min *o*-nitrobenzenesulfonylhydrazide (NBSH, 680 mg, 3.10 mmol, 1.4 equiv.) in THF (0.4 M) was added to the white suspension and the temperature was raised to -15 °C within 1 h. Stirring was continued at this temperature for 1 h and finally warmed to ambient temperature over night. TLC analysis showed complete consumption of the starting material and the reaction mixture was concentrated to about 10 mL. The residue was passed through a plug of silica twice, concentrated and dissolved in acetone/H₂O (23 mL, 2.4:1). PPTS (170 mg, 0.70 mmol, 0.3 equiv.) was added and the mixture was refluxed for 50 min. The solvent was evaporated and the crude product was directly loaded onto a silica column. Purification by flash chromatography (hexane : ethylacetate = 50 : 1 to 10 : 1) afforded a 1.2:1 mixture¹ of *rac*-**9a** and its C1-diastereomer (330 mg, 68 % 2 steps), which was easily separated by HPLC.

9a: ¹H NMR (400 MHz, CDCl₃) δ = 4.86-4.83 (m, 1H), 4.70 (s, 1H), 2.95 (qu, *J* = 8.4 Hz, 1H), 2.67 (t, *J* = 10.1 Hz, 1H), 2.57 (qu, *J* = 6.3 Hz, 1H), 2.33-2.13 (m, 2H), 1.97-1.75 (m, 3H), 1.79 (s, 3H), 1.65 (qu, *J* = 6.2 Hz, 1H), 1.48-1.32 (m, 2H).

¹³C NMR (100 MHz, CDCl₃) δ = 222.8 (C), 144.6 (C), 110.3 (CH₂), 51.1 (CH), 50.8 (CH), 41.7 (CH), 39.4 (CH₂), 28.8 (CH₂), 27.2 (CH₂), 23.3 (CH₃), 21.6 (CH₂).

IR [cm⁻¹]: 2956, 1738, 1647, 1458, 1092.

HRMS (EI) (m/z): [M]⁺ calcd. for C₁₁H₁₆O: 164.1201, found: 164.1198

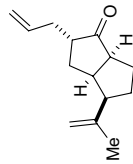
C1-diastereomer of **9a:** ¹H NMR (400 MHz, CDCl₃) δ = 4.79-4.75 (m, 1H), 4.75-4.71 (m, 1H), 2.68-2.57 (m, 2H), 2.35-2.26 (m, 2H), 2.15-1.99 (m, 3H), 1.88- 1.65 (m, 3H), 1.73 (s, 3H), 1.60-1.49 (m, 1H).

¹³C NMR (100 MHz, CDCl₃) δ = 223.4 (C), 146.1 (C), 110.4 (CH₂), 53.0 (CH), 52.2 (CH), 45.2 (CH), 36.2 (CH₂), 32.4 (CH₂), 27.6 (CH₂), 23.7 (CH₂), 20.3 (CH₃).

IR [cm⁻¹]: 2954, 1738, 1644, 1456, 1410, 1135.

HRMS (EI) (m/z): [M]⁺ calcd. for C₁₁H₁₆O: 164.1201, found: 164.1198.

¹ *rac*-(*Z*)-**8a** afforded similarly high yield but a 10:1 ratio in favor of *rac*-**9a**.



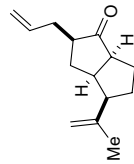
Ketone *rac*-5a. KHMDS (0.5 M in toluene, 2.0 mL, 1.02 mmol, 1.05 equiv.) was diluted with THF (6.4 mL) and cooled to -78 °C. Ketone *rac*-**9a** (160 mg, 0.97 mmol, 1 equiv.) in 2 mL THF (0.5 M) was added dropwise over a period of 5 min. After an additional 40 min at -78 °C HMPA (2 mL) was added, followed by the rapid addition of allyliodide (713 μL, 7.79 mmol, 8 equiv.). The reaction mixture was quenched after 30 min at -78 °C with saturated aqueous NaHCO₃ solution (10 mL) and the aqueous layer was extracted with Et₂O (3 x 10 mL). The combined organic fractions were washed with brine, dried (MgSO₄) and evaporated to give an oil, which was purified by gradient flash chromatography (hexane : ethylacetate = 20 : 1 to 15 : 1) to afford *rac*-**5a** (179 mg, 90%).

¹H NMR (400 MHz, CDCl₃) δ = 5.73 (ddt, *J* = 17.0, 12.1, 6.0 Hz, 1H), 5.07-4.98 (m, 2H), 4.87-4.85 (m, 1H), 4.71 (s, 1H), 2.96-2.86 (m, 1H), 2.74-2.67 (m, 1H), 2.58 (qu, *J* = 6.5 Hz, 1H), 2.44-2.35 (m, 1H), 2.22-2.14 (m, 1H), 2.09-2.00 (m, 1H), 1.96-1.78 (m, 2H), 1.78 (s, 3H), 1.73-1.64 (m, 2H), 1.61-1.53 (m, 1H), 1.41 (dq, *J* = 12.5, 7.6 Hz, 1H).

¹³C NMR (100 MHz, CDCl₃) δ = 223.9 (C), 144.9 (C), 135.7 (CH), 116.5 (CH₂), 110.3 (CH₂), 51.2 (CH), 51.2 (CH), 49.0 (CH), 38.9 (CH), 35.1 (CH₂), 29.9 (CH₂), 27.9 (CH₂), 27.6 (CH₂), 23.4 (CH₃).

IR [cm⁻¹]: 2950, 2869, 1733, 1642, 1451.

HRMS (EI) (m/z): [M]⁺ calcd. for C₁₄H₂₀O: 204.1514, found: 204.1509.



Ketone *rac*-4a. A solution of *rac*-**5a** (50 mg, 0.24 mmol, 1 equiv.) was dissolved in toluene (10 mL) and heated to 80 °C. DBU (29 μL, 0.20 mmol, 0.8 equiv.) was added and stirring was continued for 48 h. The solution was cooled to ambient temperature, the solvent was evaporated under reduced pressure and the crude residue was passed through a plug of silica (hexane :

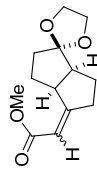
ethylacetate = 10 : 1) to afford 50 mg (99%, dr = 3:1) of *rac*-**4a** and *rac*-**5a**. For analytical purposes a small amount of *rac*-**4a** was purified by HPLC (hexane : ethylacetate = 15 : 1).

¹H NMR (600 MHz, CDCl₃) δ = 5.75 (ddt, *J* = 17.2, 12.1, 5.9 Hz, 1H), 5.05-5.01 (m, 1H), 5.00-4.96 (m, 1H), 4.83-4.81 (m, 1H), 4.67 (s, 1H), 2.85 (qu, *J* = 8.6 Hz, 1H), 2.75 (t, *J* = 9.8 Hz, 1H), 2.54 (qu, *J* = 6.1 Hz, 1H), 2.52-2.47 (m, 1H), 2.39-2.32 (m, 1H), 1.98-1.91 (m, 3H), 1.84-1.76 (m, 1H), 1.79 (s, 3H), 1.61 (qu, *J* = 6.1 Hz, 1H), 1.34 (dq, *J* = 12.5, 7.6 Hz, 1H), 0.97 (dt, *J* = 13.4, 10.4 Hz, 1H).

¹³C NMR (600 MHz, CDCl₃) δ = 223.9 (C), 144.4 (C), 136.1 (CH), 116.1 (CH₂), 110.2 (CH₂), 50.8 (CH), 50.6 (CH), 49.4 (CH), 39.2 (CH), 32.9 (CH₂), 28.2 (CH₂), 28.0 (CH₂), 26.9 (CH₂), 23.3 (CH₃).

IR [cm⁻¹]: 2950, 2869, 1733, 1642, 1451.

HRMS (EI) (*m/z*): [M]⁺ calcd. for C₁₄H₂₀O: 204.1514, found: 204.1507.



Ester (+)-7b. A solution of DIPA (6.3 mL, 45.0 mmol, 2 equiv.) in THF (110 mL) was cooled to -78 °C and treated with BuLi (2.5 M in hexane, 18.0 mL, 45.0 mmol, 2 equiv.). After 15 min methyl trimethylsilylacetate (7.4 mL, 45.0 mmol, 2 equiv.) in THF (45 mL) was added dropwise and the resulting yellow-orange mixture was stirred for 20 min at -78 °C. Ketone (+)-**6** (4.1 g, 22.50 mmol, 1 equiv.) in THF (40 mL) was transferred to the solution *via* syringe and warmed to -15 °C within 1.5 h. As TLC analysis showed complete consumption of the starting material, the mixture was partitioned between saturated aqueous NH₄Cl (150 mL) and Et₂O (100 mL). The aqueous layer was extracted with Et₂O (3 x 80 mL), the combined organic phases were washed with brine, dried over MgSO₄ and concentrated. Flash chromatography (hexane : ethylacetate = 5 : 1) afforded 5.07 g (95%, *Z:E* = 5.1:1) of unsaturated methyl ester (+)-**7b**, which was used without further purification in the next step. For analytical purposes a small amount of (+)-**7b** was purified by HPLC (hexane : ethylacetate = 3 : 1).

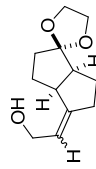
¹H NMR (400 MHz, CDCl₃) δ = 5.72-5.70 (m, 1H), 3.95-3.90 (m, 4H), 3.83-3.74 (m, 1H), 3.68 (s, 3H), 2.66-2.55 (m, 2H), 2.41-2.31 (m, 2H), 1.85-1.73 (m, 3H), 1.70-1.62 (m, 1H), 1.47-1.37 (m, 1H).

¹³C NMR (100 MHz, CDCl₃) δ = 173.1 (C), 166.7 (C), 118.7 (C), 110.7 (CH), 64.9 (CH₂), 64.1 (CH₂), 50.8 (CH₂), 50.6 (CH), 45.7 (CH), 35.9 (CH₂), 35.3 (CH₂), 29.6 (CH₂), 25.1 (CH₂).

IR [cm⁻¹]: 2951, 1717, 1654, 1213, 1137.

HRMS (EI) (*m/z*): [M]⁺ calcd. for C₁₃H₁₈O₄: 238.1205, found: 238.1201.

[α]_D²⁰ = +208.6 (*c* = 0.95, CHCl₃)



Allyl alcohol (+)-8b. Methyl ester (+)-**7b** (4.9 g, 20.56 mmol, 1 equiv., 5.1:1 *Z:E*-mixture) was dissolved CH₂Cl₂ (51 mL) and cooled to -50 °C. DIBAL-H (1.5 M in toluene, 34.3 mL, 51.41 mmol, 2.5 equiv.) was added *via* syringe and the solution was warmed to -30 °C within 1 h. The reaction mixture was diluted with CH₂Cl₂ (100 mL) and finally quenched with saturated aqueous Na-K-tartrate solution (100 mL). The resulting suspension was stirred until a clear phase separation was achieved (3 h). The aqueous layer was extracted with CH₂Cl₂ (3 x 20 mL), dried over MgSO₄ and concentrated. The residue was filtered over a small plug of silica (hexane : ethylacetate = 3 : 1 to 1 : 1) to afford 4.24 g (98%) of (+)-**8b** as an oil. The crude product was used in the next step without further purification.

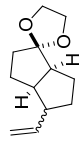
¹H NMR (400 MHz, CDCl₃) δ = 5.48-5.40 (m, 1H), 4.22-4.07 (m, 2H), 3.96-3.86 (m, 4H), 3.22-3.13 (m, 1H), 2.52 (dt, *J* = 8.5, 5.1 Hz, 2H), 2.29-2.18 (m, 1H), 2.10-1.98 (m, 1H), 1.84-1.57 (m, 4H), 1.41 (m, 1H), 1.21 (bs, 1H).

¹³C NMR (100 MHz, CDCl₃) δ = 152.8 (C), 119.0 (CH), 118.6 (C), 64.9 (CH₂), 64.0 (CH₂), 60.7 (CH₂), 50.7 (CH), 43.1 (CH), 35.4 (CH₂), 34.2 (CH₂), 30.4 (CH₂), 26.2 (CH₂).

IR [cm⁻¹]: 3416 (b), 2954, 2879, 1080, 1006.

HRMS (EI) (*m/z*): [M-H₂O]⁺ calcd. for C₁₂H₁₆O₂: 192.1150, found: 192.1155.

[α]_D²⁰ = +155.0 (*c* = 0.40, CHCl₃)



Ketal (+)-16. To a solution of PPh₃ (8.46 g, 32.26 mmol, 1.6 equiv.) in 81 mL THF (0.4 M) was slowly added DEAD (5.62 g, 32.26 mmol, 1.6 equiv.) at -15 °C. The orange mixture was stirred

for 15 min at -15 °C, cooled to -40 °C and allylalcohol (+)-**8b** (4.24 g, 20.16 mmol, 1 equiv.) was added via syringe (0.4 M in THF). After 5 min NBSH (6.13 g, 28.23 mmol, 1.4 equiv.) in THF (0.4 M) was added to the white suspension and the temperature was raised to -15 °C within 1 h. Stirring was continued at this temperature for 1 h and finally warmed to ambient temperature over night. TLC analysis showed complete consumption of the starting material and the reaction mixture was concentrated to 50 mL. The residue was passed through a plug of silica twice (pentane : Et₂O = 5 : 1), the solvent was evaporated and purification by column chromatography and HPLC (hexane : ethylacetate = 15 : 1) afforded 2.14 g (+)-**16** and 0.61 g of its C1-diastereomer (70% for both diastereomers).

(+)-**16**: ¹H NMR (400 MHz, CDCl₃) δ = 5.88 (ddd, *J* = 17.1, 10.5, 6.7 Hz, 1H), 5.06-4.98 (m, 2H), 3.94-3.85 (m, 4H), 2.61 (qu, *J* = 7.8 Hz, 1H), 2.56-2.44 (m, 1H), 1.83-1.76 (m, 1H), 1.72-1.65 (m, 1H), 1.65-1.49 (m, 5H), 1.49-1.41 (m, 1H), 1.41-1.32 (m, 1H).

¹³C NMR (100 MHz, CDCl₃) δ = 139.6 (CH), 118.9 (C), 114.4 (CH₂), 64.9 (CH₂), 64.1 (CH₂), 49.8 (CH), 47.9 (CH), 45.5 (CH), 35.9 (CH₂), 30.2 (CH₂), 26.4 (CH₂), 24.0 (CH₂).

IR [cm⁻¹]: 2953, 2360, 1558, 1094, 1040.

HRMS (EI) (m/z): [M]⁺ calcd. for C₁₂H₁₈O₂: 194.1307, found: 194.1302.

[α]_D²⁰ = +82.0 (*c* = 1.00, CHCl₃)

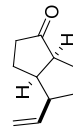
C1-diastereomer of (+)-**16**: ¹H NMR (400 MHz, CDCl₃) δ = 5.76 (ddd, *J* = 17.3, 10.0, 7.4 Hz, 1H), 4.99 (ddd, *J* = 17.2, 1.9, 1.2 Hz, 1H), 4.90 (ddd, *J* = 10.2, 2.0, 0.9 Hz, 1H), 3.94-3.83 (m, 4H), 2.40 (q, *J* = 9.0 Hz, 1H), 2.28-2.19 (m, 1H), 2.12-2.03 (m, 1H), 1.86-1.74 (m, 1H), 1.72-1.66 (m, 5H), 1.59-1.49 (m, 1H), 1.43-1.27 (m, 1H).

¹³C NMR (100 MHz, CDCl₃) δ = 142.2 (CH), 118.7 (C), 113.1 (CH₂), 64.7 (CH₂), 63.7 (CH₂), 52.2 (CH), 49.8 (CH), 47.7 (CH), 34.4 (CH₂), 33.5 (CH₂), 27.8 (CH₂), 26.8 (CH₂).

IR [cm⁻¹]: 2950, 2873, 1638, 1284, 1074.

HRMS (EI) (m/z): [M-H]⁺ calcd. for C₁₂H₁₇O₂: 193.1229, found: 193.1226.

[α]_D²⁰ = +61.3 (*c* = 1.05, CHCl₃)



Ketone (+)-9b. To a solution of (+)-**16** (1.7 g, 8.75 mmol, 1 equiv.) in acetone/H₂O (2.4 : 1, 88 mL) was added PPTS (660 mg, 2.62 mmol, 0.3 equiv.). The mixture was heated to reflux for 40

min, cooled to ambient temperature and diluted with H₂O (50 mL). The aqueous layer was extracted with CH₂Cl₂ (3 x 20 mL), the combined organic fractions were washed with brine (30 mL), dried (MgSO₄) and concentrated. Purification by flash chromatography (pentane : Et₂O = 10 : 1 to 5 : 1) furnished volatile ketone (+)-**9b** (1.23 g, 94%) as a colorless oil.

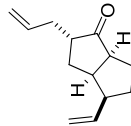
¹H NMR (400 MHz, CDCl₃) δ = 5.87 (ddd, *J* = 17.1, 10.5, 6.8 Hz, 1H), 5.11-5.04 (m, 2H), 2.85 (qu, *J* = 8.2 Hz, 1H), 2.76-2.61 (m, 2H), 2.32-2.14 (m, 2H), 1.96-1.81 (m, 3H), 1.72 (dqu, *J* = 6.4, 2.7 Hz, 1H), 1.60-1.48 (m, 1H), 1.47-1.35 (m, 1H).

¹³C NMR (100 MHz, CDCl₃) δ = 222.7 (C), 138.1 (CH), 115.3 (CH₂), 51.3 (CH), 48.0 (CH), 44.5 (CH), 39.4 (CH₂), 29.5 (CH₂), 28.6 (CH₂), 22.3 (CH₂).

IR [cm⁻¹]: 2955, 2874, 1738, 1654, 1521, 1360.

HRMS (EI) (m/z): [M]⁺ calcd. for C₁₀H₁₄O: 150.1045, found: 150.1042.

[α]_D²⁰ = +151.9 (*c* = 1.00, CHCl₃)



Ketone (+)-5b. KHMDS (0.5 M in toluene, 16.8 mL, 8.39 mmol, 1.05 equiv.) was diluted with THF (65 mL) and cooled to -78 °C. Ketone (+)-**9b** (1.2 g, 7.99 mmol, 1 equiv.) in 16 mL THF (0.5 M) was added dropwise over a period of 10 min. After stirred for an additional 1 h at -78 °C, HMPA (4 mL) was added, followed by the rapid addition of allyliodide (5.84 mL, 63.90 mmol, 8 equiv.). The reaction mixture was quenched with saturated aqueous NaHCO₃ solution after 10 min and the aqueous layer was extracted with Et₂O (3 x 50 mL). The combined organic fractions were washed with brine, dried (MgSO₄) and evaporated to give an oil, which was purified by gradient flash chromatography (hexane : ethylacetate = 20 : 1 to 10 : 1) to afford pure ketone (+)-**5b** (1.36 g, 89%).

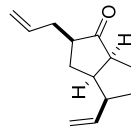
¹H NMR (600 MHz, CDCl₃) δ = 5.89-5.82 (m, 1H), 5.76-5.69 (m, 1H), 5.09-5.00 (m, 4H), 2.83 (dq, *J* = 8.5, 5.5 Hz, 1H), 2.76-2.70 (m, 1H), 2.67 (dt, *J* = 9.3, 2.6 Hz, 1H), 2.43-2.38 (m, 1H), 2.17 (dq, *J* = 8.3, 5.3 Hz, 1H), 2.07-2.01 (m, 1H), 1.94-1.83 (m, 3H), 1.74 (dqu, *J* = 6.4, 2.4 Hz, 1H), 1.66-1.60 (m, 1H), 1.37 (dq, *J* = 11.7, 7.6 Hz, 1H).

¹³C NMR (150 MHz, CDCl₃) δ = 223.8 (C), 138.4 (CH), 135.7 (CH), 116.6 (CH₂), 115.3 (CH₂), 51.9 (CH), 48.9 (CH), 47.7 (CH), 41.5 (CH), 34.9 (CH₂), 30.3 (CH₂), 29.6 (CH₂), 28.2 (CH₂).

IR [cm⁻¹]: 2951, 2871, 1733, 1640, 1116.

HRMS (EI) (m/z): [M]⁺ calcd. for C₁₃H₁₈O: 190.1358, found: 190.1353.

[α]_D²⁰ = +209.8 (c = 1.10, CHCl₃)



Ketone (+)-4b. A solution of ketone (+)-5b (1.24 g, 5.78 mmol, 1 equiv.) was dissolved in toluene (65 mL) and heated to 110 °C. DBU (777 μL, 5.20 mmol, 0.8 equiv.) was added and stirring was continued over night. The solution was cooled to ambient temperature, the solvent was evaporated under reduced pressure and the crude residue was purified by a short silica column (hexane : ethylacetate = 20 : 1) to afford 1.24 g (99%, dr = 2.3:1) of ketone (+)-4b and (+)-5b, which could be used in the next step without further purification. For analytical purposes a small amount was separated by HPLC (hexane : ethylacetate = 30 : 1).

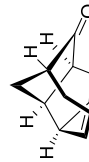
¹H NMR (400 MHz, CDCl₃) δ = 5.89 (ddd, J = 17.1, 10.4, 6.6 Hz, 1H), 5.76 (ddt, J = 17.1, 12.1, 5.9 Hz, 1H), 5.11-4.97 (m, 4H), 2.80-2.71 (m, 2H), 2.71-2.63 (m, 1H), 2.55-2.46 (m, 1H), 2.40-2.30 (m, 1H), 2.10-2.00 (m, 1H), 2.03-1.93 (m, 1H), 1.93-1.77 (m, 2H), 1.67 (qu, J = 6.3 Hz, 1H), 1.33 (dq, J = 11.5, 9.3 Hz, 1H), 1.08 (dt, J = 13.3, 9.6 Hz, 1H).

¹³C NMR (100 MHz, CDCl₃) δ = 221.0 (C), 138.0 (CH), 136.1 (CH), 116.1 (CH₂), 115.3 (CH₂), 51.0 (CH), 49.6 (CH), 48.0 (CH), 42.0 (CH), 32.9 (CH₂), 28.9 (CH₂), 28.8 (CH₂), 28.2 (CH₂).

IR [cm⁻¹]: 2954, 2872, 1733, 1641, 1455.

HRMS (EI) (m/z): [M]⁺ calcd. for C₁₃H₁₈O: 190.1358, found: 190.1354.

[α]_D²⁰ = +40.0 (c = 0.40, CHCl₃)



Tricyclic ketone (+)-3b. The 2.3:1 mixture of (+)-4b and (+)-5b (1.1 g, 5.78 mmol, 1 equiv.) was dissolved in degassed CH₂Cl₂ (1.2 L, 4.5 mM) and heated to reflux. Grubbs II catalyst (245 mg, 0.289 mmol, 0.05 equiv.) in degassed CH₂Cl₂ (20 mL) was added *via* syringe pump within 16 h. After 18 h air was bubbled through the solution for 30 min and the mixture was concentrated. Purification of the dark brown oil by gradient column chromatography (hexane : ethylacetate = 50 : 1 to 15 : 1) yielded tricyclic ketone (+)-3b as a pale-red oil (549 mg, 59%, 84% based on pure diastereomer (+)-4b).

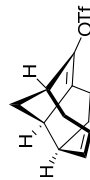
¹H NMR (400 MHz, CDCl₃) δ = 5.35-5.22 (m, 2H), 2.85 (qu, J = 7.7 Hz, 1H), 2.84-2.77 (bs, 1H), 2.65-2.47 (m, 3H), 2.41-2.31 (m, 1H), 2.32-2.23 (m, 1H), 2.05-1.95 (m, 2H), 1.05-1.74 (m, 1H), 1.71-1.60 (m, 2H).

¹³C NMR (100 MHz, CDCl₃) δ = 228.0 (C), 131.6 (CH), 124.8 (CH), 52.1 (CH), 46.4 (CH), 45.0 (2 x CH), 36.2 (CH₂), 34.9 (CH₂), 28.8 (CH₂), 26.2 (CH₂).

IR [cm⁻¹]: 2938, 2358, 1733, 1426, 1102.

HRMS (EI) (m/z): [M]⁺ calcd. for C₁₁H₁₄O: 162.1045, found: 162.1043.

[α]_D²⁰ = +81.7 (c = 1.00, CHCl₃)



Vinyltriflate (+)-17. To a solution of PhNTf₂ (2.29 g, 6.41 mmol, 2 equiv.) and tricyclic ketone (+)-3b (520 mg, 3.21 mmol, 1 equiv.) in THF (65 mL) was added KHMDS (0.5 M in toluene, 12.8 mL, 6.41 mmol, 2 equiv.) at -78 °C. The mixture was stirred for 1 h at -78 °C and 0.5 h at -30 °C. TLC analysis showed complete conversion of the starting material and the reaction mixture was quenched by the addition of NH₄Cl (50 mL). The phases were separated, extracted with a 1 : 1 mixture of Et₂O-hexane (3 x 30 mL), washed with brine, dried over MgSO₄ and evaporated to dryness. The residual oil was diluted with a small amount of hexane and purification by flash chromatography (hexane : ethylacetate = 50 : 1) afforded 753 mg (80%) of vinyltriflate (+)-17 as a colorless oil.

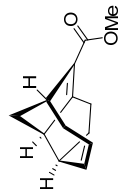
¹H NMR (400 MHz, CDCl₃) δ = 5.21-5.08 (m, 2H), 3.12-3.06 (bs, 1H), 2.95 (t, J = 7.0 Hz, 1H), 2.76-2.69 (b, 1H), 2.59 (bd, J = 18.8 Hz, 1H), 2.47-2.39 (m, 1H), 2.35-2.25 (m, 1H), 2.29-2.19 (m, 1H), 2.24-2.12 (m, 1H), 2.19-2.07 (m, 1H), 1.79 (d, J = 12.8 Hz, 1H), 1.78-1.71 (m, 1H).

¹³C NMR (100 MHz, CDCl₃) δ = 142.9 (C), 139.1 (C), 131.0 (CH), 122.5 (CH), 117.1 (C), 49.2 (CH), 44.5 (CH), 42.8 (CH), 37.5 (CH₂), 32.9 (CH₂), 27.3 (CH₂), 20.7 (CH₂).

IR [cm⁻¹]: 2941, 1420, 1246, 1209, 1142, 1036.

HRMS (EI) (m/z): [M]⁺ calcd. for C₁₂H₁₃O₃F₃S: 294.0538, found: 294.0529.

[α]_D²⁰ = +62.1 (c = 0.63, CHCl₃)



Ester (+)-18. Pd(OAc)₂ (7.8 mg, 0.035 mmol, 0.03 equiv.), PPh₃ (18 mg, 0.069 mmol, 0.06 equiv.) and NEt₃ (229 μL, 2.31 mmol, 2 equiv.) was sequentially added to a solution of vinyltriflate **20** (340 mg, 1.16 mmol, 1 equiv.) in DMF (14 mL) and MeOH (14 mL). CO was bubbled through the orange solution for 10 min. The flask was sealed under 1 atm of CO and heated to 50 °C for 1 h. The solution was cooled to ambient temperature, diluted with Et₂O (100 mL) and washed with brine (2 x 50 mL). The organic phase was dried over MgSO₄, filtered and concentrated. The crude residue was purified by flash chromatography (hexane : ethylacetate = 50 : 1 to 20 : 1) to yield α,β-unsaturated ester (+)-**18** (227 mg, 96%) as a colorless oil.

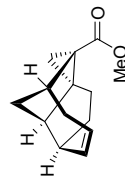
¹H NMR (400 MHz, CDCl₃) δ = 5.18-5.11 (m, 1H), 5.09-5.02 (m, 1H), 3.71 (s, 3H), 3.36-3.30 (bs, 1H), 3.09 (t, *J* = 7.6 Hz, 1H), 2.83 (dddd, *J* = 14.7, 9.0, 5.5 Hz, 1H), 2.73-2.61 (m, 2H), 2.42-2.30 (m, 1H), 2.35-2.21 (m, 2H), 2.10-2.01 (m, 1H), 1.78-1.70 (m, 1H), 1.70 (d, *J* = 12.9 Hz, 1H).

¹³C NMR (100 MHz, CDCl₃) δ = 166.5 (C), 165.9 (C), 130.5 (CH), 126.9 (C), 123.8 (CH), 54.4 (CH), 50.9 (CH₃), 46.4 (CH), 41.2 (CH), 39.7 (CH₂), 33.5 (CH₂), 28.5 (CH₂), 23.8 (CH₂).

IR [cm⁻¹]: 2936, 1706, 1436, 1293, 1109.

HRMS (EI) (m/z): [M]⁺ calcd. for C₁₃H₁₆O₂: 204.1150, found: 204.1146.

[α]_D²⁰ = -52.5 (c = 1.1, CHCl₃)



Tetracyclic Ester (+)-10. To an ice-cooled mixture of NaH (60%, 49 mg, 1.23 mmol, 1.2 equiv.) and trimethylsulfonium iodide (272 mg, 1.23 mmol, 1.2 equiv.) was added DMSO (1.8 mL) dropwise over 5 min. After stirred for 10 min at 10 °C, methylester (+)-**18** (210 mg, 1.03 mmol, 1 equiv.) in DMSO (1 mL, 2 x 0.4 mL rinse) was added *via* syringe. The mixture was stirred for 40 min at ambient temperature, diluted with Et₂O (50 mL), quenched by the addition of saturated aqueous NH₄Cl (50 mL) and extracted with Et₂O/hexane = 1 : 1 (3 x 20 mL). The organic phase was washed with brine (20 mL), the solvent was evaporated and the crude product was filtered over a small plug of silica (hexane : ethylacetate = 20 : 1) to afford tetracyclic ester (+)-**10** (222 mg, 99%) as a colorless oil.

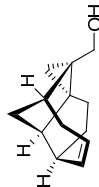
¹H NMR (400 MHz, CDCl₃) δ = 5.32-5.21 (m, 2H), 3.62 (s, 3H), 3.02 (bd, *J* = 17.4 Hz, 1H), 2.67-2.60 (b, 1H), 2.60-2.54 (bs, 1H), 2.32 (t, *J* = 6.4 Hz, 1H), 2.15 (dt, *J* = 13.6, 9.0 Hz, 1H), 2.01-1.93 (m, 2H), 1.89-1.81 (m, 1H), 1.79-1.72 (m, 1H), 1.66-1.54 (m, 2H), 1.35 (d, *J* = 4.5 Hz, 1H), 1.06 (d, *J* = 4.5 Hz, 1H).

¹³C NMR (100 MHz, CDCl₃) δ = 173.8 (C), 132.0 (CH), 124.4 (CH), 50.9 (CH₃), 48.9 (CH), 45.3 (CH), 44.2 (C), 39.2 (CH), 35.9 (C), 34.5 (CH₂), 33.6 (CH₂), 30.5 (CH₂), 24.0 (CH₂), 22.2 (CH₂).

IR [cm⁻¹]: 2946, 1717, 1433, 1338, 1156, 1132.

HRMS (EI) (m/z): [M]⁺ calcd. for C₁₄H₁₈O₂: 218.1307, found: 218.1304.

[α]_D²⁰ = +120.4 (c = 0.45, CHCl₃)



Alcohol (+)-19. Ester (+)-**10** (213 mg, 0.98 mmol, 1 equiv.) in CH₂Cl₂ (5 mL) was cooled to -30 °C and treated with DIBAL (1.5 M in toluene, 1.43 mL, 2.15 mmol, 2.2 equiv.). After 20 min TLC analysis showed complete consumption of the starting material and the temperature was raised to 0 °C. The reaction was diluted with CH₂Cl₂ (50 mL) and quenched by the careful addition of saturated aqueous Na/K-Tartrate (20 mL) solution. Stirring was continued for 2 h and the phases were separated. The aqueous layer was extracted with CH₂Cl₂ (3 x 10 mL), the combined organic fractions were dried (MgSO₄), filtered and the solvent was evaporated to yield

pure primary alcohol (+)-**19** (160 mg, 86%) as a white solid. The crude product was used in the next step without further purification.

¹H NMR (400 MHz, CDCl₃) δ = 5.37-5.23 (m, 2H), 3.76 (dd, J = 11.4, 5.4 Hz, 1H), 3.14 (dd, J = 11.4, 2.4 Hz, 1H), 2.64-2.57 (bs, 1H), 2.56-2.43 (m, 2H), 2.33-2.28 (bt, J = 6.8 Hz, 1H), 2.10-1.93 (m, 2H), 1.79-1.67 (m, 2H), 1.59-1.47 (m, 2H), 1.21-1.15 (bs, 1H), 0.66 (dd, J = 4.8, 1.5 Hz, 1H), 0.57 (d, J = 5.0 Hz, 1H).

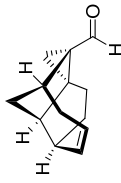
¹³C NMR (100 MHz, CDCl₃) δ = 133.0 (CH), 124.0 (CH), 65.4 (CH₂), 49.3 (CH), 45.4 (CH), 39.7 (CH), 38.5 (C), 33.8 (CH₂), 33.1 (CH₂), 33.0 (C), 30.3 (CH₂), 23.7 (CH₂), 16.3 (CH₂).

IR [cm⁻¹]: 3333, 2933, 2335, 1023.

HRMS (EI) (m/z): [M-H₂O]⁺ calcd. for C₁₃H₁₆: 172.1252, found: 172.1248.

[α]_D²⁰ = +88.7 (c = 0.55, CHCl₃)

mp 85-89 °C



Aldehyde (+)-**20**. Alcohol (+)-**19** (150 mg, 0.79 mmol, 1 equiv.) was dissolved in DMSO (3 mL) and treated with IBX (662 mg, 2.37 mmol, 3 equiv.). After 1 h at room temperature, the reaction mixture was directly loaded onto a silica column. Elution of the product (hexane : ethylacetate = 10 : 1 to 3 : 1) afforded (+)-**20** as a white solid (145 mg, 98%).

¹H NMR (400 MHz, CDCl₃) δ = 9.28 (s, 1H), 5.35-5.22 (m, 2H), 3.07 (bd, J = 18.4 Hz, 1H), 2.71-2.63 (m, 2H), 2.39 (t, J = 6.4 Hz, 1H), 2.12 (bd, J = 18.4 Hz, 1H), 2.03-1.93 (m, 1H), 1.93-1.80 (m, 2H), 1.80-1.71 (m, 1H), 1.72-1.66 (m, 2H), 1.65 (d, J = 5.2 Hz, 1H), 1.20 (d, J = 5.2 Hz, 1H).

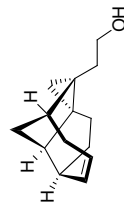
¹³C NMR (100 MHz, CDCl₃) δ = 200.9 (C), 132.5 (CH), 123.6 (CH), 48.4 (CH), 45.4 (CH), 43.1 (C), 38.1 (CH), 33.4 (CH₂), 33.3 (CH₂), 31.4 (CH₂), 24.2 (CH₂), 21.2 (CH₂).

IR [cm⁻¹]: 2945, 1705, 1675, 1306, 1157, 1036.

HRMS (EI) (m/z): [M]⁺ calcd. for C₁₃H₁₆O: 188.1201, found: 188.1200.

[α]_D²⁰ = +287.6 (c = 0.55, CHCl₃)

mp 125-128 °C



Alcohol (+)-**12**. MeOCH₂PPh₃Cl (528 mg, 1.54 mmol, 2 equiv.) was dissolved in 5.1 mL THF and cooled to 0 °C. *t*-BuOK (173 mg, 1.54 mmol, 2 equiv.) was added in one portion and the resulting red-orange suspension was stirred for 1 h at 0 °C. Aldehyde (+)-**20** (145 mg, 0.77 mmol, 1 equiv.) in 1.5 mL THF was added *via* syringe, whereas the solution turned pale-orange. After stirring for 20 min at ambient temperature, the reaction was quenched by slow addition of 6 N HCl (4 mL). As TLC analysis showed complete hydrolysis of the enol ether (1.5 h), the mixture was carefully adjusted to pH = 7 with solid NaHCO₃. NaBH₄ (146 mg, 3.85 mmol, 5 equiv.) was added in 5 portions, stirring was continued for further 10 min and the reaction was finally quenched by 1% HCl. The layers were separated, the aqueous phase was extracted with Et₂O (4 x 5 mL), the combined organic fractions were dried (MgSO₄) and concentrated. Purification by column chromatography (hexane : ethylacetate = 5 : 1 to 2 : 1) yielded solid alcohol (+)-**12** (125 mg, 80%).

¹H NMR (400 MHz, CDCl₃) δ = 5.33-5.20 (m, 2H), 3.86-3.71 (m, 2H), 2.61-2.54 (bs, 1H), 2.36-2.24 (b, 3H), 2.05-1.91 (m, 2H), 1.81-1.67 (m, 2H), 1.68-1.59 (m, 1H), 1.58-1.51 (m, 2H), 1.55-1.47 (m, 1H), 1.39 (bs, OH), 0.87 (dt, J = 14.8, 7.4 Hz, 1H), 0.58 (d, J = 4.8 Hz, 1H), 0.34 (d, J = 4.8 Hz, 1H).

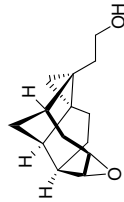
¹³C NMR (100 MHz, CDCl₃) δ = 132.8 (CH), 123.6 (CH), 62.4 (CH₂), 49.0 (CH), 45.3 (CH), 40.2 (CH), 37.2 (C), 33.8 (CH₂), 33.4 (CH₂), 33.1 (CH₂), 30.4 (CH₂), 28.2 (C), 23.2 (CH₂), 16.2 (CH₂).

IR [cm⁻¹]: 3309, 2936, 1637, 1456.

HRMS (EI) (m/z): [M]⁺ calcd. for C₁₄H₂₀O: 204.1514, found: 204.1515.

[α]_D²⁰ = +92.9 (c = 1.00, CHCl₃)

mp 63-69 °C



Epoxide (+)-21. Primary alcohol (+)-**12** (115 mg, 0.56 mmol, 1 equiv.) in acetone (1 mL) was treated with DMDO (0.07 M in acetone, 24 mL, 1.69 mmol, 3.0 equiv.) at room temperature for 30 min. The solvent was evaporated to give pure epoxide (+)-**21** (118 mg, 95%, dr = 16.6:1) as a white solid.

¹H NMR (400 MHz, CDCl₃) δ = 3.87-3.71 (m, 2H), 3.15 (t, *J* = 4.5 Hz, 1H), 2.90 (dt, *J* = 3.8, 1.3 Hz, 1H), 2.71 (t, *J* = 7.4 Hz, 1H), 2.24-2.08 (m, 5H), 1.94-1.85 (m, 1H), 1.83-1.71 (m, 3H), 1.58-1.48 (m, 1H), 1.39 (b, OH), 1.30-1.20 (m, 1H), 0.99 (ddd, *J* = 13.8, 8.6, 6.8 Hz, 1H), 0.57 (dd, *J* = 5.0, 1.8 Hz, 1H), 0.38 (d, *J* = 5.0 Hz, 1H).

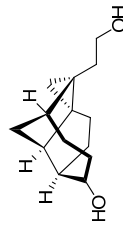
¹³C NMR (100 MHz, CDCl₃) δ = 63.9 (CH), 62.2 (CH₂), 58.2 (CH), 47.5 (CH), 40.3 (CH), 39.8 (CH), 38.8 (C), 33.3 (CH₂), 30.9 (CH₂), 29.8 (C), 29.4 (CH₂), 27.0 (CH₂), 23.8 (CH₂), 16.4 (CH₂).

IR [cm⁻¹]: 3418, 2914, 1718, 1448, 1269.

HRMS (EI) (*m/z*): [M]⁺ calcd. for C₁₄H₂₀O₂: 220.1463, found: 220.1465.

[α]_D²⁰ = +43.6 (*c* = 1.00, CHCl₃)

mp 93-95 °C



Diol (+)-14. To an ice-cold solution of epoxide (+)-**21** (80 mg, 0.36 mmol, 1 equiv.) in THF (7 mL) was added LAH (4M in Et₂O, 454 μL, 1.82 mmol, 5 equiv.) *via* syringe. The solution was warmed to room temperature within 5 min and heated to 50 °C for 2 h. The mixture was cooled to ambient temperature, quenched by careful addition of 1% HCl and the aqueous layer was extracted with CH₂Cl₂ (4 x 10 mL). The combined organic fractions were dried (MgSO₄), filtered and concentrated. Purification by column chromatography (hexane : ethylacetate = 3 : 1 to 1 : 2) gave diol (+)-**14** (58 mg, 73%) as a white solid.

¹H NMR (400 MHz, CDCl₃) δ = 3.87-3.70 (m, 3H), 2.27-2.16 (m, 3H), 2.11-2.00 (m, 1H), 2.00-1.91 (m, 2H), 1.81 (dt, *J* = 12.1, 4.6 Hz, 1H), 1.77-1.23 (m, 9H), 0.97 (ddd, *J* = 13.7, 8.4, 6.7 Hz, 1H), 0.60 (dd, *J* = 4.8, 1.8 Hz, 1H), 0.27 (d, *J* = 4.8 Hz, 1H).

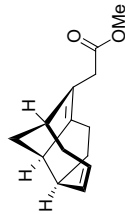
¹³C NMR (100 MHz, CDCl₃) δ = 76.2 (CH), 62.8 (CH₂), 47.8 (CH), 47.1 (CH), 40.0 (CH), 39.5 (C), 32.9 (CH₂), 30.2 (C), 29.6 (CH₂), 29.0 (CH₂), 27.6 (CH₂), 25.0 (CH₂), 24.3 (CH₂), 15.9 (CH₂).

IR [cm⁻¹]: 3390, 2935, 1648, 1549, 1091.

HRMS (EI) (*m/z*): [M]⁺ calcd. for C₁₄H₂₂O₂: 222.1620, found: 222.1621.

[α]_D²⁰ = +43.8 (*c* = 1.10, CHCl₃)

mp 131-133 °C



Ester (+)-11. Flame dried LiOAc (138 mg, 2.10 mmol, 2.5 equiv.), Pd(PPh₃)₄ (97 mg, 0.08 mmol, 0.1 equiv.) and (1-methoxyvinyl)oxytrimethylsilane² (246 mg, 1.68 mmol, 2 equiv.) were sequentially added to a solution of vinyltriflate (+)-**17** (250 mg, 0.84 mmol, 1 equiv.) in degassed THF (8 mL). The mixture was refluxed for 3 h, cooled to ambient temperature and finally H₂O (20 mL) was added. The aqueous layer was extracted with Et₂O (3 x 20 mL), the combined organic fractions were dried and concentrated in *vacuo*. Purification by column chromatography (hexane : ethylacetate = 30 : 1) gave methyl ester (+)-**11** (98 mg, 54%) as a colorless oil.

¹H NMR (400 MHz, CDCl₃) δ = 5.14-5.06 (m, 1H), 5.03-4.95 (m, 1H), 3.67 (s, 3H), 3.06 (d, *J* = 15.2 Hz, 1H), 3.06-3.01 (b, 1H), 2.92 (d, *J* = 15.6 Hz, 1H), 2.88 (bt, *J* = 7.7 Hz, 1H), 2.61-2.54 (m, 1H), 2.38 (bd, *J* = 18.7 Hz, 1H), 2.28-2.04 (m, 5H), 1.70 (d, *J* = 12.6 Hz, 1H), 1.68-1.63 (m, 1H).

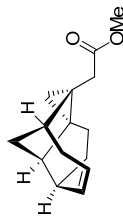
¹³C NMR (100 MHz, CDCl₃) δ = 172.4 (C), 146.7 (C), 132.1 (CH), 127.8 (C), 122.3 (C), 52.7 (CH), 51.8 (CH₃), 47.9 (CH), 42.2 (CH), 38.3 (CH₂), 33.4 (CH₂), 32.8 (CH₂), 29.4 (CH₂), 21.4 (CH₂).

IR [cm⁻¹]: 2931, 1739, 1436, 1271, 1131.

² Oisaki, K.; Suto, Y.; Kanai, M.; Shibasaki, M. *J. Am. Chem. Soc.*, **2003**, 125, 5644-5645.

HRMS (EI) (m/z): [M]⁺ calcd. for C₁₄H₁₈O₂: 218.1307, found: 218.1303.

[α]_D²⁰ = +46.6 (c = 0.49, CHCl₃)



Tetracyclic ester (+)-13. To a solution of methyl ester (+)-11 (86 mg, 0.39 mmol, 1 equiv.) in CH₂Cl₂ (7 mL) at 0 °C was added CH₂I₂ (44 μL, 0.55 mmol, 1.4 equiv.), followed by Et₂Zn (1.1 M in toluene, 890 μL, 0.98 mmol, 2.5 equiv.). After stirring for 1.5 h at 0 °C TLC analysis showed complete consumption of the starting material and the reaction was quenched by the addition of saturated aqueous NH₄Cl (2 mL) solution. After stirring for 30 min with saturated aqueous Na/K-Tartrate solution (5 mL), the aqueous layer was extracted with CH₂Cl₂ (3 x 10 mL). The combined organic phases were dried over MgSO₄ and filtered. Evaporation of the solvent and purification by flash chromatography (hexane : ethylacetate = 30 : 1) afforded tetracyclic ester (+)-13 (80 mg, 89%) as a colorless oil.

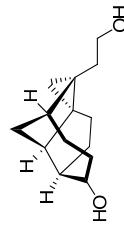
¹H NMR (400 MHz, CDCl₃) δ = 5.34-5.20 (m, 2H), 3.68 (s, 3H), 2.62-2.55 (b, 1H), 2.46 (dd, *J* = 15.4, 1.5 Hz, 1H), 2.46-2.43 (b, 1H), 2.37 (bd, *J* = 17.9 Hz, 1H), 2.29 (t, *J* = 6.2 Hz, 1H), 2.03-1.93 (m, 2H), 1.74-1.64 (m, 2H), 1.59 (d, *J* = 15.7 Hz, 1H), 1.58-1.53 (m, 2H), 1.39 (dt, *J* = 13.1, 9.0 Hz, 1H), 0.71 (dd, *J* = 5.3, 1.5 Hz, 1H), 0.49 (d, *J* = 5.3 Hz, 1H).

¹³C NMR (100 MHz, CDCl₃) δ = 174.1 (C), 133.0 (CH), 123.6 (CH), 51.4 (CH₃), 49.1 (CH), 45.4 (CH), 40.4 (CH), 38.1 (C), 35.6 (C), 33.9 (CH₂), 33.5 (CH₂), 30.3 (CH₂), 28.5 (CH₂), 23.5 (CH₂), 16.6 (CH₂).

IR [cm⁻¹]: 2943, 1743, 1435, 1281, 1160.

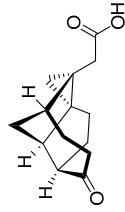
HRMS (EI) (m/z): [M]⁺ calcd. for C₁₅H₂₀O₂: 232.1463, found: 232.1457.

[α]_D²⁰ = +90.3 (c = 0.88, CHCl₃)



Diol (+)-14. Tetracyclic ester (+)-13 (40 mg, 0.17 mmol, 1 equiv.) was treated with DMDO (0.07 M in acetone, 3 mL, 0.21 mmol, 1.2 equiv.) at room temperature for 30 min. The solvent was evaporated to give crude epoxide (dr = 5:1), which was directly used in the next step.

To an ice-cold solution of the epoxide in THF (7 mL) was added LAH (4 M in Et₂O, 344 μL, 1.38 mmol, 8 equiv.) dropwise. The solution was warmed to room temperature within 5 min and heated to 50 °C for 2.5 h. The mixture was cooled to 0 °C, quenched by careful addition of 1% HCl and the aqueous layer was extracted with CH₂Cl₂ (4 x 10 mL). The combined organic fractions were dried (MgSO₄), filtered and concentrated. Purification by column chromatography (hexane : ethylacetate = 3 : 1 to 1 : 2) gave diol (+)-14 (34 mg, 90%, dr = 5:1). Analytical data matched those reported on pp. S17-S18.



Ketoacid (+)-15. Diol (+)-14 (55 mg, 0.25 mmol, 1 equiv.) and IBX (346 mg, 1.24 mmol, 5 equiv.) in DMSO (2 mL) were stirred for 1 h at ambient temperature. The reaction mixture was diluted with Et₂O (50 mL), washed with H₂O (3 x 10 mL) and the aqueous layer was extracted with Et₂O/hexane (1:1, 3 x 10 mL). The combined organic fractions were dried (MgSO₄), filtered over a small plug of silica and concentrated.

The crude product was dissolved in *t*-BuOH (2.5 mL), treated with 2-methyl-2-butene (1 mL / mmol, 0.25 mL), and cooled to 5 °C. NaClO₂ (419 mg, 3.71 mmol, 15 equiv.) and 419 mg NaH₂PO₄ were dissolved in H₂O (2.5 mL, 1.5 M) and added over a period of 5 min. After 15 min at room temperature TLC analysis showed complete consumption and the reaction mixture was separated between CH₂Cl₂ (50 mL) and brine (20 mL). The aqueous layer was extracted with three portions of CH₂Cl₂ (15 mL) and the combined organic extracts were dried over MgSO₄. Evaporation of the solvent gave crude ketoacid, which was purified by flash chromatography (hexane : ethylacetate from 2 : 1 to 1 : 2) to give 41 mg (72 %) of (+)-15 as a white solid.

¹H NMR (600 MHz, CDCl₃) δ = 10.5 (b, OH), 2.93 (t, *J* = 8.5 Hz, 1H), 2.63 (dd, *J* = 15.3, 1.7 Hz, 1H), 2.55 (dt, *J* = 7.2, 3.6 Hz, 1H), 2.47-2.42 (m, 1H), 2.40 (t, *J* = 8.1 Hz, 1H), 2.22-2.17 (m,

1H), 2.15-1.96 (m, 4H), 1.82 (d, $J = 15.5$ Hz, 1H), 1.70-1.63 (m, 2H), 1.59-1.52 (m, 1H), 1.39 (d, $J = 14.4$ Hz, 1H), 0.84 (dd, $J = 5.7, 1.5$ Hz, 1H), 0.63 (d, $J = 6.0$ Hz, 1H).

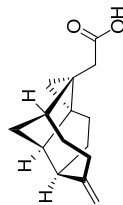
^{13}C NMR (150 MHz, CDCl_3) $\delta = 214.7$ (C), 177.5 (C), 55.5 (CH), 46.6 (CH), 39.7 (CH), 35.9 (CH_2), 35.1 (CH_2), 30.5 (CH_2), 30.2 (C), 28.8 (CH_2), 27.8 (CH_2), 25.3 (CH_2), 16.2 (CH_2).

IR [cm^{-1}]: 2943, 1701, 1697, 1459, 1220, 1063.

HRMS (ESI) (m/z): $[\text{M}-\text{H}]^+$ calcd. for $\text{C}_{14}\text{H}_{17}\text{O}_3$: 233.1178, found: 233.1175.

$[\alpha]_{\text{D}}^{20} = +43.8$ ($c = 1.10$, CHCl_3)

mp 131-133 °C



Echinopine A (1). To a solution of MePPh_3Br (290 mg, 0.811 mmol, 5 equiv.) in THF (2.7 mL) was added *t*-BuOK at room temperature. After 30 min ketoacid (+)-**15** (38 mg, 0.162 mmol, 1 equiv.) in THF (1 mL, 2 x 0.3 mL rinse) was transferred to the yellow-orange solution *via* syringe. The mixture was refluxed for 45 min, cooled to ambient temperature, diluted with Et_2O (50 mL) and quenched by the addition of 0.1% HCl (10 mL). The phases were separated, the aqueous layer was extracted with Et_2O (4 x 10 mL), the combined organic fractions were dried and concentrated. Purification by column chromatography (hexane : ethylacetate = 3 : 1 to 1 : 1) afforded Echinopine A (**1**) (23 mg, 62%) as a white solid. Crystallization from hexane at ambient temperature afforded crystals suitable for X-ray analysis.

^1H NMR (600 MHz, CDCl_3) $\delta = 10.81$ (b, OH), 4.64 (d, $J = 2.6$ Hz, 1H), 4.61 (d, $J = 2.6$ Hz, 1H), 2.81 (dt, $J = 9.3, 2.4$ Hz, 1H), 2.67 (dd, $J = 15.3, 1.7$ Hz, 1H), 2.44 (m, 1H), 2.27 (t, $J = 8.1$ Hz, 1H), 2.17 (m, 1H), 2.13 (dt, $J = 13.2, 4.2$ Hz, 1H), 1.96 (m, 1H), 1.94 (m, 1H), 1.83 (dt, $J = 13.2, 4.2$ Hz, 1H), 1.76 (d, $J = 15.5$ Hz, 1H), 1.66 (dddd, $J = 14.0, 9.7, 6.8, 2.8$ Hz, 1H), 1.53 (ddd, $J = 13.7, 9.7, 4.1$ Hz, 1H), 1.45 (ddd, $J = 13.8, 7.6, 1.5$ Hz, 1H), 1.37 (d, $J = 13.6$ Hz, 1H), 1.28 (ddd, $J = 13.5, 4.2, 2.6$ Hz, 1H), 0.74 (dd, $J = 5.3, 1.5$ Hz, 1H), 0.51 (d, $J = 5.3$ Hz, 1H).

^{13}C NMR (150 MHz, CDCl_3) $\delta = 178.6$ (C), 154.3 (C), 112.1 (CH_2), 48.7 (CH), 48.5 (CH), 41.4 (C), 40.5 (CH), 35.2 (CH_2), 32.7 (CH_2), 31.0 (CH_2), 30.5 (CH_2), 29.9 (C), 29.4 (CH_2), 25.7 (CH_2), 16.1 (CH_2).

IR [cm^{-1}]: 3064, 2930, 2877, 1702, 1699, 1455, 1306 1131.

HRMS (EI) (m/z): $[\text{M}]^+$ calcd. for $\text{C}_{15}\text{H}_{20}\text{O}_2$: 232.1463, found: 232.1464.

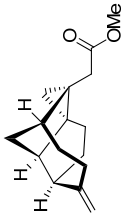
$[\alpha]_{\text{D}}^{22} = +26.1$ ($c = 0.70$, CHCl_3), lit.³ $[\alpha]_{\text{D}}^{22} = +23$ ($c = 0.11$, CHCl_3)

mp 99-102 °C

³ Dong, M.; Cong, B.; Yu, S.-H.; Sauriol, F.; Huo, C.-H.; Shi, Q.-W.; Gu, Y.-C.; Zamir, L. O.; Kiyota, H. *Org. Lett.*, **2008**, 10, 701-704.

Comparison of the NMR Spectral Data of Natural and Synthetic Echinopine A

| | Natural ¹ H-NMR (500 MHz, CHCl ₃) | Synthetic ¹ H-NMR (600 MHz, CHCl ₃) | Natural ¹³ C-NMR (500 MHz, CHCl ₃) | Synthetic ¹³ C-NMR (600 MHz, CHCl ₃) |
|-----|--|--|---|---|
| 1 | 2.81 (dt, <i>J</i> = 9.1, 2.1 Hz) | 2.81 (dt, <i>J</i> = 9.3, 2.4 Hz) | 48.7 | 48.7 |
| 2a | 2.16 (m) | 2.17 (m) | 31.1 | 31.0 |
| 2b | 1.66 (dddd, <i>J</i> = 14.0, 9.7, 6.7, 2.6 Hz) | 1.66 (dddd, <i>J</i> = 14.0, 9.7, 6.8, 2.8 Hz) | | |
| 3a | 1.95 (m) | 1.96 (m) | 25.8 | 25.7 |
| 3b | 1.52 (ddd, <i>J</i> = 13.5, 9.7, 4.9 Hz) | 1.53 (ddd, <i>J</i> = 13.7, 9.7, 4.1 Hz) | | |
| 4 | | | 41.5 | 41.4 |
| 5 | 2.26 (t, <i>J</i> = 8.0 Hz) | 2.27 (t, <i>J</i> = 8.1 Hz) | 48.6 | 48.5 |
| 6a | 1.44 (ddd, <i>J</i> = 13.7, 7.4, 0.9 Hz) | 1.45 (ddd, <i>J</i> = 13.8, 7.6, 1.5 Hz) | 30.5 | 30.5 |
| 6b | 1.36 (d, <i>J</i> = 13.7 Hz) | 1.37 (d, <i>J</i> = 13.6 Hz) | | |
| 7 | 2.43 (m) | 2.44 (m) | 40.6 | 40.5 |
| 8a | 1.93 (m) | 1.94 (m) | 32.8 | 32.7 |
| 8b | 1.27 (ddd, <i>J</i> = 13.6, 4.0, 3.0 Hz) | 1.28 (ddd, <i>J</i> = 13.5, 4.2, 2.6 Hz) | | |
| 9a | 2.12 (dt, <i>J</i> = 13.3, 4.7 Hz) | 2.13 (dt, <i>J</i> = 13.2, 4.2 Hz) | 29.4 | 29.9 |
| 9b | 1.82 (dt, <i>J</i> = 13.3, 4.2 Hz) | 1.83 (dt, <i>J</i> = 13.2, 4.2 Hz) | | |
| 10 | | | 154.3 | 154.3 |
| 11a | 2.66 (dd, <i>J</i> = 15.3, 1.3 Hz) | 2.67 (dd, <i>J</i> = 15.3, 1.7 Hz) | 35.2 | 35.2 |
| 11b | 1.75 (d, <i>J</i> = 15.3 Hz) | 1.76 (d, <i>J</i> = 15.5 Hz) | | |
| 12 | | | 178.2 | 178.6 |
| 13 | | | 30.0 | 29.9 |
| 14a | 4.63 (d, <i>J</i> = 2.6 Hz) | 4.64 (d, <i>J</i> = 2.6 Hz) | 112.1 | 112.1 |
| 14b | 4.60 (d, <i>J</i> = 2.6 Hz) | 4.61 (d, <i>J</i> = 2.6 Hz) | | |
| 15a | 0.73 (dd, <i>J</i> = 5.3, 1.3 Hz) | 0.74 (dd, <i>J</i> = 5.3, 1.5 Hz) | 16.1 | 16.1 |
| 15b | 0.50 (d, <i>J</i> = 5.3 Hz) | 0.51 (d, <i>J</i> = 5.3 Hz) | | |
| OH | 10.53 (b) | 10.81 (b) | | |



Echinopine B (2). A solution CH₂N₂ (1 M in Et₂O, 130 μ L, 0.13 mmol, 5 equiv.) was added dropwise to Echinopine A (6 mg, 0.026 mmol, 1 equiv.) in Et₂O (1 mL). After 30 min excess CH₂N₂ and solvent was removed under reduced pressure to afford 6 mg (95%) of pure Echinopine B **2** as a colorless oil.

¹H NMR (600 MHz, CDCl₃) δ = 4.63 (d, *J* = 2.6 Hz, 1H), 4.61 (d, *J* = 2.6 Hz, 1H), 3.68 (s, 3H), 2.81 (dt, *J* = 9.1, 1.9 Hz, 1H), 2.63 (dd, *J* = 15.0, 1.4 Hz, 1H), 2.39 (m, 1H), 2.26 (t, *J* = 8.1 Hz, 1H), 2.15 (m, 1H), 2.12 (m, 1H), 1.94 (m, 1H), 1.93 (m, 1H), 1.83 (dt, *J* = 13.2, 4.2 Hz, 1H), 1.73 (d, *J* = 15.1 Hz, 1H), 1.65 (dddd, *J* = 14.0, 9.7, 6.7, 2.8 Hz, 1H), 1.53 (ddd, *J* = 13.5, 9.7, 3.9 Hz, 1H), 1.44 (ddd, *J* = 14.0, 7.4, 1.1 Hz, 1H), 1.35 (d, *J* = 13.6 Hz, 1H), 1.26 (ddt, *J* = 13.5, 4.0, 3.1 Hz, 1H), 0.69 (dd, *J* = 5.3, 1.3 Hz, 1H), 0.47 (d, *J* = 5.3 Hz, 1H).

¹³C NMR (150 MHz, CDCl₃) δ = 173.7 (C), 154.4 (C), 112.0 (CH₂), 51.5 (CH₃), 48.7 (CH), 48.6 (CH), 41.4 (C), 40.6 (CH), 35.3 (CH₂), 32.8 (CH₂), 31.1 (CH₂), 30.5 (CH₂), 30.1 (C), 29.5 (CH₂), 25.7 (CH₂), 16.0 (CH₂).

IR [cm⁻¹]: 2931, 1745, 1636, 1434, 1272, 1166.

HRMS (EI) (m/z): [M]⁺ calcd. for C₁₆H₂₂O₂: 246.1620, found: 246.1611.

[α]_D²² = +21.6 (*c* = 0.60, CHCl₃), lit. [α]_D²² = +21 (*c* = 0.14, CHCl₃)

Comparison of the NMR Spectral Data of Natural and Synthetic Echinopine B

| | Natural ¹ H-NMR (500 MHz, CHCl ₃) | Synthetic ¹ H-NMR (600 MHz, CHCl ₃) | Natural ¹³ C-NMR (500 MHz, CHCl ₃) | Synthetic ¹³ C-NMR (600 MHz, CHCl ₃) |
|-----|--|--|---|---|
| 1 | 2.80 (br, <i>J</i> = 10 Hz) | 2.81 (dt, <i>J</i> = 9.3, 2.4 Hz) | 48.5 | 48.7 |
| 2a | 2.14 (m) | 2.15 (m) | 30.9 | 31.1 |
| 2b | 1.65 (m) | 1.65 (dddd, <i>J</i> = 14.0, 9.7, 6.7, 2.8 Hz) | | |
| 3a | 1.93 (m) | 1.93 (m) | 25.4 | 25.7 |
| 3b | 1.51 (m) | 1.53 (ddd, <i>J</i> = 13.5, 9.7, 3.9 Hz) | | |
| 4 | | | 29.8 | 30.1 |
| 5 | 2.25 (t, <i>J</i> = 9 Hz) | 2.26 (t, <i>J</i> = 8.1 Hz) | 48.4 | 48.6 |
| 6a | 1.44 (m) | 1.45 (ddd, <i>J</i> = 14.0, 7.4, 1.1 Hz) | 30.3 | 30.5 |
| 6b | 1.35 (d, <i>J</i> = 13.8 Hz) | 1.35 (d, <i>J</i> = 13.6 Hz) | | |
| 7 | 2.39 (m) | 2.39 (m) | 40.4 | 40.6 |
| 8a | 1.93 (m) | 1.94 (m) | 32.5 | 32.8 |
| 8b | 1.26 (m) | 1.26 (ddt, <i>J</i> = 13.5, 4.0, 3.1 Hz) | | |
| 9a | 2.11 (m) | 2.12 (m) | 29.3 | 29.5 |
| 9b | 1.82 (dt, <i>J</i> = 13.2, 4.0 Hz) | 1.83 (dt, <i>J</i> = 13.2, 4.2 Hz) | | |
| 10 | | | 154.3 | 154.4 |
| 11a | 2.65 (dd, <i>J</i> = 15.1, 1.3 Hz) | 2.63 (dd, <i>J</i> = 15.0, 1.4 Hz) | 35.1 | 35.3 |
| 11b | 1.72 (d, <i>J</i> = 15.1 Hz) | 1.73 (d, <i>J</i> = 15.2 Hz) | | |
| 12 | | | 173.7 | 173.7 |
| 13 | | | 41.0 | 41.3 |
| 14a | 4.63 (d, <i>J</i> = 2.6 Hz) | 4.63 (d, <i>J</i> = 2.6 Hz) | 111.9 | 112.0 |
| 14b | 4.60 (d, <i>J</i> = 2.6 Hz) | 4.61 (d, <i>J</i> = 2.6 Hz) | | |
| 15a | 0.69 (dd, <i>J</i> = 5.1, 1.3 Hz) | 0.69 (dd, <i>J</i> = 5.3, 1.3 Hz) | 15.7 | 16.0 |
| 15b | 0.47 (d, <i>J</i> = 5.1 Hz) | 0.47 (d, <i>J</i> = 5.3 Hz) | | |
| OMe | 3.68 (s) | 3.68 (s) | 51.3 | 51.5 |

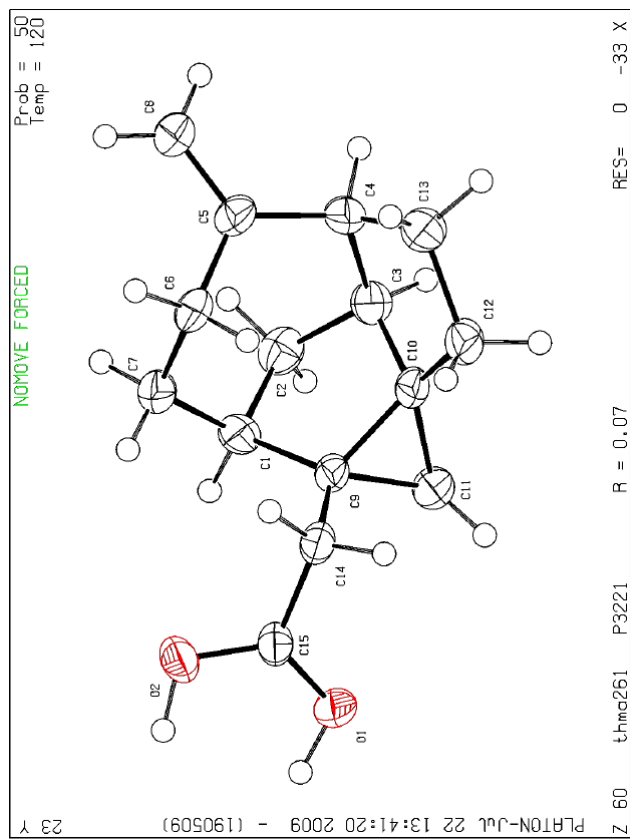
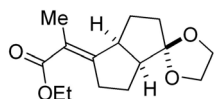


Figure 1: ORTEP view of Echinopine A with labeling scheme. The thermal ellipsoids are drawn at 50% probability level.

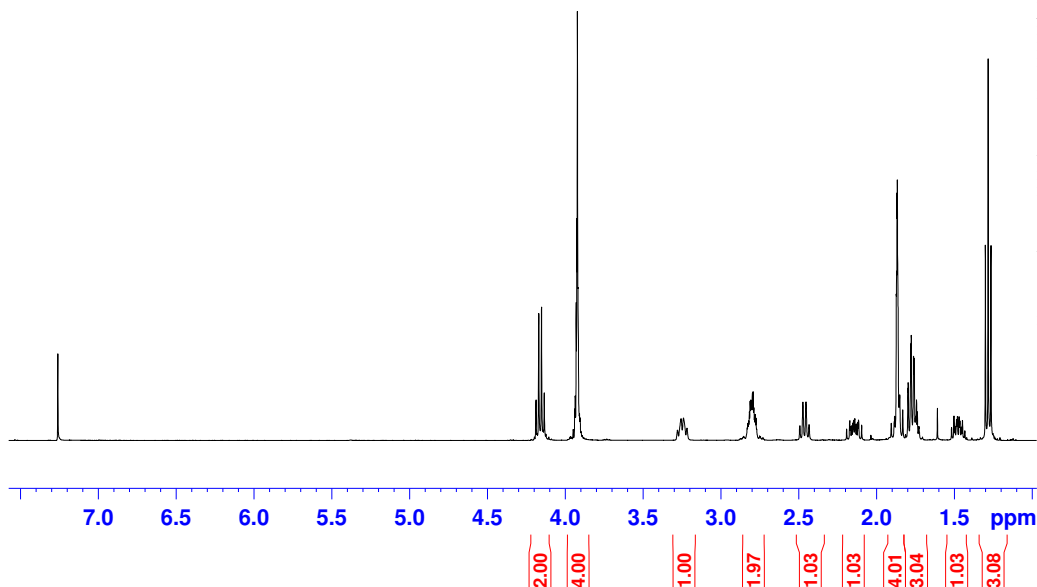
Compound E-7a



```

NAME      THMA E067
EXPNO     10
PROCNO    1
Date_     20090403
Time      14.17
INSTRUM   avance400
PROBHD    5 mm BBO BB-1H
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        8278.146 Hz
FIDRES     0.126314 Hz
AQ         3.9584243 sec
RG         161.3
DW         60.400 usec
DE         6.00 usec
TE         300.2 K
D1         1.00000000 sec
D10        1

```



```

===== CHANNEL f1 =====
NUC1      1H
P1         8.75 usec
PL1        -3.00 dB
SFO1      400.1324710 MHz
SI         32768
SF         400.1300094 MHz
WDW        EM
SSB         0
LB         0.30 Hz
GB          0
PC         1.00

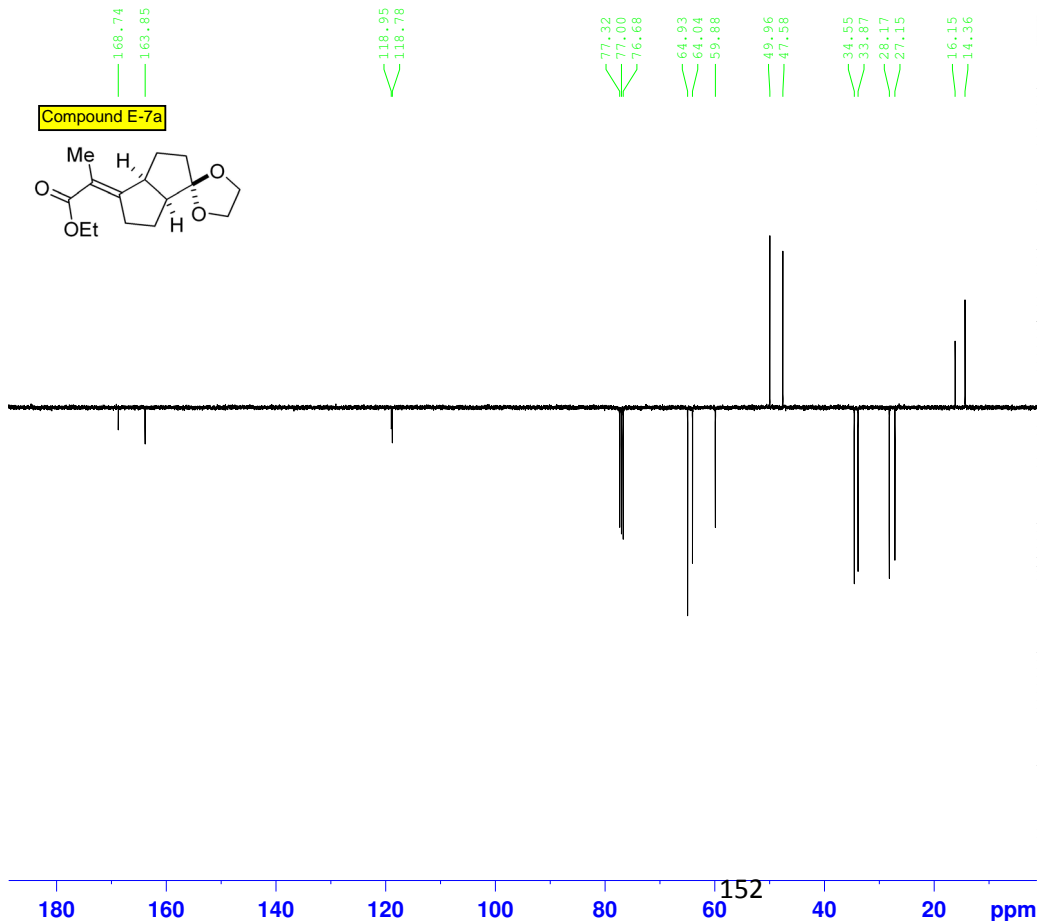
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```

NAME      THMA E067
EXPNO     11
PROCNO    1
Date_     20090403
Time      15.26
INSTRUM   avance400
PROBHD    5 mm BBO BB-1H
PULPROG   jmod
TD         65536
SOLVENT   CDCl3
NS         1200
DS         2
SWH       25062.656 Hz
FIDRES     0.382426 Hz
AQ         1.3074932 sec
RG         5160.6
DW         19.950 usec
DE         6.00 usec
TE         300.2 K
CNST2      145.0000000
CNST11     1.0000000
D1          2.0000000 sec
d20         0.00689655 sec
DELTA       0.00001311 sec
D10         1

```



```

===== CHANNEL f1 =====
NUC1      13C
P1         10.30 usec
p2         20.60 usec
PL1        -1.00 dB
SFO1      100.6233329 MHz

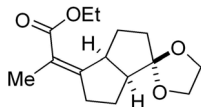
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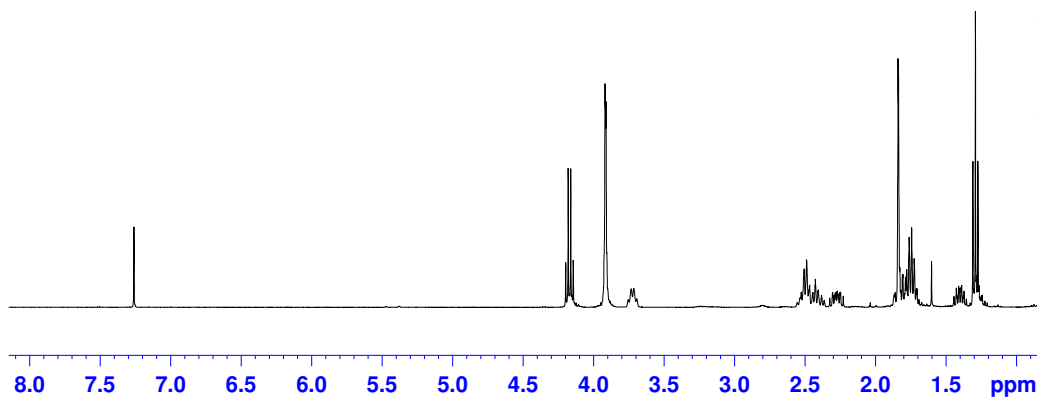
===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2       1H
PCPD2      100.00 usec
PL2        -3.00 dB
PL12       16.00 dB
SFO2      400.1316005 MHz
SI         32768
SF         100.6127704 MHz
WDW        EM
SSB         0
LB         1.00 Hz
GB          0
PC         1.40

```

Compound Z-7a

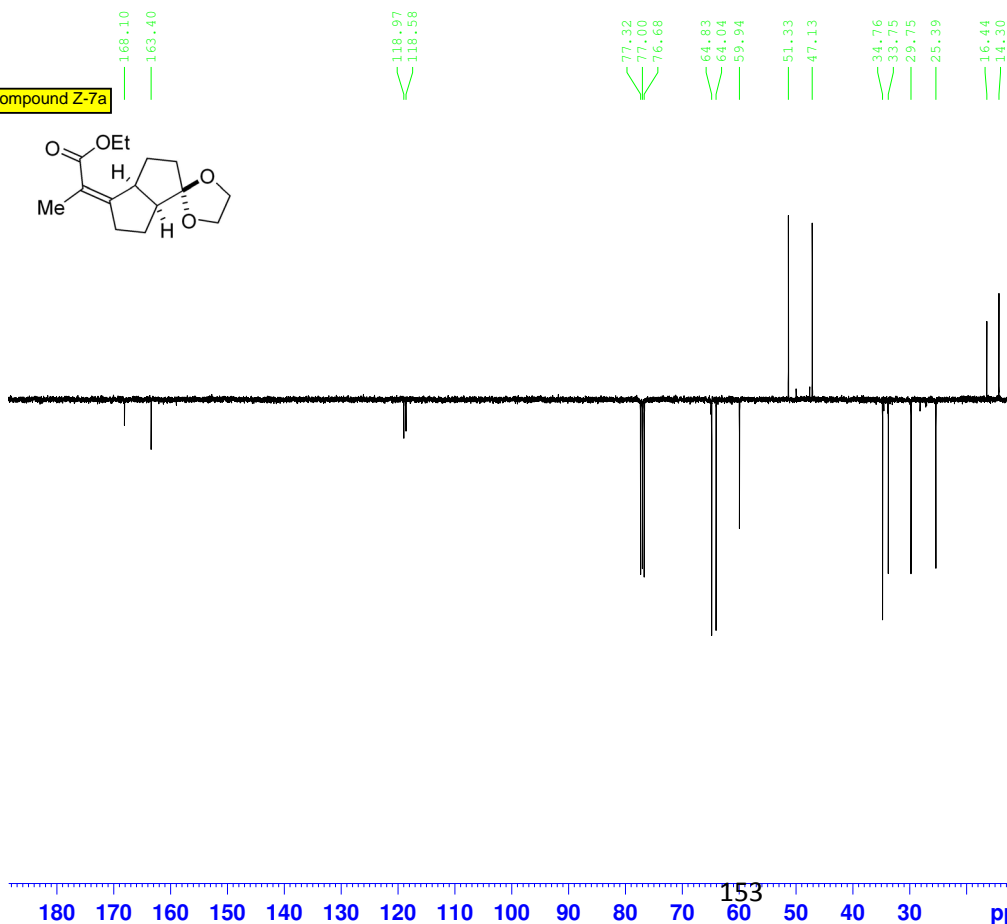
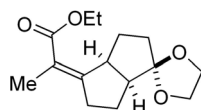


NAME THMA E067
EXPNO 20
PROCNO 1
Date_ 20090404
Time 2.39
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 181
DW 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.00000000 sec
TD0 1



===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 -3.00 dB
SFO1 400.1324710 MHz
SI 32768
SF 400.1300092 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

Compound Z-7a

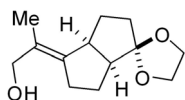


NAME THMA E067
EXPNO 21
PROCNO 1
Date_ 20090404
Time 3.46
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 1200
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3074932 sec
RG 4096
DW 19.950 usec
DE 6.00 usec
TE 300.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
d20 0.00689655 sec
DELTA 0.00001311 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.30 usec
p2 20.60 usec
PL1 -1.00 dB
SFO1 100.6233329 MHz

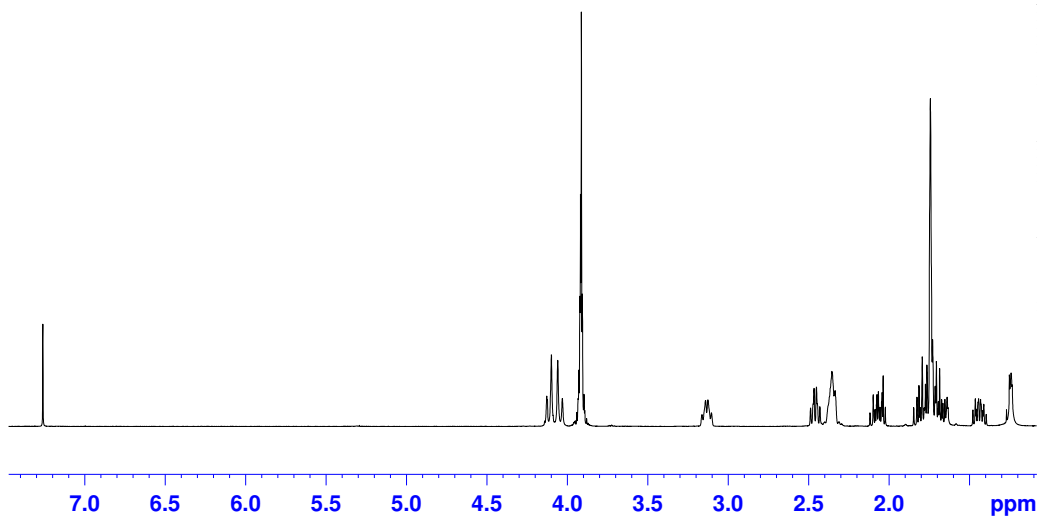
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -3.00 dB
PL12 16.00 dB
SFO2 400.1316005 MHz
SI 32768
SF 100.6127701 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 3.00

Compound E-8a

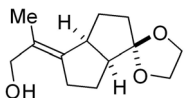


NAME THMA E079
EXPNO 30
PROCNO 1
Date_ 20090404
Time 5.47
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 161.3
DW 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 -3.00 dB
SFO1 400.1324710 MHz
SI 32768
SF 400.1300092 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



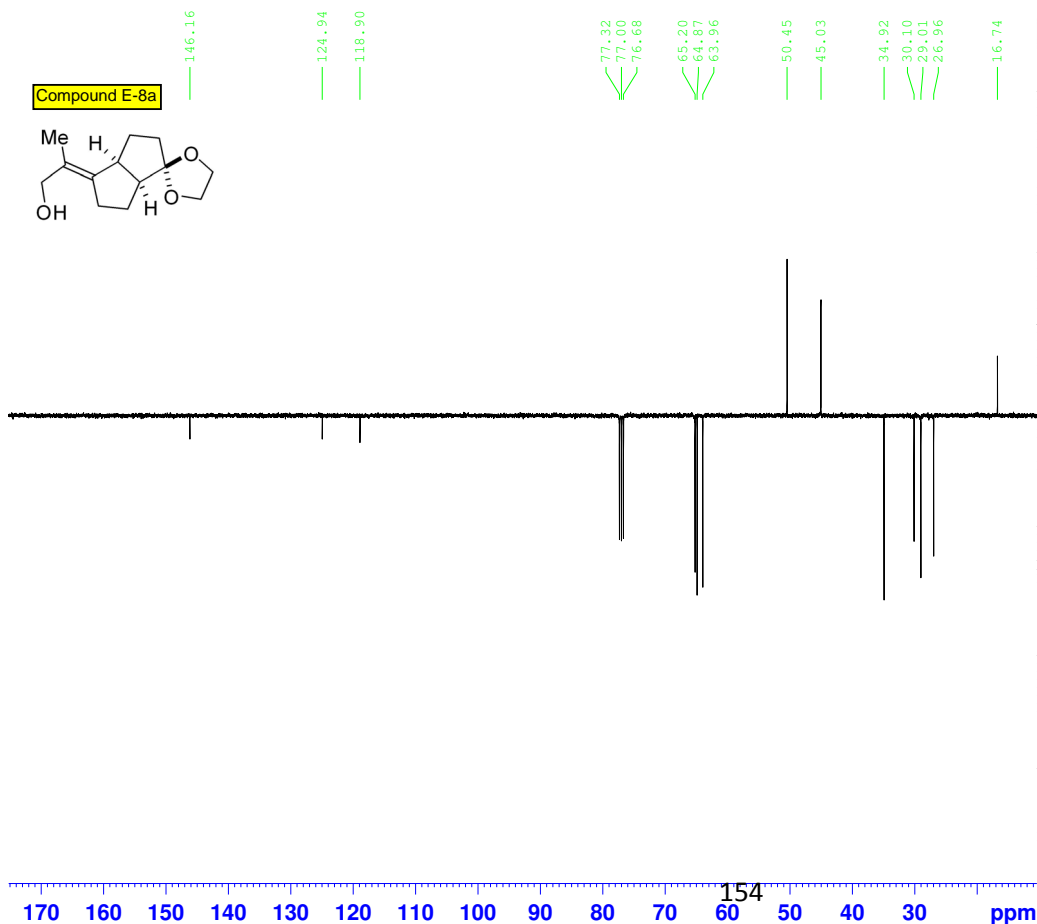
Compound E-8a



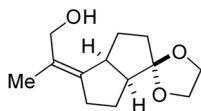
NAME THMA E079
EXPNO 31
PROCNO 1
Date_ 20090404
Time 6.55
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 1200
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3074932 sec
RG 3251
DW 19.950 usec
DE 6.00 usec
TE 300.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
d20 0.00689655 sec
DELTA 0.00001311 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.30 usec
p2 20.60 usec
PL1 -1.00 dB
SFO1 100.6233329 MHz

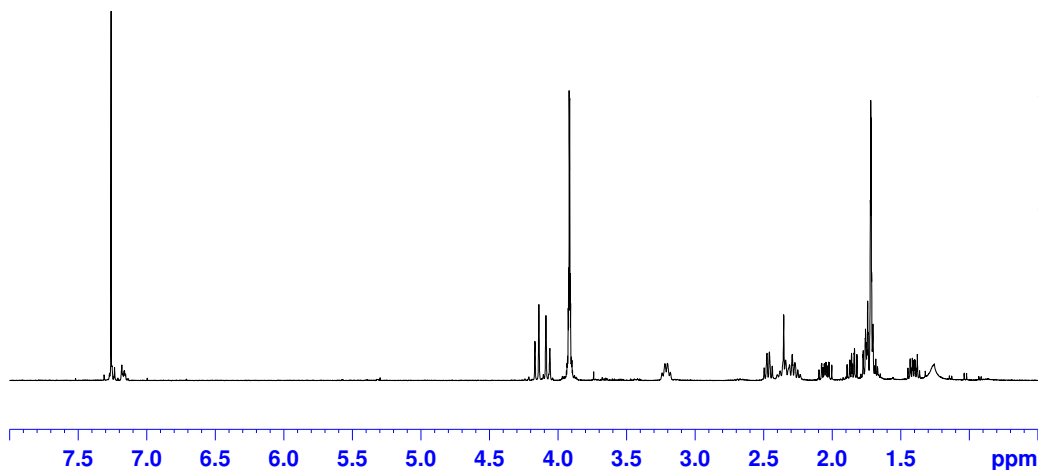
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -3.00 dB
PL12 16.00 dB
SFO2 400.1316005 MHz
SI 32768
SF 100.6127706 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 3.00



Compound Z-8a

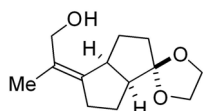


NAME THMAE KTE7
EXPNO 340
PROCNO 1
Date_ 20090407
Time 17.21
INSTRUM AVIII400
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 4
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 575
DW 60.800 usec
DE 6.50 usec
TE 298.2 K
D1 1.00000000 sec
TD0 1

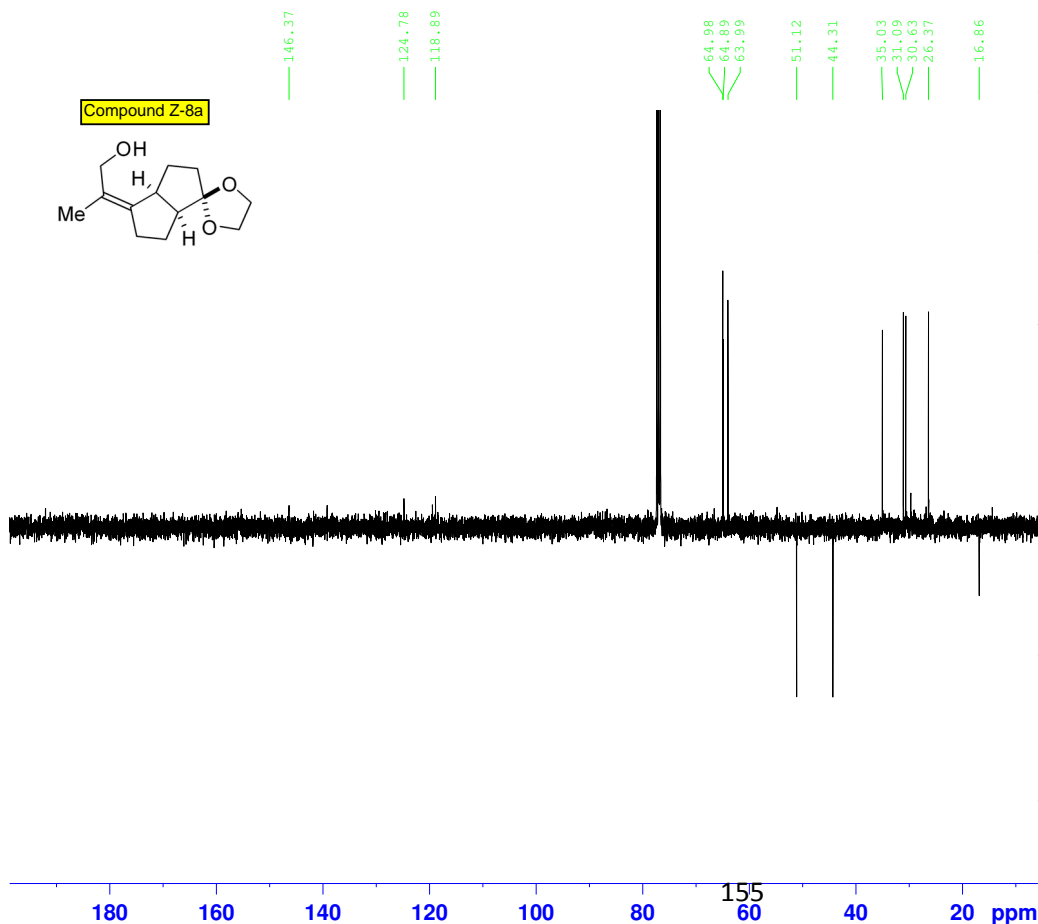


===== CHANNEL f1 =====
NUC1 1H
P1 13.50 usec
PL1 -1.80 dB
PL1W 15.28361320 W
SFO1 400.2724718 MHz
SI 32768
SF 400.2700104 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

Compound Z-8a



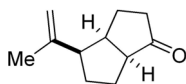
NAME THMAE KTE7
EXPNO 171
PROCNO 1
Date_ 20090721
Time 3.11
INSTRUM AVIII400
PROBHD 5 mm PABBO BB-
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 7000
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 298.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
D20 0.00689655 sec
TD0 1



===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
P2 20.00 usec
PL1 -1.50 dB
PL1W 47.89980698 W
SFO1 100.6580364 MHz

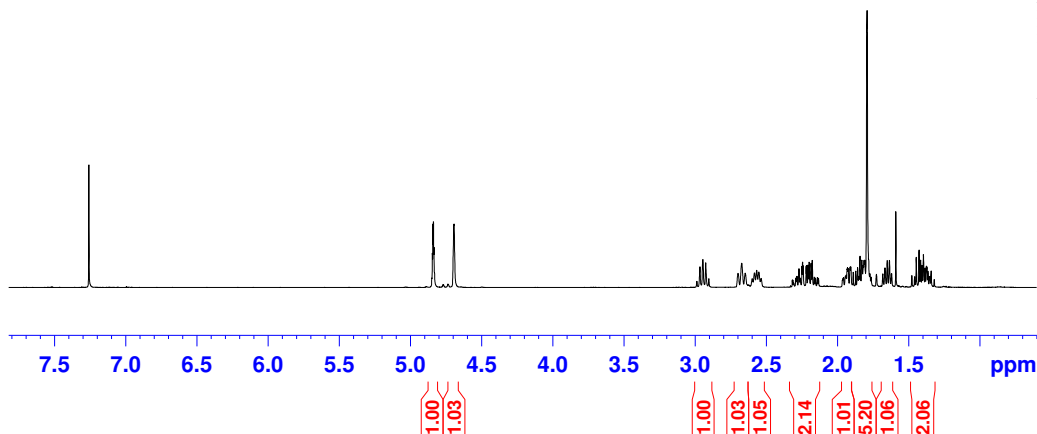
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -1.80 dB
PL12 14.70 dB
PL2W 15.28361320 W
PL12W 0.34215751 W
SFO2 400.2716011 MHz
SI 32768
SF 100.6479720 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Compound R-9a

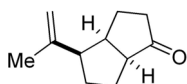


NAME THMA E114
EXPNO 20
PROCNO 1
Date_ 20090430
Time 21.47
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 181
DW 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 -3.00 dB
SFO1 400.1324710 MHz
SI 32768
SF 400.1300094 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



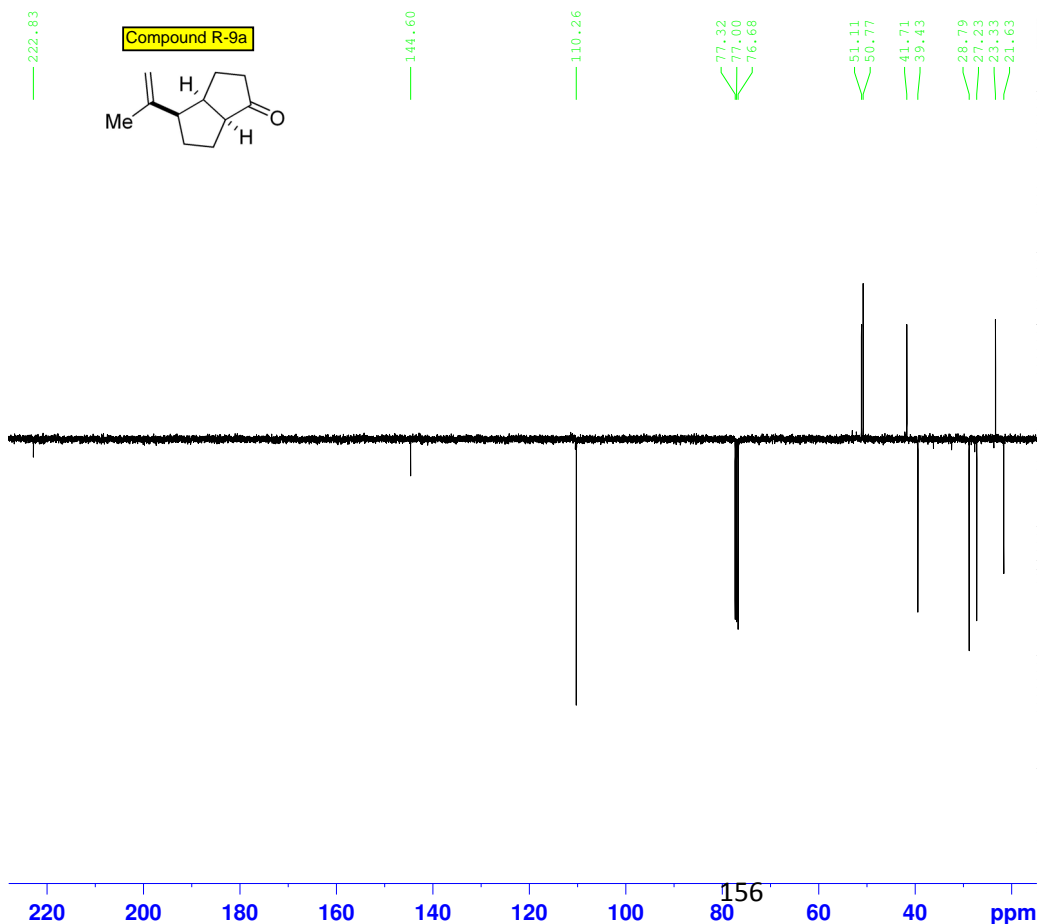
Compound R-9a



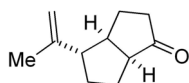
NAME THMA E114
EXPNO 21
PROCNO 1
Date_ 20090430
Time 22.44
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 1000
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3074932 sec
RG 8192
DW 19.950 usec
DE 6.00 usec
TE 300.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
d20 0.00689655 sec
DELTA 0.00001311 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.30 usec
p2 20.60 usec
PL1 -1.00 dB
SFO1 100.6233329 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -3.00 dB
PL12 16.00 dB
SFO2 400.1316005 MHz
SI 32768
SF 100.6127702 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.80

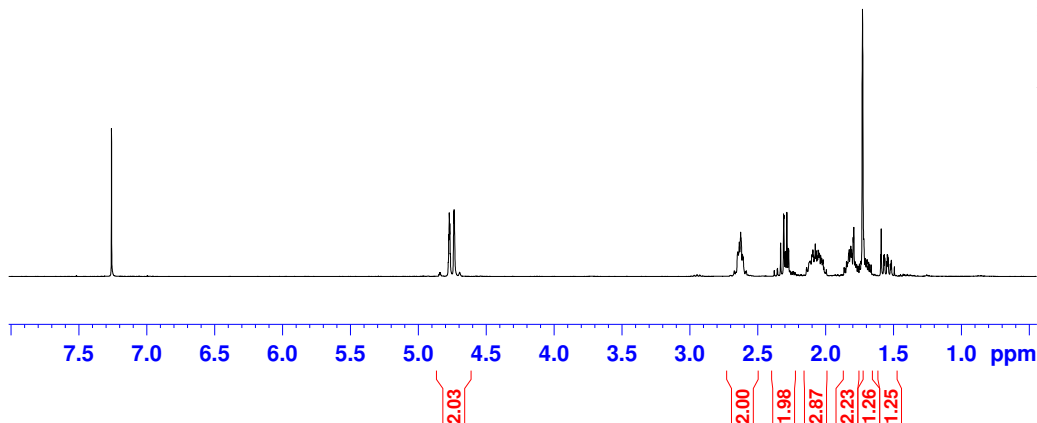


Compound S-9a

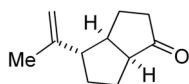


NAME THMA E114
EXPNO 30
PROCNO 1
Date_ 20090501
Time 0.44
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 203.2
DW 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 -3.00 dB
SFO1 400.1324710 MHz
SI 32768
SF 400.1300094 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



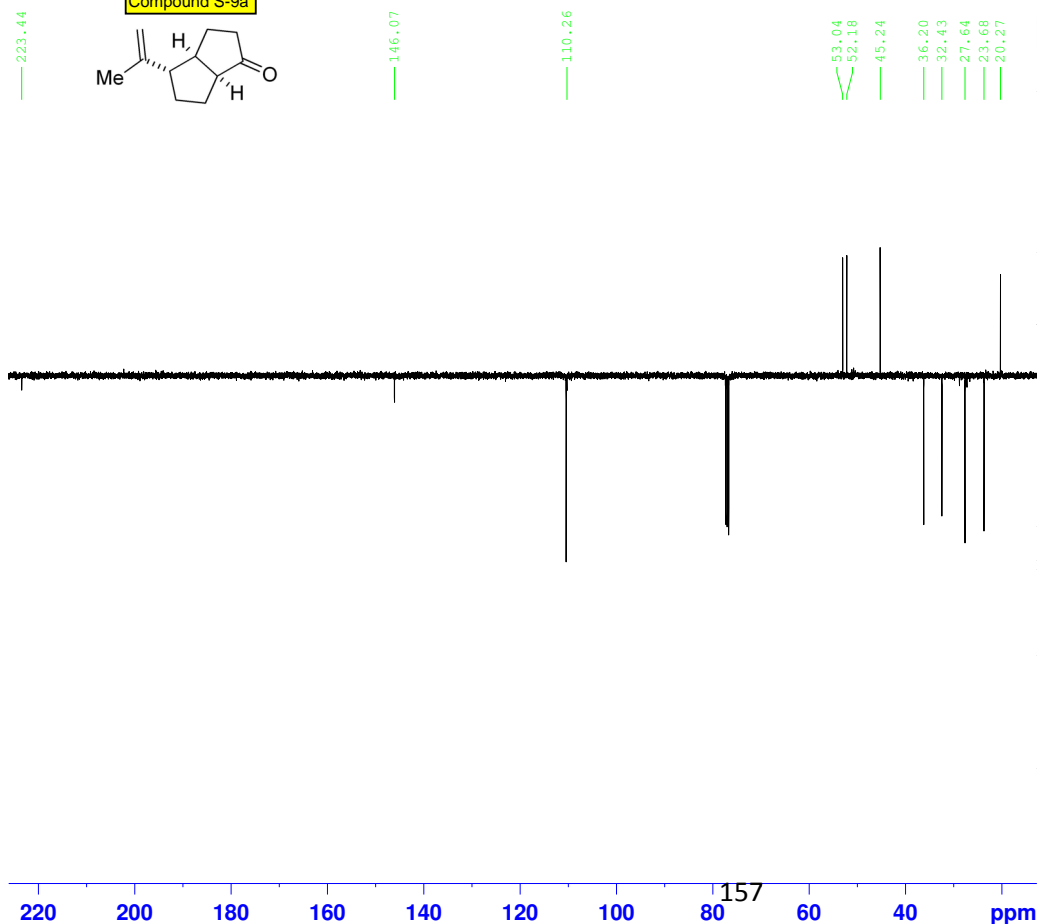
Compound S-9a



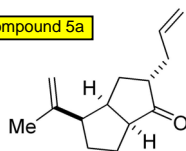
NAME THMA E114
EXPNO 31
PROCNO 1
Date_ 20090501
Time 1.41
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 1000
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3074932 sec
RG 14596.5
DW 19.950 usec
DE 6.00 usec
TE 300.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
d20 0.00689655 sec
DELTA 0.00001311 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.30 usec
p2 20.60 usec
PL1 -1.00 dB
SFO1 100.6233329 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -3.00 dB
PL12 16.00 dB
SFO2 400.1316005 MHz
SI 32768
SF 100.6127701 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

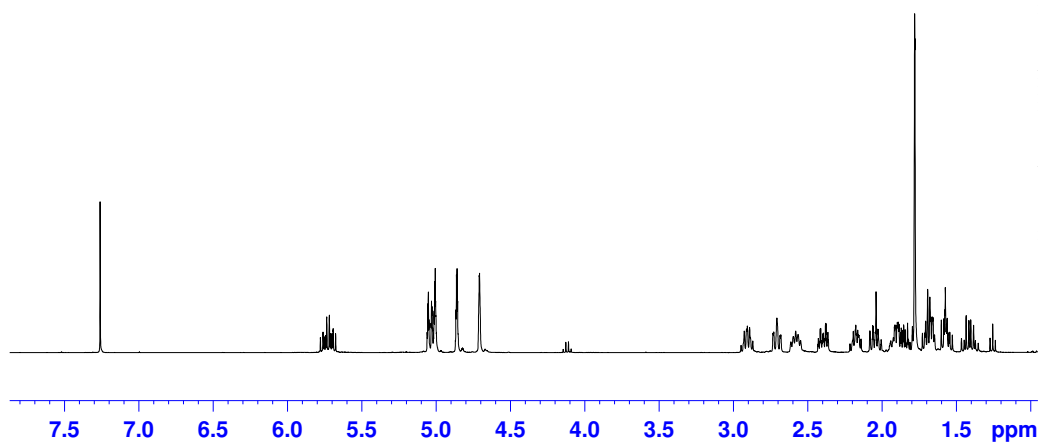


Compound 5a



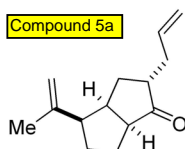
NAME KTE 10F3
EXPNO 80
PROCNO 1
Date_ 20090418
Time 5.22
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 181
DW 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 -3.00 dB
SFO1 400.1324710 MHz
SI 32768
SF 400.1300094 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



223.92

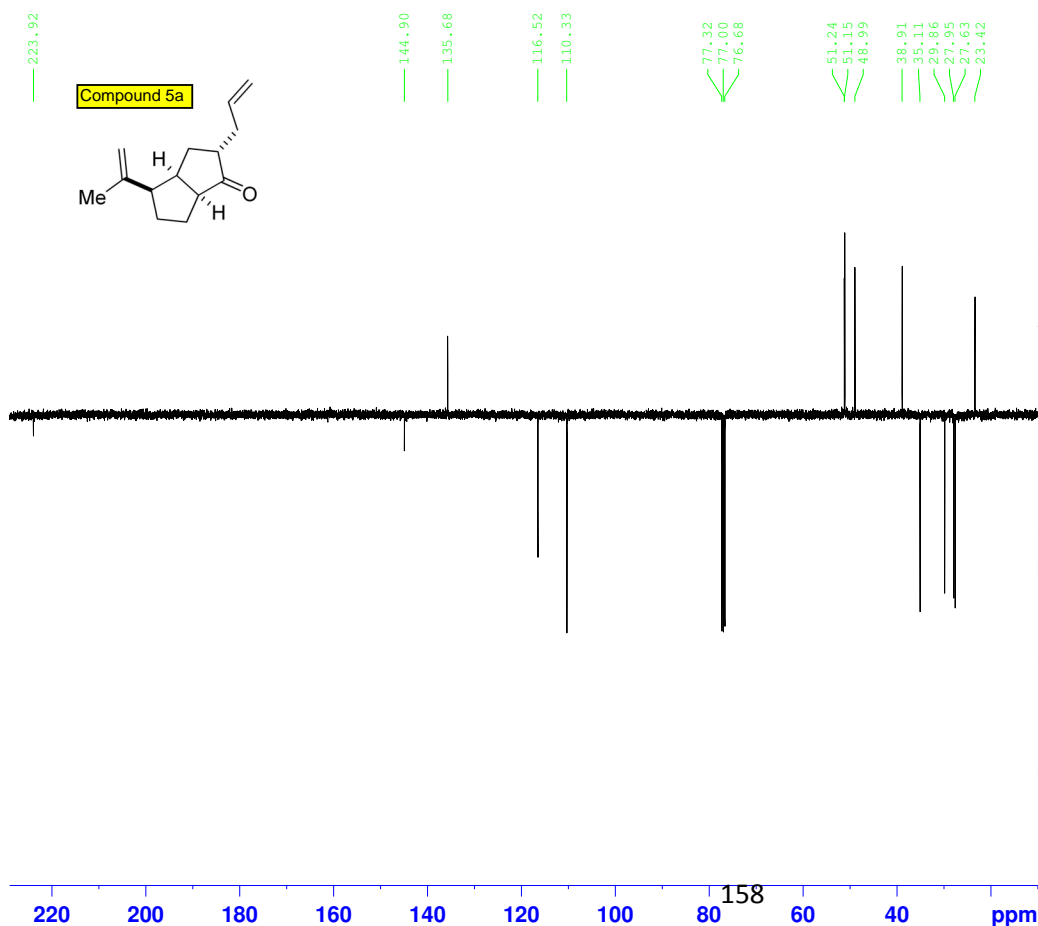
Compound 5a



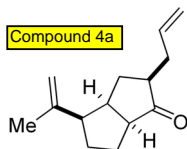
NAME KTE 10F3
EXPNO 81
PROCNO 1
Date_ 20090418
Time 6.19
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 1000
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3074932 sec
RG 9195.2
DW 19.950 usec
DE 6.00 usec
TE 300.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
d20 0.00689655 sec
DELTA 0.00001311 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.30 usec
p2 20.60 usec
PL1 -1.00 dB
SFO1 100.6233329 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -3.00 dB
PL12 16.00 dB
SFO2 400.1316005 MHz
SI 32768
SF 100.6127698 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

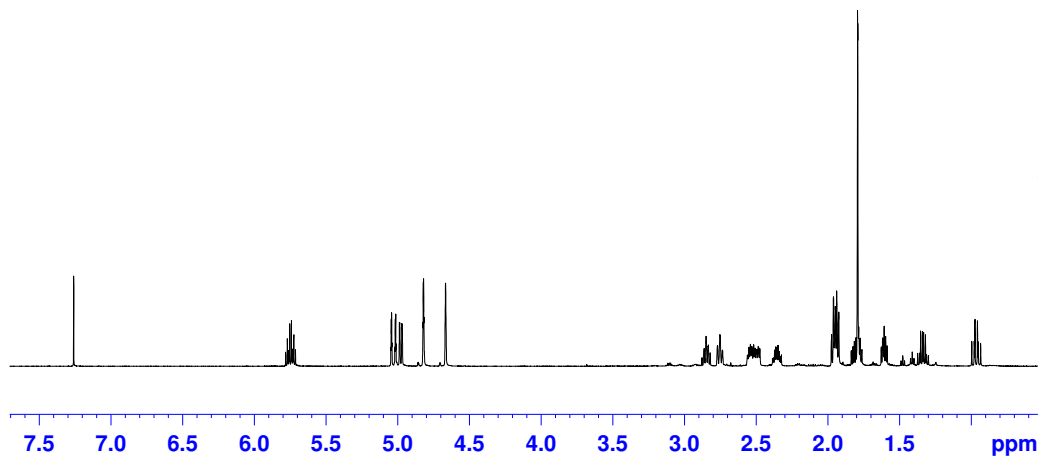


Compound 4a

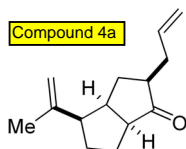


NAME THMA E098
EXPNO 10
PROCNO 1
Date_ 20090423
Time 14.05
INSTRUM advance600
PROBHD 5 mm PAQNP Swi
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 12376.237 Hz
FIDRES 0.188846 Hz
AQ 2.6477449 sec
RG 181
DW 40.400 usec
DE 6.00 usec
TE 298.1 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.85 usec
PL1 -6.00 dB
SFO1 600.1337060 MHz
SI 32768
SF 600.1300174 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 1.00



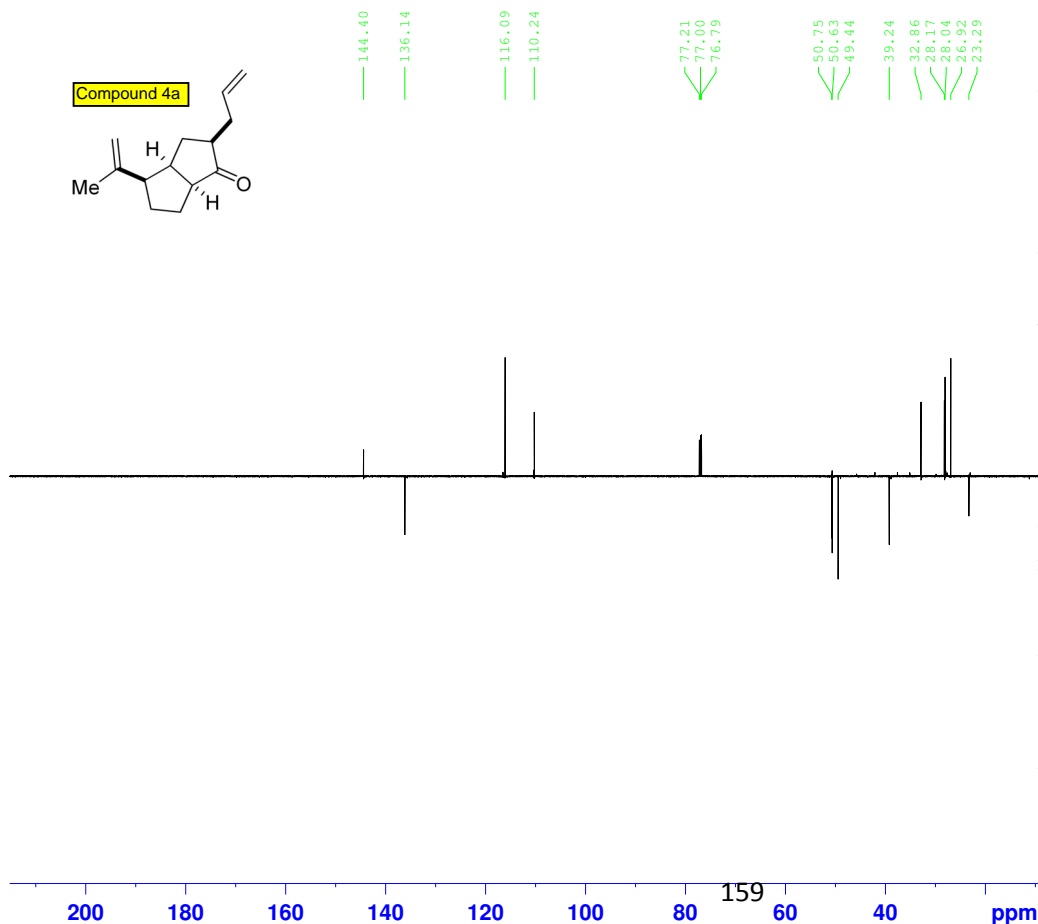
Compound 4a



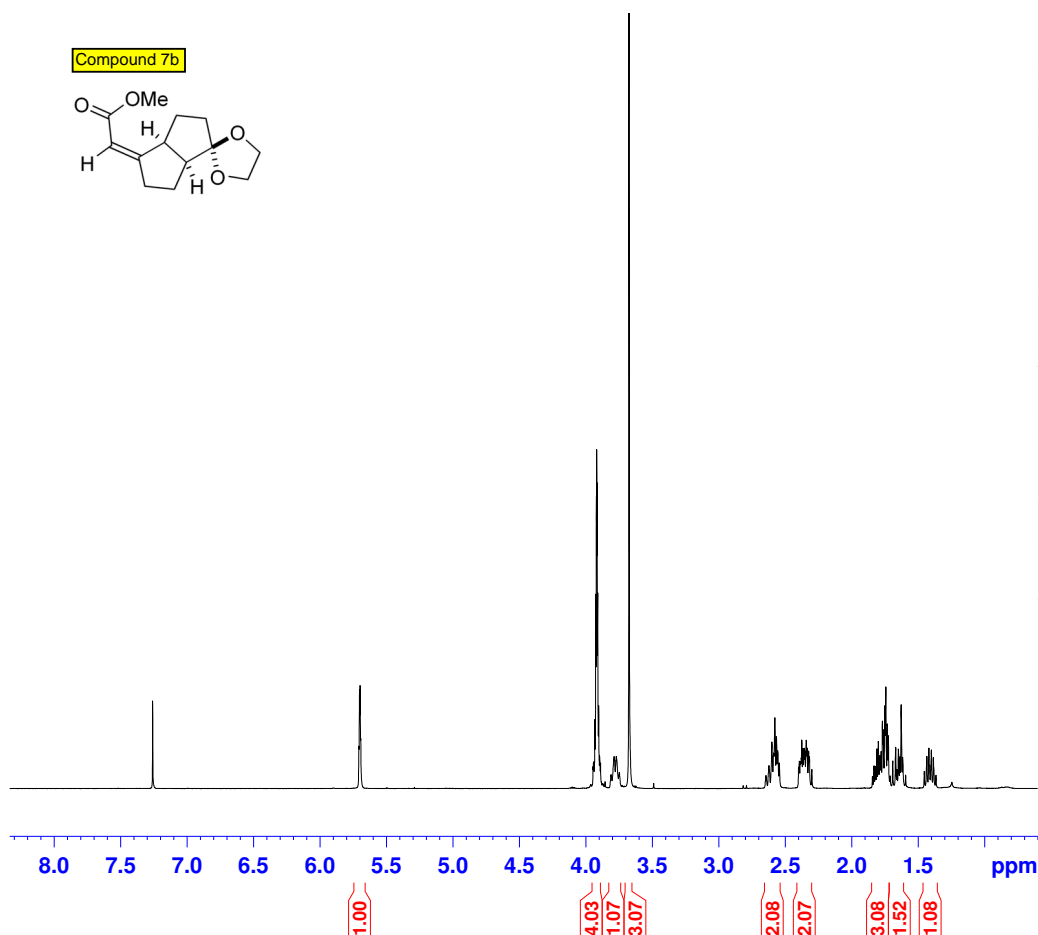
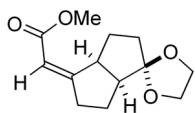
NAME THMA E098
EXPNO 11
PROCNO 1
Date_ 20090423
Time 16.10
INSTRUM advance600
PROBHD 5 mm PAQNP Swi
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 2500
DS 4
SWH 35971.223 Hz
FIDRES 0.548877 Hz
AQ 0.9110143 sec
RG 2298.8
DW 13.900 usec
DE 6.00 usec
TE 298.1 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
d20 0.00689655 sec
DELTA 0.00000828 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 6.50 usec
p2 13.00 usec
PL1 0.00 dB
SFO1 150.9178988 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -6.00 dB
PL12 12.20 dB
SFO2 600.1324005 MHz
SI 32768
SF 150.9028143 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 6.00



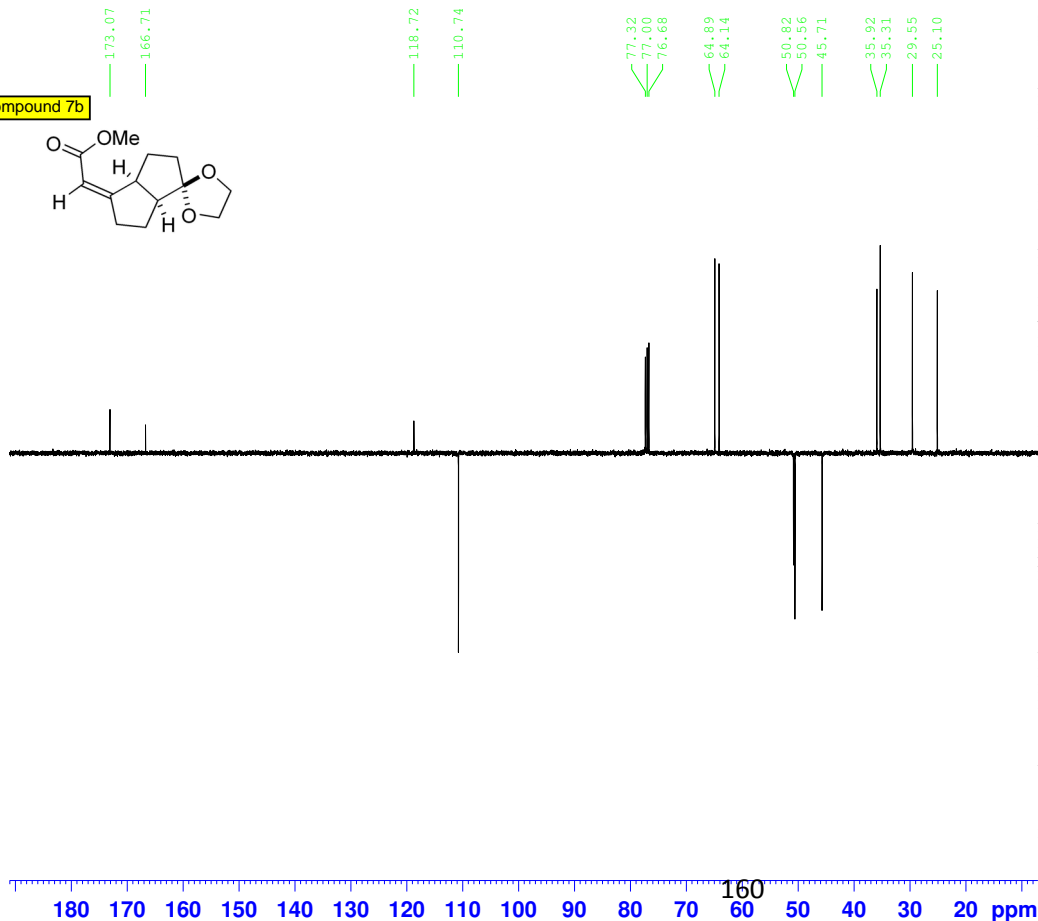
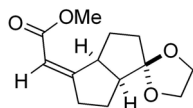
Compound 7b



NAME THMA E236-F1
EXPNO 20
PROCNO 1
Date_ 20090717
Time 11.06
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 143.7
DW 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 -3.00 dB
SFO1 400.1324710 MHz
SI 32768
SF 400.1300092 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

Compound 7b

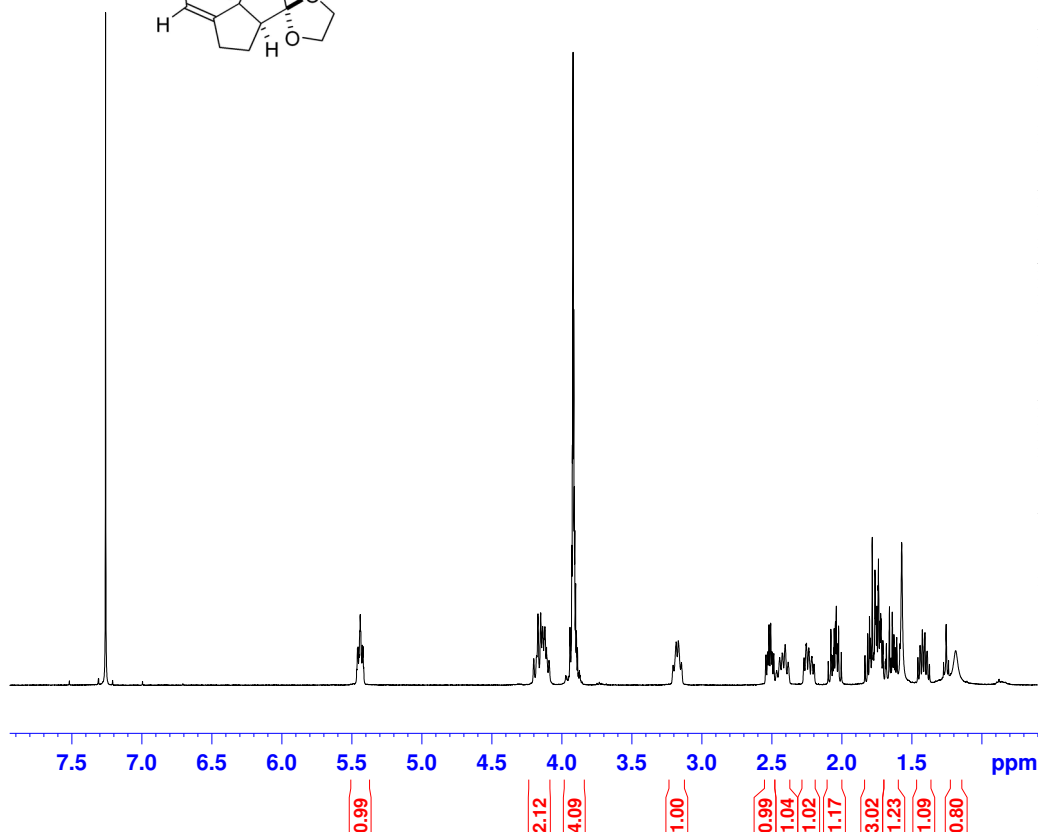
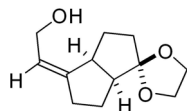


NAME THMA E236-F1
EXPNO 21
PROCNO 1
Date_ 20090717
Time 11.58
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 931
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3074932 sec
RG 9195.2
DW 19.950 usec
DE 6.00 usec
TE 300.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
d20 0.00689655 sec
DELTA 0.00001311 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.30 usec
p2 20.60 usec
PL1 -1.00 dB
SFO1 100.6233329 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -3.00 dB
PL12 16.00 dB
SFO2 400.1316005 MHz
SI 32768
SF 100.6127702 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 3.00

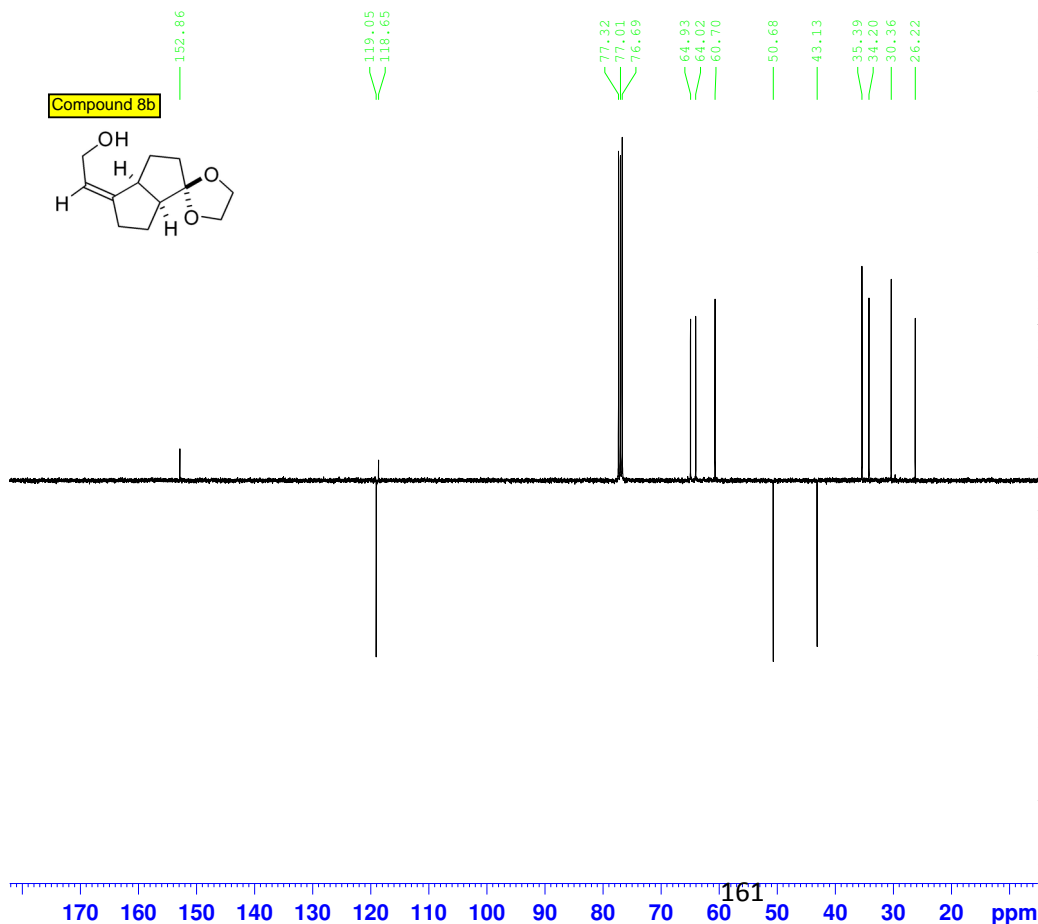
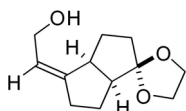
Compound 8b



NAME THMA E263
EXPNO 270
PROCNO 1
Date_ 20090717
Time 14.33
INSTRUM AVIII400
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 575
DW 60.800 usec
DE 6.50 usec
TE 298.2 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.50 usec
PL1 -1.80 dB
PL1W 15.28361320 W
SFO1 400.2724718 MHz
SI 32768
SF 400.2700105 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

Compound 8b

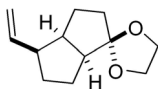


NAME THMA E263
EXPNO 271
PROCNO 1
Date_ 20090718
Time 2.14
INSTRUM AVIII400
PROBHD 5 mm PABBO BB-
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 6000
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 298.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
D20 0.00689655 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
P2 20.00 usec
PL1 -1.50 dB
PL1W 47.89980698 W
SFO1 100.6580364 MHz

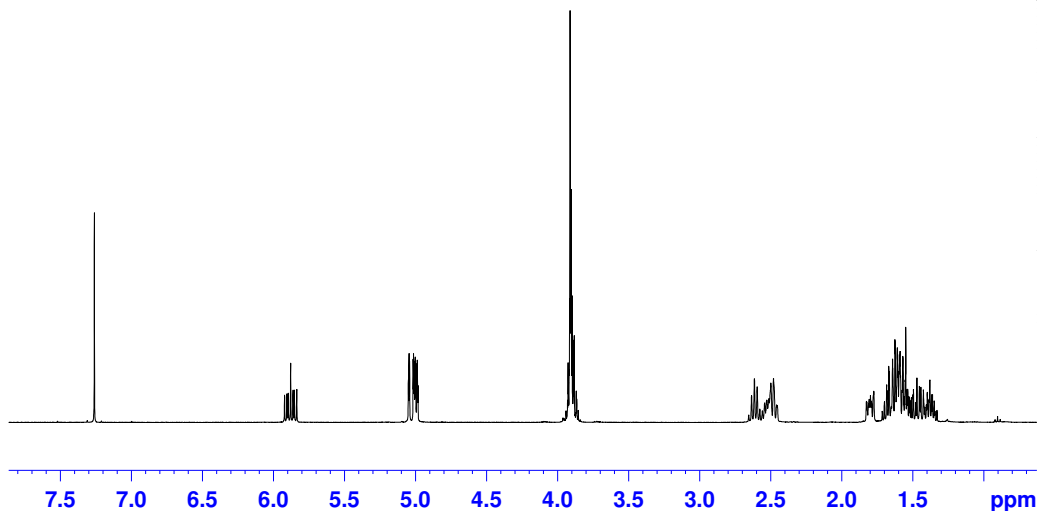
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -1.80 dB
PL12 14.70 dB
PL2W 15.28361320 W
PL12W 0.34215751 W
SFO2 400.2716011 MHz
SI 32768
SF 100.6479720 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

Compound S-16



NAME THMA E242AB
EXPNO 210
PROCNO 1
Date_ 20090702
Time 15.22
INSTRUM AVIII400
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 406
DW 60.800 usec
DE 6.50 usec
TE 298.2 K
D1 1.00000000 sec
TD0 1

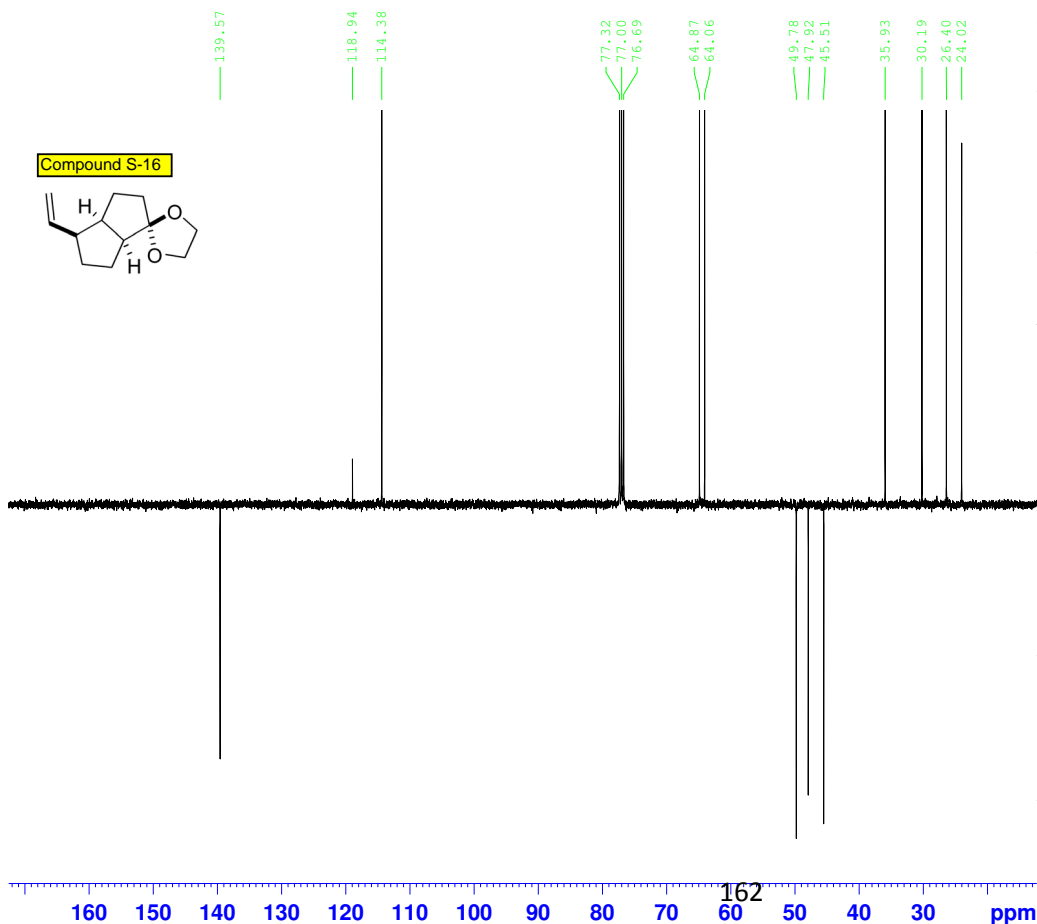
===== CHANNEL f1 =====
NUC1 1H
P1 13.50 usec
PL1 -1.80 dB
PL1W 15.28361320 W
SFO1 400.2724718 MHz
SI 32768
SF 400.2700105 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



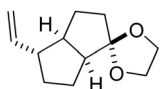
NAME THMA E242AB
EXPNO 211
PROCNO 1
Date_ 20090703
Time 0.01
INSTRUM AVIII400
PROBHD 5 mm PABBO BB-
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 3000
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 298.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
D20 0.00689655 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
P2 20.00 usec
PL1 -1.50 dB
PL1W 47.89980698 W
SFO1 100.6580364 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -1.80 dB
PL12 14.70 dB
PL2W 15.28361320 W
PL12W 0.34215751 W
SFO2 400.2716011 MHz
SI 32768
SF 100.6479720 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



Compound R-16



```

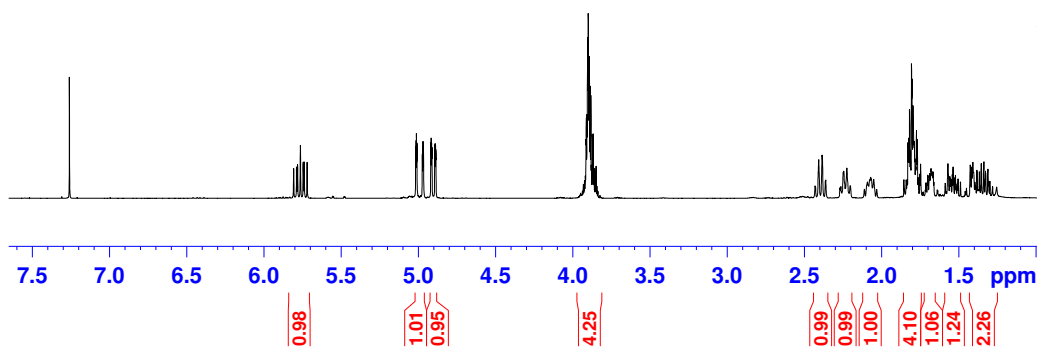
NAME      THMA E242AB
EXPNO     220
PROCNO    1
Date_     20090702
Time      15.27
INSTRUM   AVIII400
PROBHD    5 mm PABBO BB-
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        8223.685 Hz
FIDRES     0.125483 Hz
AQ         3.9846387 sec
RG         181
DW         60.800 usec
DE         6.50 usec
TE         298.2 K
D1         1.00000000 sec
TD0        1

```

```

===== CHANNEL f1 =====
NUC1      1H
P1        13.50 usec
PL1       -1.80 dB
PL1W      15.28361320 W
SFO1      400.2724718 MHz
SI        32768
SF        400.2700107 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00

```



```

NAME      THMA E242AB
EXPNO     222
PROCNO    1
Date_     20090703
Time      2.55
INSTRUM   AVIII400
PROBHD    5 mm PABBO BB-
PULPROG   jmod
TD         65536
SOLVENT   CDCl3
NS         3000
DS         4
SWH        24038.461 Hz
FIDRES     0.366798 Hz
AQ         1.3631988 sec
RG         2050
DW         20.800 usec
DE         6.50 usec
TE         298.2 K
CNST2     145.0000000
CNST11    1.0000000
D1         2.00000000 sec
D20       0.00689655 sec
TD0        1

```

```

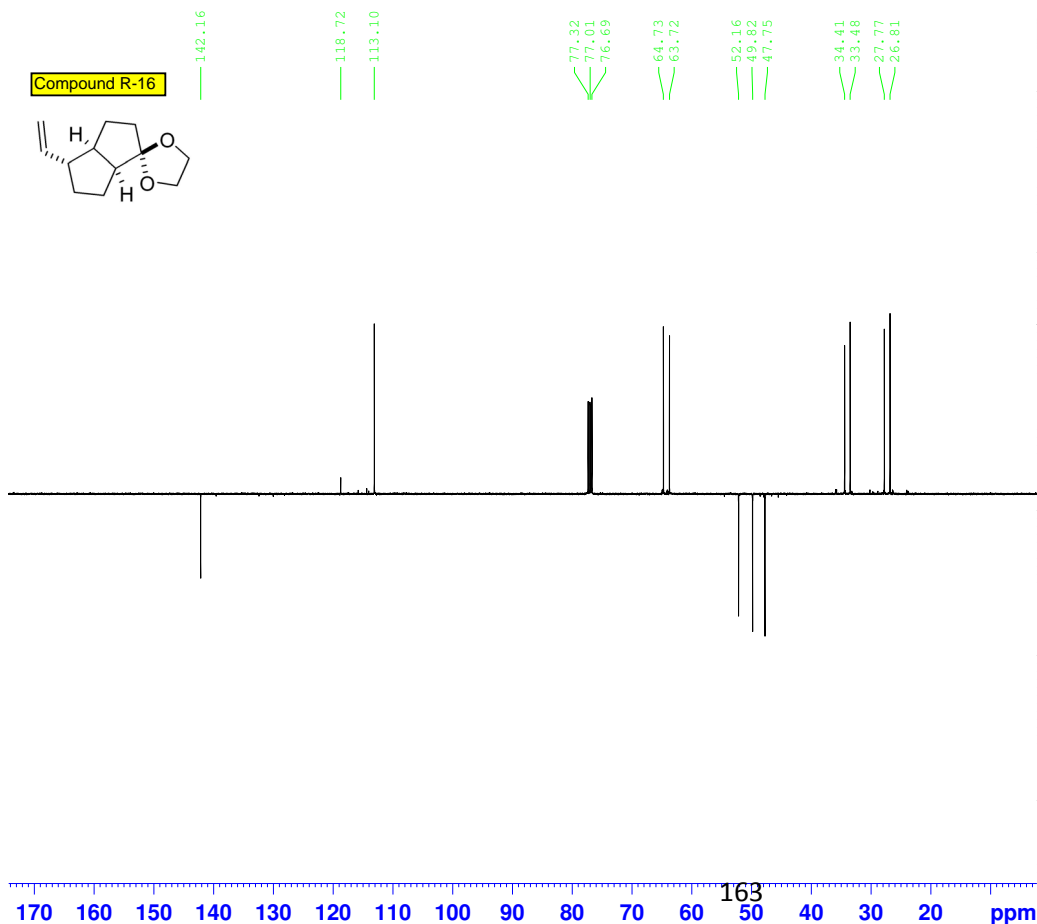
===== CHANNEL f1 =====
NUC1      13C
P1        10.00 usec
P2        20.00 usec
PL1       -1.50 dB
PL1W      47.89980698 W
SFO1      100.6580364 MHz

```

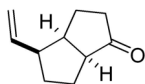
```

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2       1H
PCPD2     100.00 usec
PL2       -1.80 dB
PL12      14.70 dB
PL2W      15.28361320 W
PL12W     0.34215751 W
SFO2      400.2716011 MHz
SI        32768
SF        100.6479720 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        5.00

```

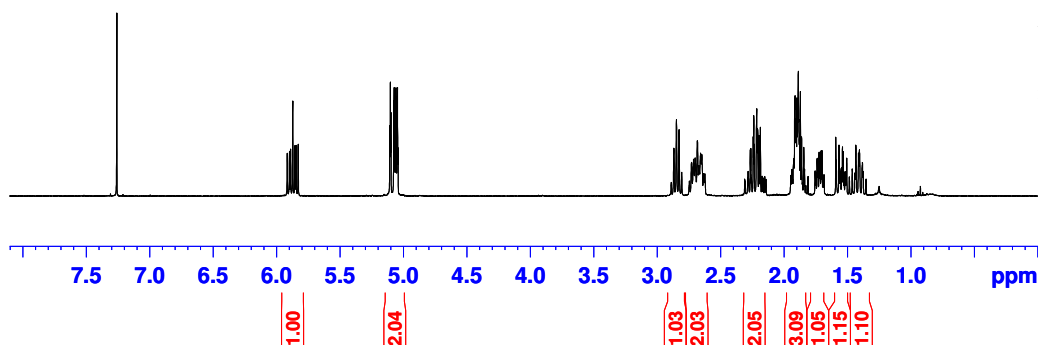


Compound 9b

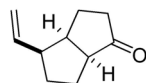


NAME THMA E243
EXPNO 280
PROCNO 1
Date_ 20090702
Time 19.06
INSTRUM AVIII400
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 1
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 322
DW 60.800 usec
DE 6.50 usec
TE 298.3 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.50 usec
PL1 -1.80 dB
PL1W 15.28361320 W
SFO1 400.2724718 MHz
SI 32768
SF 400.2700107 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



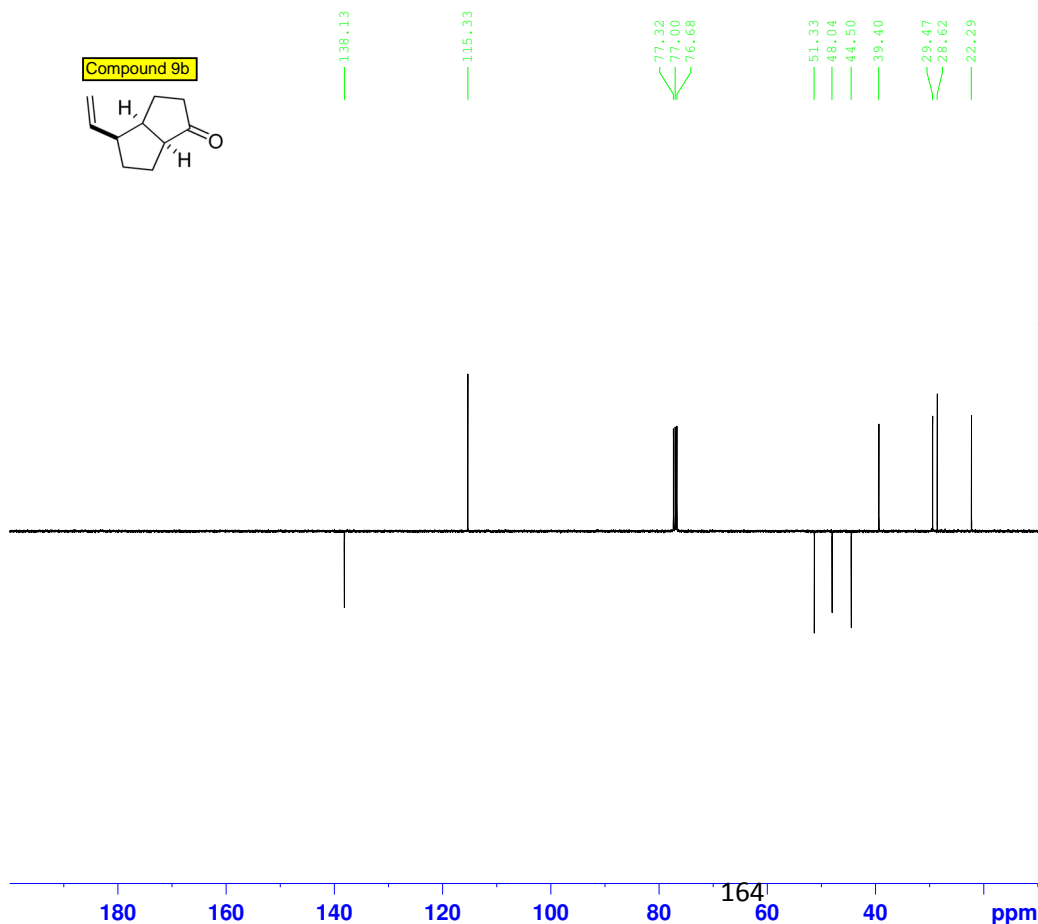
Compound 9b



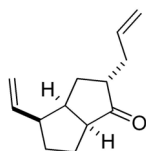
NAME THMA E243
EXPNO 281
PROCNO 1
Date_ 20090703
Time 4.52
INSTRUM AVIII400
PROBHD 5 mm PABBO BB-
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 2000
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 298.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
D20 0.00689655 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
P2 20.00 usec
PL1 -1.50 dB
PL1W 47.89980698 W
SFO1 100.6580364 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -1.80 dB
PL12 14.70 dB
PL2W 15.28361320 W
PL12W 0.34215751 W
SFO2 400.2716011 MHz
SI 32768
SF 100.6479732 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 4.00



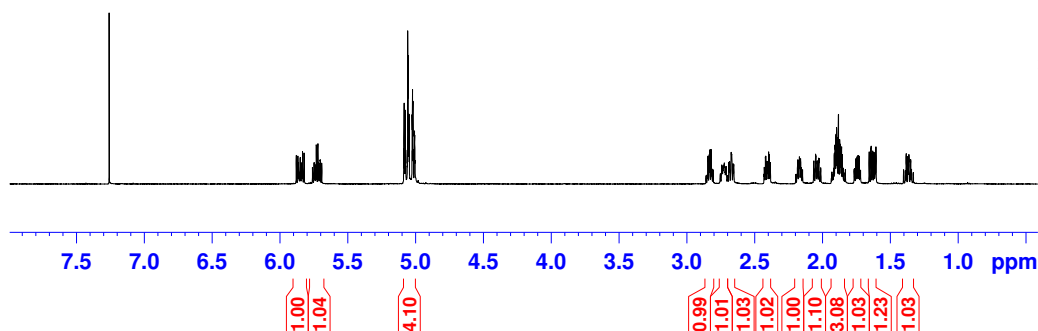
Compound 5b



```

NAME      THMA E152
EXPNO     20
PROCNO    1
Date_     20090519
Time      1.34
INSTRUM   avance600
PROBHD    5 mm PAQNP Swi
PULPROG   zg30
TD        65536
SOLVENT   CDCl3
NS        32
DS        2
SWH       12376.237 Hz
FIDRES    0.188846 Hz
AQ        2.6477449 sec
RG        181
DW        40.400 usec
DE        6.00 usec
TE        298.1 K
D1        1.00000000 sec
TD0       1

```

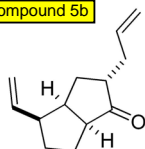


```

===== CHANNEL f1 =====
NUC1      1H
P1        13.85 usec
PL1       -6.00 dB
SFO1      600.1337060 MHz
SI        32768
SF        600.1300174 MHz
WDW       no
SSB       0
LB        0.00 Hz
GB        0
PC        1.00

```

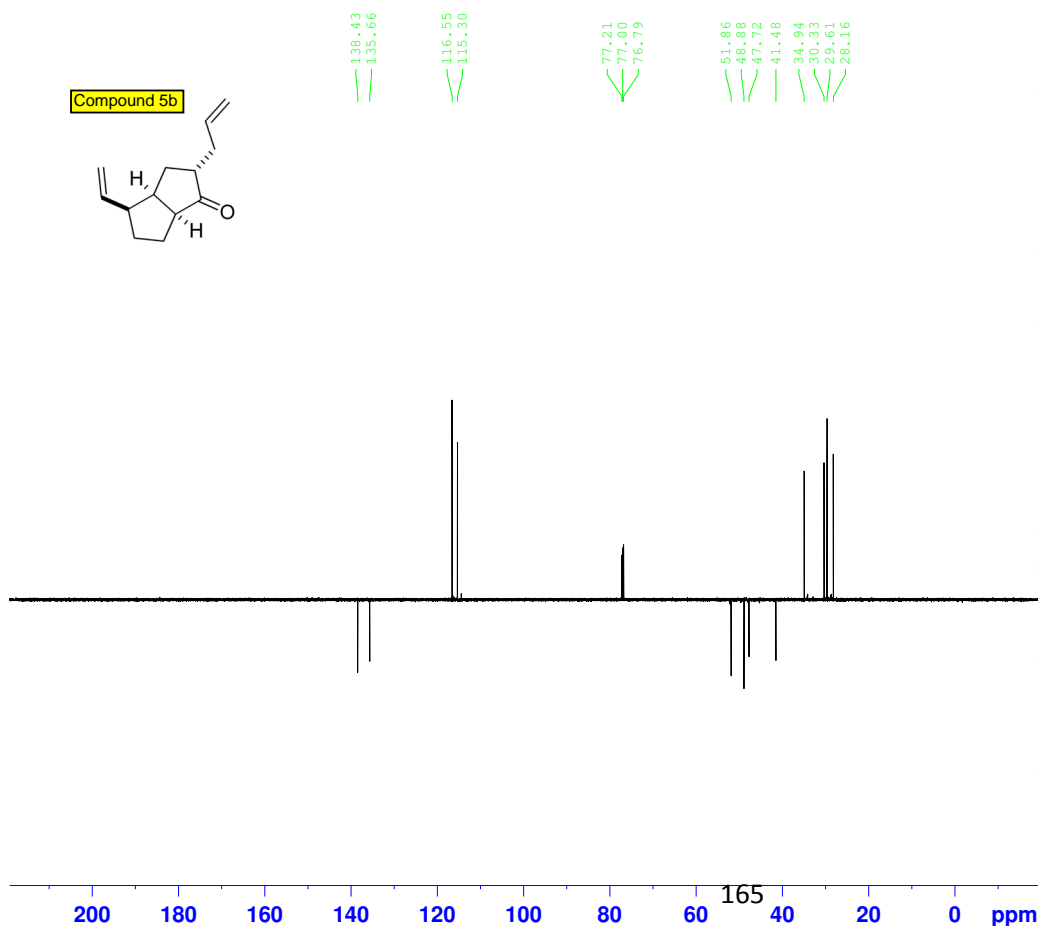
Compound 5b



```

NAME      THMA E152
EXPNO     25
PROCNO    1
Date_     20090519
Time      4.35
INSTRUM   avance600
PROBHD    5 mm PAQNP Swi
PULPROG   jmod
TD        65536
SOLVENT   CDCl3
NS        1200
DS        4
SWH       35971.223 Hz
FIDRES    0.548877 Hz
AQ        0.9110143 sec
RG        2048
DW        13.900 usec
DE        6.00 usec
TE        298.1 K
CNST2     145.0000000
CNST11    1.0000000
D1        2.00000000 sec
d20       0.00689655 sec
DELTA     0.00000828 sec
TD0       1

```



```

===== CHANNEL f1 =====
NUC1      13C
P1        6.50 usec
p2        13.00 usec
PL1       0.00 dB
SFO1      150.9178988 MHz

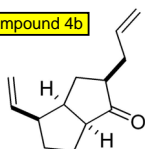
```

```

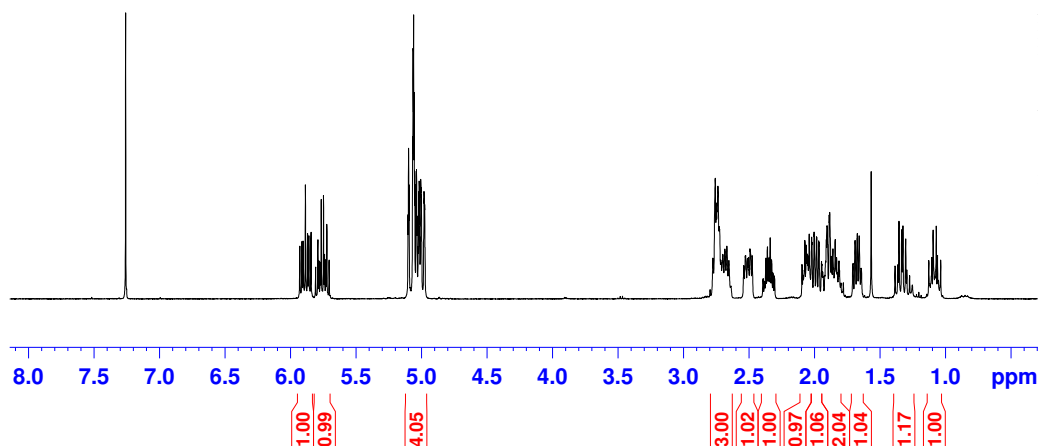
===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2      1H
PCPD2     80.00 usec
PL2       -6.00 dB
PL12      12.20 dB
SFO2      600.1324005 MHz
SI        32768
SF        150.9028144 MHz
WDW       no
SSB       0
LB        0.00 Hz
GB        0
PC        3.00

```

Compound 4b

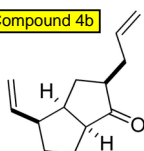


NAME THMA E245-F1
EXPNO 20
PROCNO 1
Date_ 20090710
Time 12.59
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 287.4
DW 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.00000000 sec
TD0 1

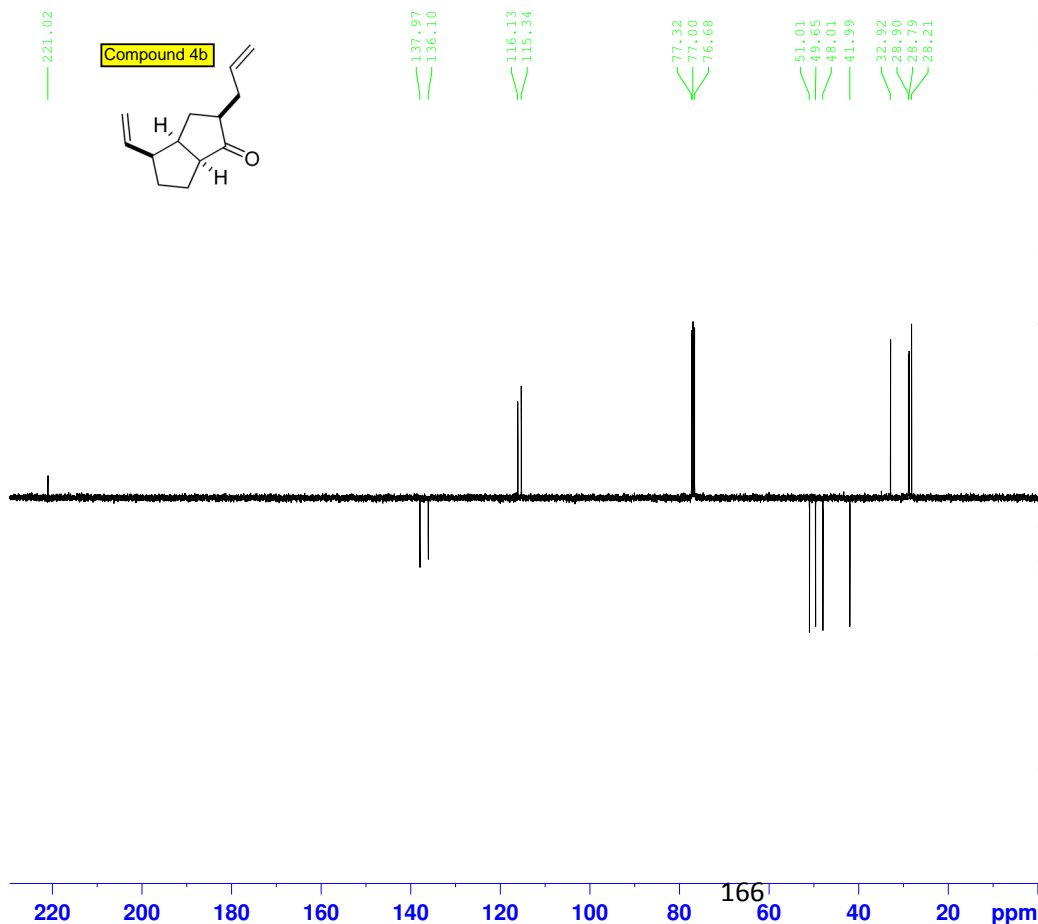


===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 -3.00 dB
SFO1 400.1324710 MHz
SI 32768
SF 400.1300093 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

Compound 4b



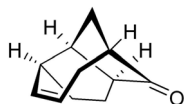
NAME THMA E245-F1
EXPNO 21
PROCNO 1
Date_ 20090710
Time 13.02
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 1000
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3074932 sec
RG 8192
DW 19.950 usec
DE 6.00 usec
TE 300.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
d20 0.00689655 sec
DELTA 0.00001311 sec
TD0 1



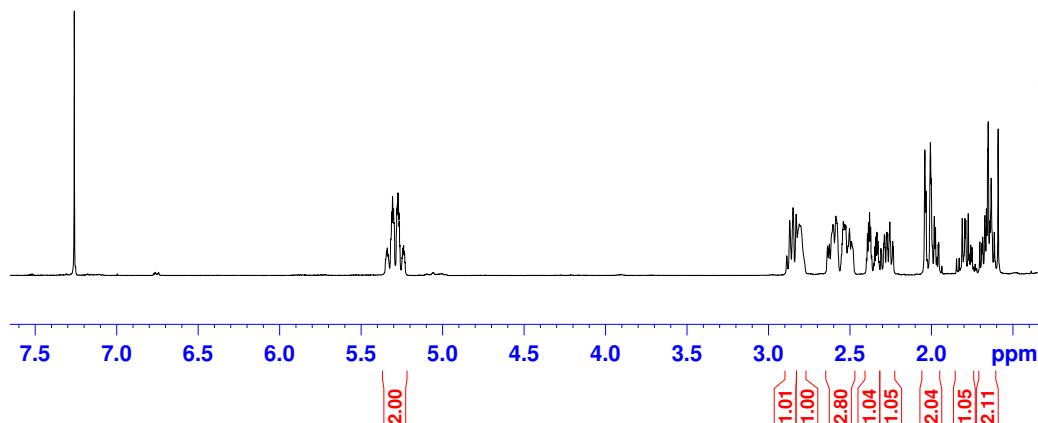
===== CHANNEL f1 =====
NUC1 13C
P1 10.30 usec
p2 20.60 usec
PL1 -1.00 dB
SFO1 100.6233329 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -3.00 dB
PL12 16.00 dB
SFO2 400.1316005 MHz
SI 32768
SF 100.6127699 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 3.00

Compound 3b

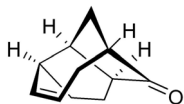


NAME THMA E163
EXPNO 20
PROCNO 1
Date_ 20090527
Time 10.24
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 203.2
DW 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.00000000 sec
TD0 1

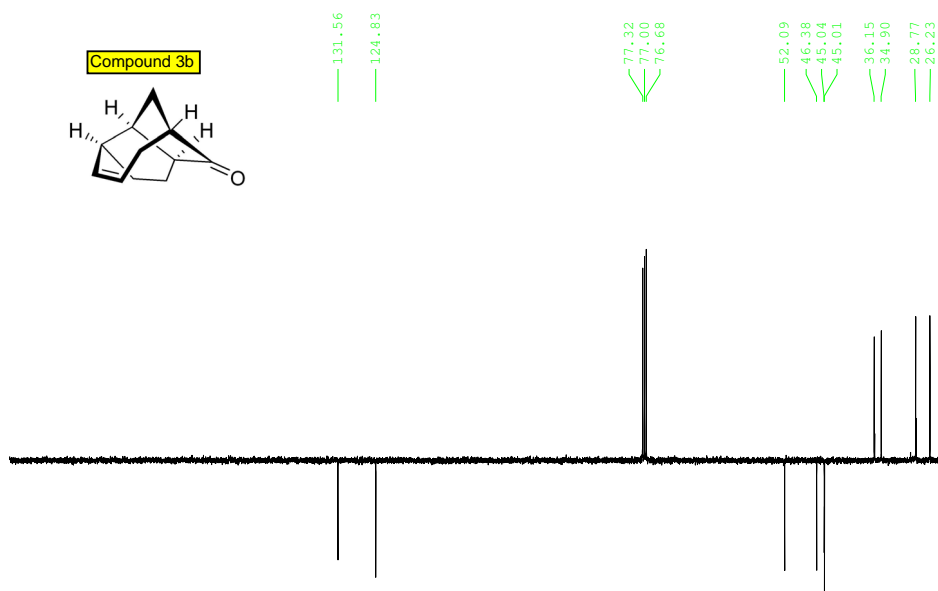


===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 -3.00 dB
SFO1 400.1324710 MHz
SI 32768
SF 400.1300093 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

Compound 3b

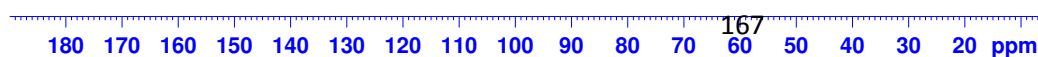


NAME THMA E163
EXPNO 21
PROCNO 1
Date_ 20090527
Time 10.43
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 1600
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3074932 sec
RG 9195.2
DW 19.950 usec
DE 6.00 usec
TE 300.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
d20 0.00689655 sec
DELTA 0.00001311 sec
TD0 1

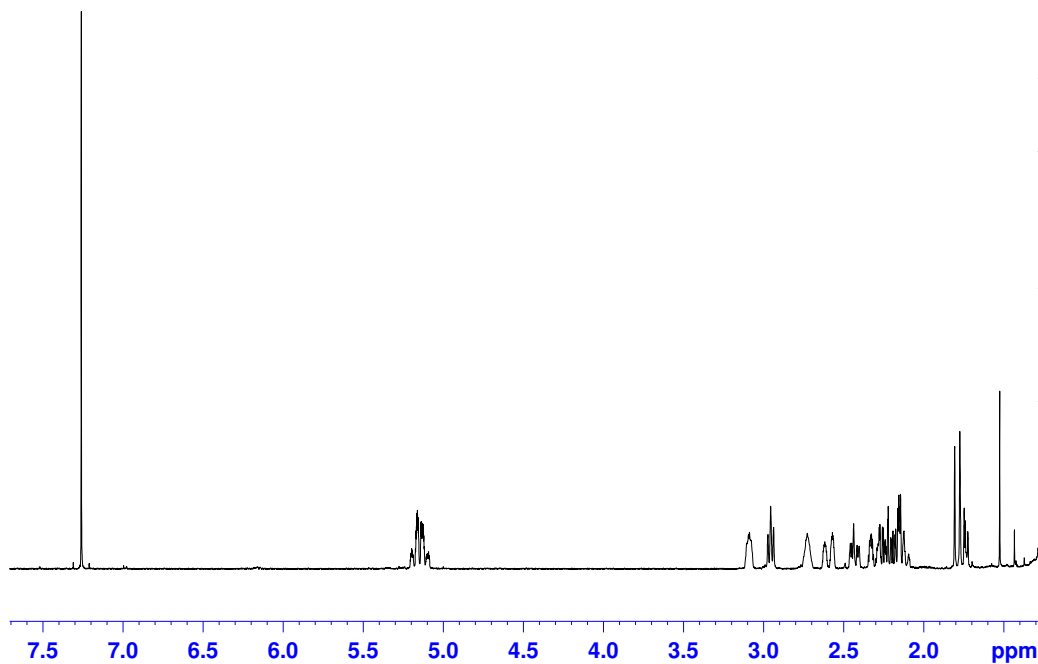
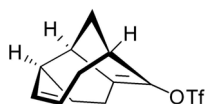


===== CHANNEL f1 =====
NUC1 13C
P1 10.30 usec
p2 20.60 usec
PL1 -1.00 dB
SFO1 100.6233329 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -3.00 dB
PL12 16.00 dB
SFO2 400.1316005 MHz
SI 32768
SF 100.6127707 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 8.00



Compound 17



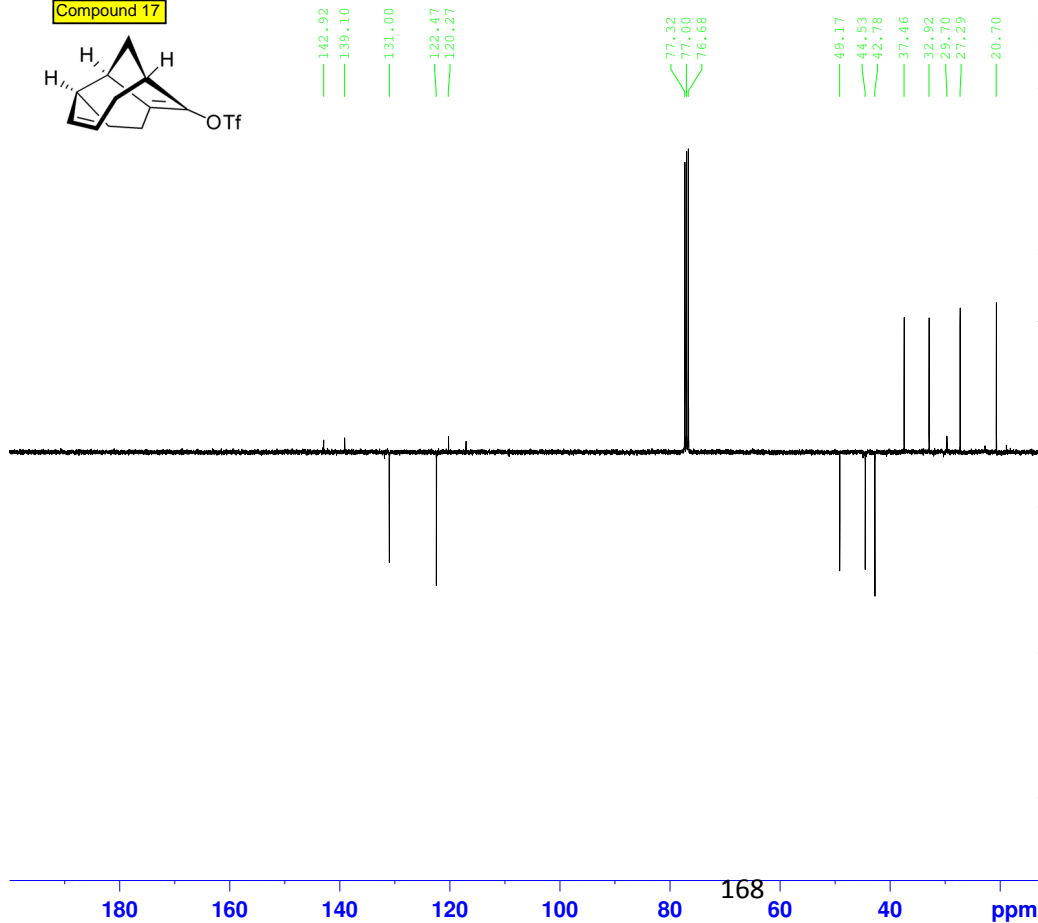
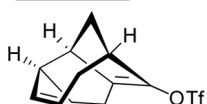
```

NAME      THMA E251
EXPNO     220
PROCNO    1
Date_     20090707
Time      14.57
INSTRUM   AVIII400
PROBHD    5 mm PABBO BB-
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         1
DS         2
SWH        8223.685 Hz
FIDRES     0.125483 Hz
AQ         3.9846387 sec
RG         575
DW         60.800 usec
DE         6.50 usec
TE         298.2 K
D1         1.00000000 sec
TD0        1
  
```

```

===== CHANNEL f1 =====
NUC1       1H
P1         13.50 usec
PL1        -1.80 dB
PL1W       15.28361320 W
SFO1       400.2724718 MHz
SI         32768
SF         400.2700105 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
  
```

Compound 17



```

NAME      THMA E251
EXPNO     221
PROCNO    1
Date_     20090708
Time      4.11
INSTRUM   AVIII400
PROBHD    5 mm PABBO BB-
PULPROG   jmod
TD         65536
SOLVENT   CDCl3
NS         4000
DS         4
SWH        24038.461 Hz
FIDRES     0.366798 Hz
AQ         1.3631988 sec
RG         2050
DW         20.800 usec
DE         6.50 usec
TE         298.2 K
CNST2     145.0000000
CNST11    1.0000000
D1         2.00000000 sec
D20        0.00689655 sec
TD0        1
  
```

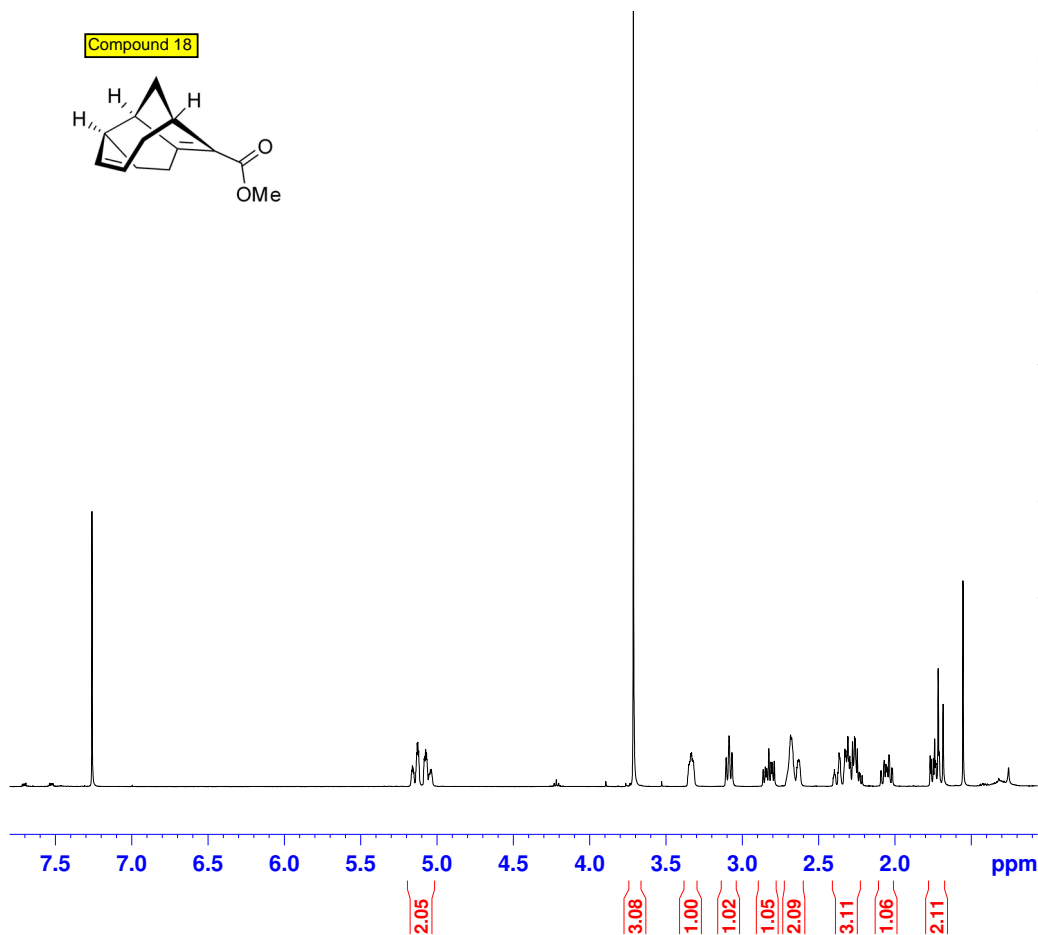
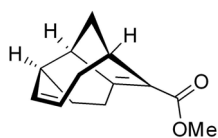
```

===== CHANNEL f1 =====
NUC1       13C
P1         10.00 usec
P2         20.00 usec
PL1        -1.50 dB
PL1W       47.89980698 W
SFO1       100.6580364 MHz
  
```

```

===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2        1H
PCPD2      100.00 usec
PL2        -1.80 dB
PL12       14.70 dB
PL2W       15.28361320 W
PL12W      0.34215751 W
SFO2       400.2716011 MHz
SI         32768
SF         100.6479720 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         3.40
  
```

Compound 18



```

NAME      THMA E175
EXPNO     430
PROCNO    1
Date_     20090530
Time      18.43
INSTRUM   avance400
PROBHD    5 mm BBO BB-1H
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         256
DS         2
SWH        8278.146 Hz
FIDRES     0.126314 Hz
AQ         3.9584243 sec
RG         456.1
DW         60.400 usec
DE         6.00 usec
TE         300.2 K
D1         1.00000000 sec
TD0        1
  
```

```

===== CHANNEL f1 =====
NUC1      1H
P1         8.75 usec
PL1        -3.00 dB
SFO1      400.1324710 MHz
SI         32768
SF         400.1300094 MHz
WDW        EM
SSB         0
LB         0.30 Hz
GB          0
PC         1.00
  
```



```

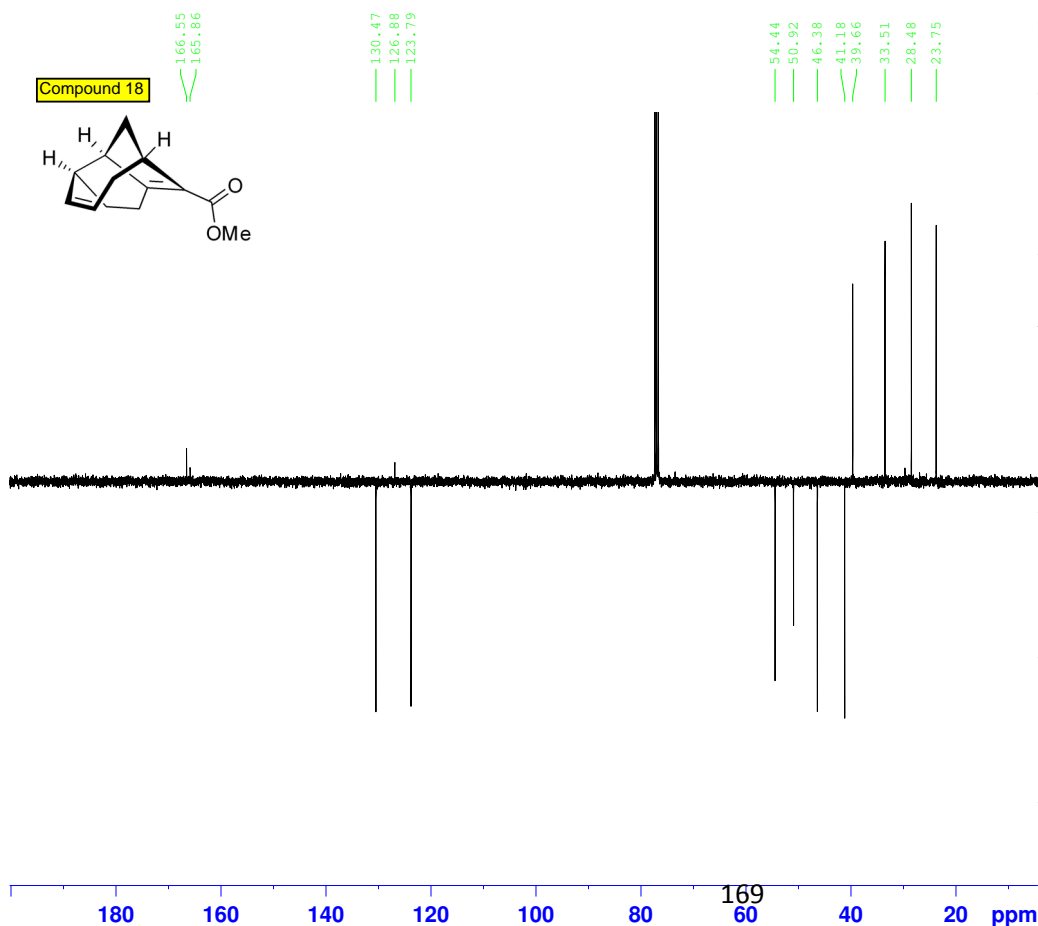
NAME      THMA E252
EXPNO     431
PROCNO    1
Date_     20090708
Time      7.05
INSTRUM   AVIII400
PROBHD    5 mm PABBO BB-
PULPROG   jmod
TD         65536
SOLVENT   CDCl3
NS         3000
DS         4
SWH        24038.461 Hz
FIDRES     0.366798 Hz
AQ         1.3631988 sec
RG         2050
DW         20.800 usec
DE         6.50 usec
TE         298.2 K
CNST2     145.0000000
CNST11    1.0000000
D1         2.00000000 sec
D20        0.00689655 sec
TD0        1
  
```

```

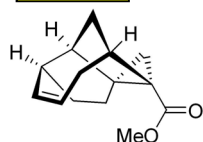
===== CHANNEL f1 =====
NUC1      13C
P1         10.00 usec
P2         20.00 usec
PL1        -1.50 dB
PL1W       47.89980698 W
SFO1      100.6580364 MHz
  
```

```

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2       1H
PCPD2     100.00 usec
PL2        -1.80 dB
PL12       14.70 dB
PL2W       15.28361320 W
PL12W      0.34215751 W
SFO2      400.2716011 MHz
SI         32768
SF         100.6479720 MHz
WDW        EM
SSB         0
LB         1.00 Hz
GB          0
PC         1.60
  
```

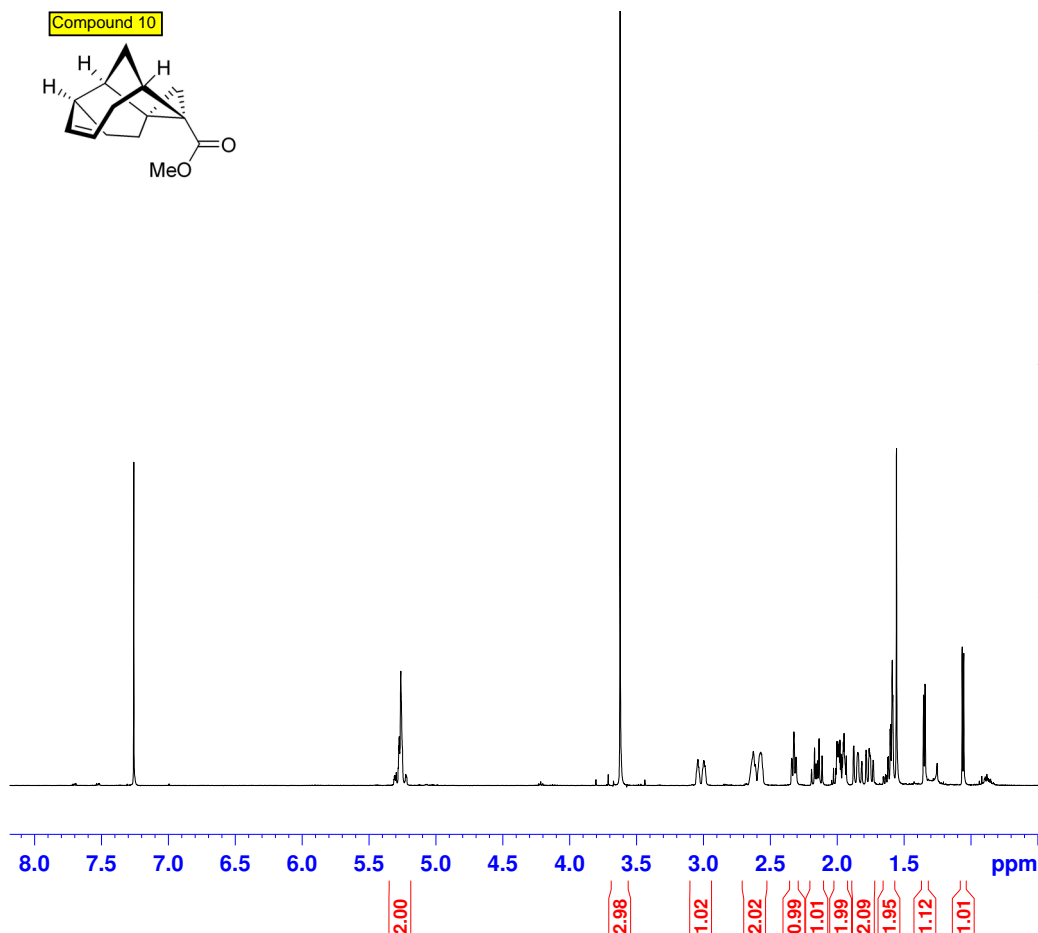


Compound 10



NAME THMA EK784HF
EXPNO 20
PROCNO 1
Date_ 20090603
Time 14.01
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 287.4
DW 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.00000000 sec
TD0 1

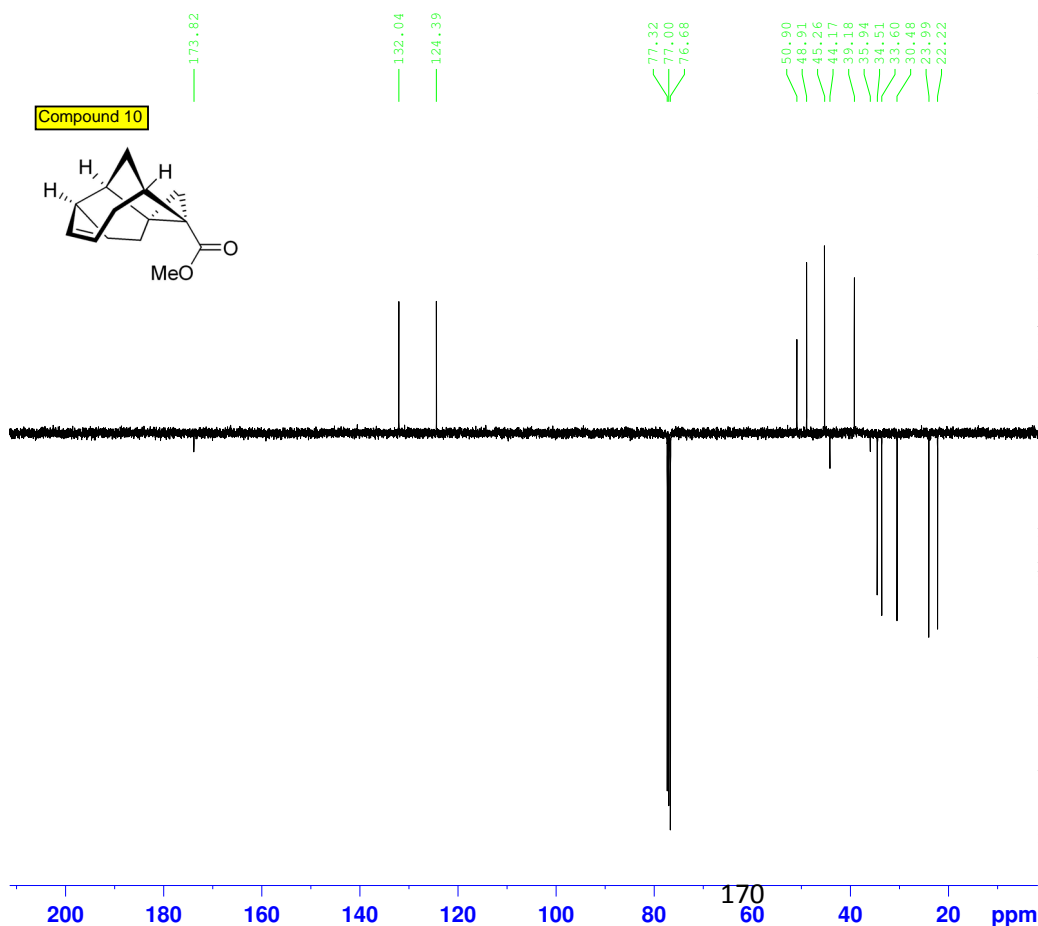
===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 -3.00 dB
SFO1 400.1324710 MHz
SI 32768
SF 400.1300095 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



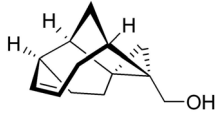
NAME THMA EK784HF
EXPNO 21
PROCNO 1
Date_ 20090603
Time 15.09
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 2500
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3074932 sec
RG 5160.6
DW 19.950 usec
DE 6.00 usec
TE 300.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
d20 0.00689655 sec
DELTA 0.00001311 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.30 usec
p2 20.60 usec
PL1 -1.00 dB
SFO1 100.6233329 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -3.00 dB
PL12 16.00 dB
SFO2 400.1316005 MHz
SI 32768
SF 100.6127697 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

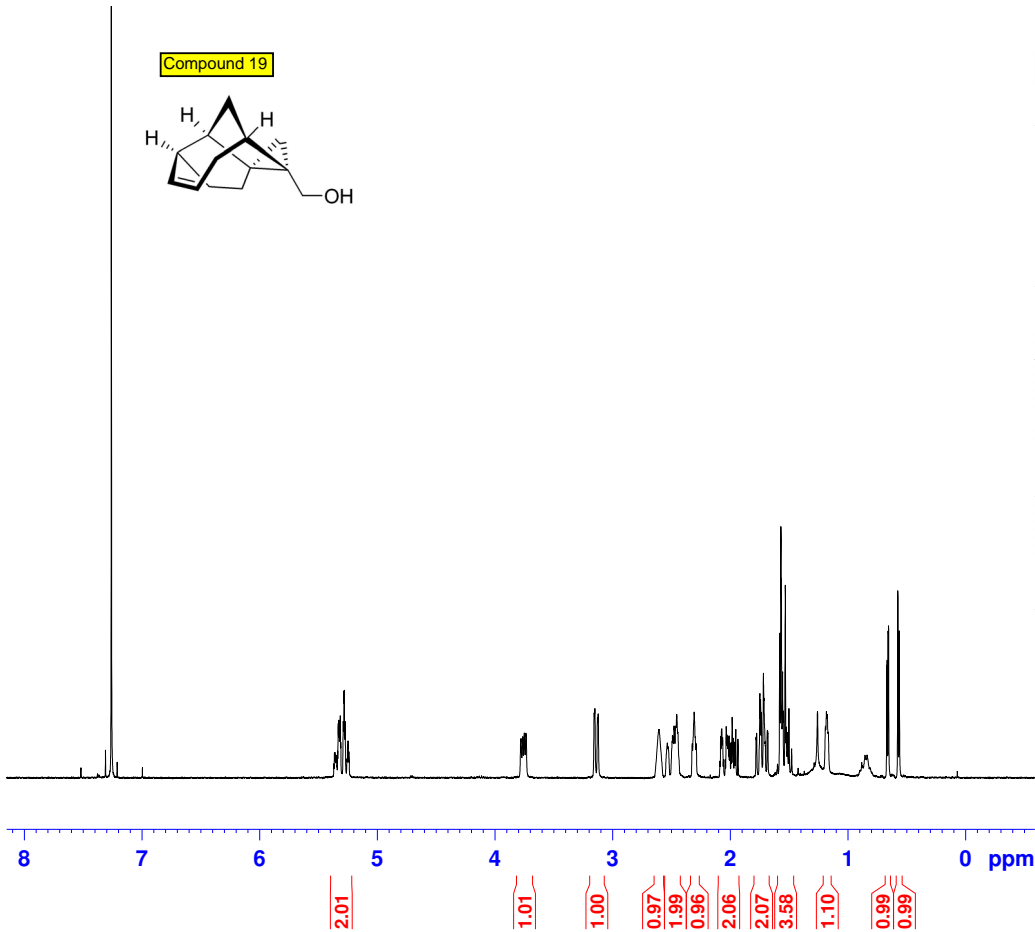


Compound 19

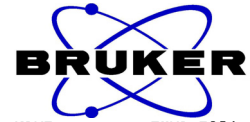
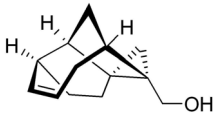


NAME THMA E254
EXPNO 20
PROCNO 1
Date_ 20090708
Time 9.54
INSTRUM AVIII400
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 912
DW 60.800 usec
DE 6.50 usec
TE 298.2 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.50 usec
PL1 -1.80 dB
PL1W 15.28361320 W
SFO1 400.2724718 MHz
SI 32768
SF 400.2700107 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



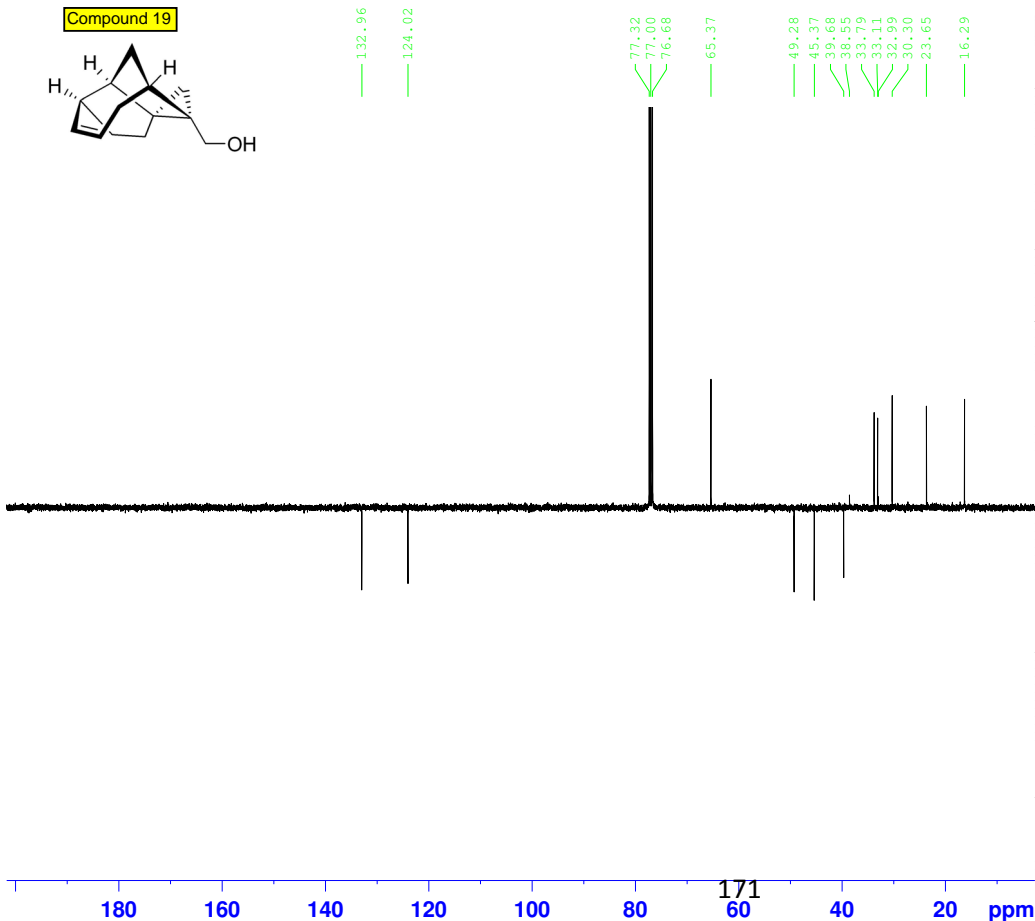
Compound 19



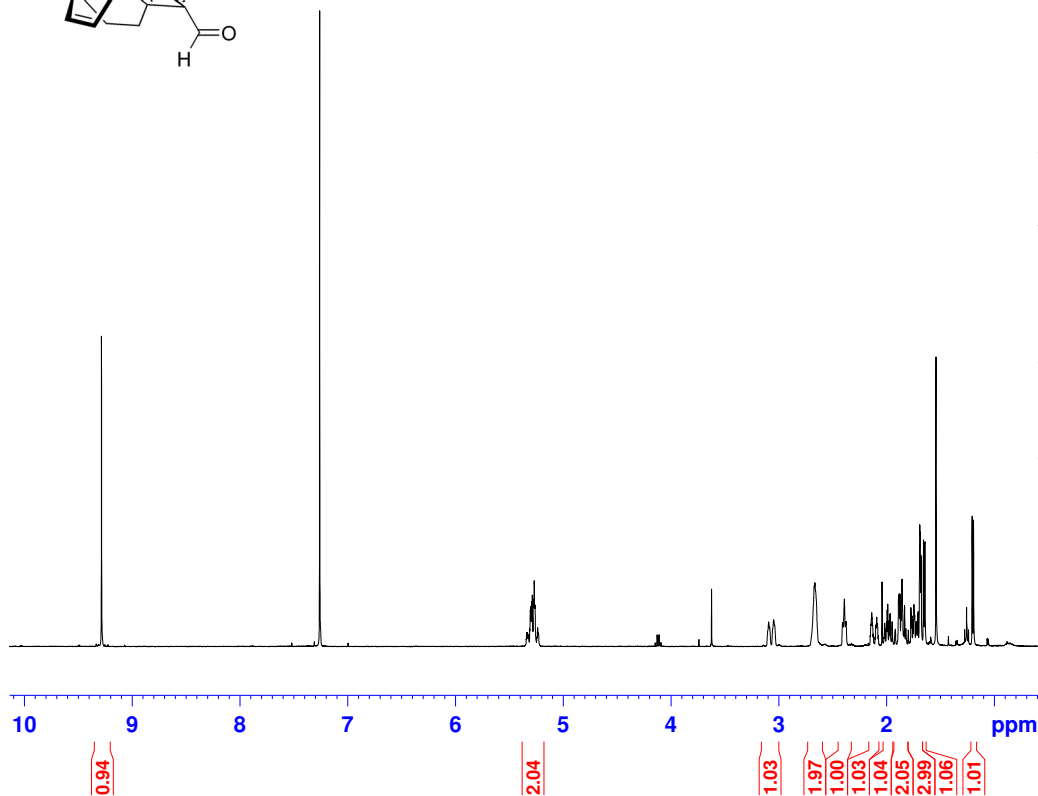
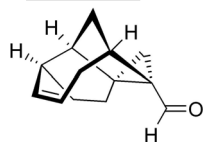
NAME THMA E254
EXPNO 21
PROCNO 1
Date_ 20090709
Time 3.43
INSTRUM AVIII400
PROBHD 5 mm PABBO BB-
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 4500
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 298.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
D20 0.00689655 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
P2 20.00 usec
PL1 -1.50 dB
PL1W 47.89980698 W
SFO1 100.6580364 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -1.80 dB
PL12 14.70 dB
PL2W 15.28361320 W
PL12W 0.34215751 W
SFO2 400.2716011 MHz
SI 32768
SF 100.6479720 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



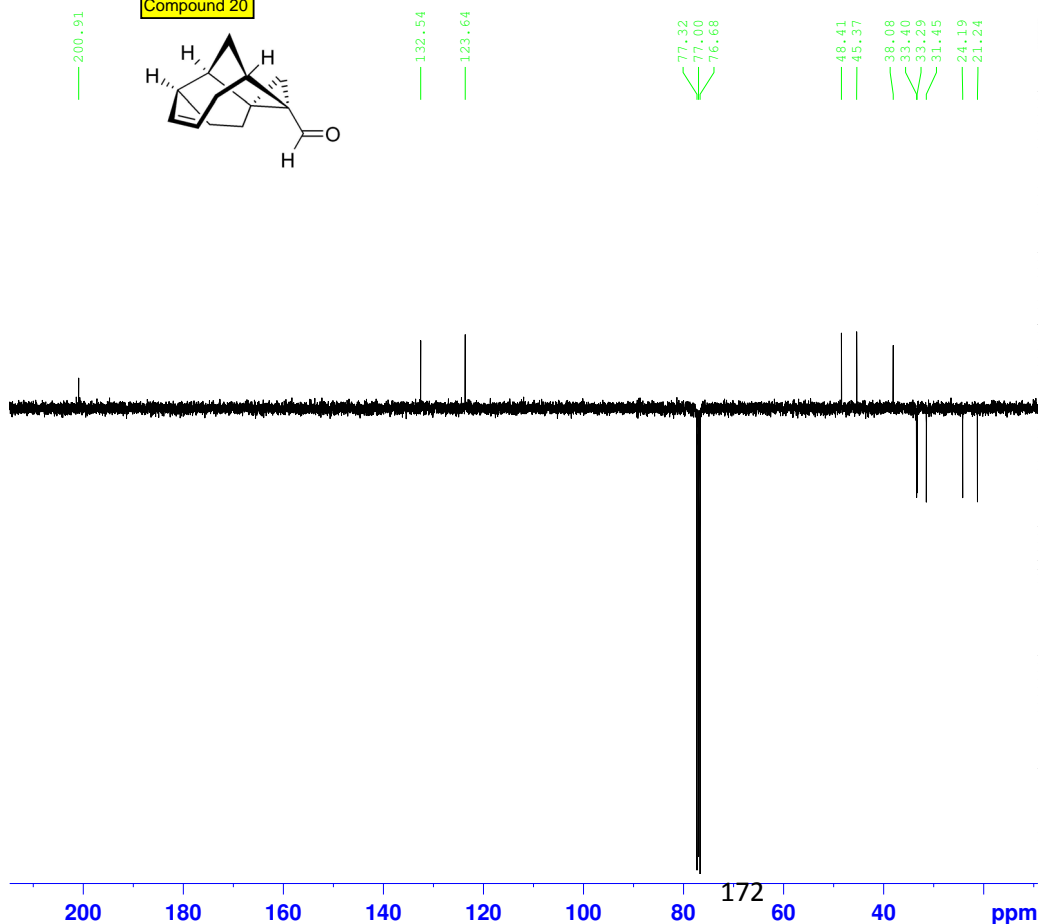
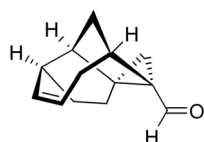
Compound 20



NAME THMA E206
EXPNO 60
PROCNO 1
Date_ 20090613
Time 0.49
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 32
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 645.1
DW 60.400 usec
DE 6.00 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 -3.00 dB
SFO1 400.1324710 MHz
SI 32768
SF 400.1300094 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

Compound 20

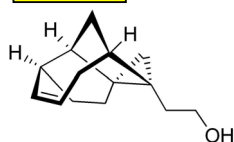


NAME THMA E206
EXPNO 61
PROCNO 1
Date_ 20090613
Time 2.42
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG jmod
TD 65536
SOLVENT CDCl₃
NS 2000
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3074932 sec
RG 7298.2
DW 19.950 usec
DE 6.00 usec
TE 300.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
d20 0.00689655 sec
DELTA 0.00001311 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.30 usec
p2 20.60 usec
PL1 -1.00 dB
SFO1 100.6233329 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -3.00 dB
PL12 16.00 dB
SFO2 400.1316005 MHz
SI 32768
SF 100.6127694 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

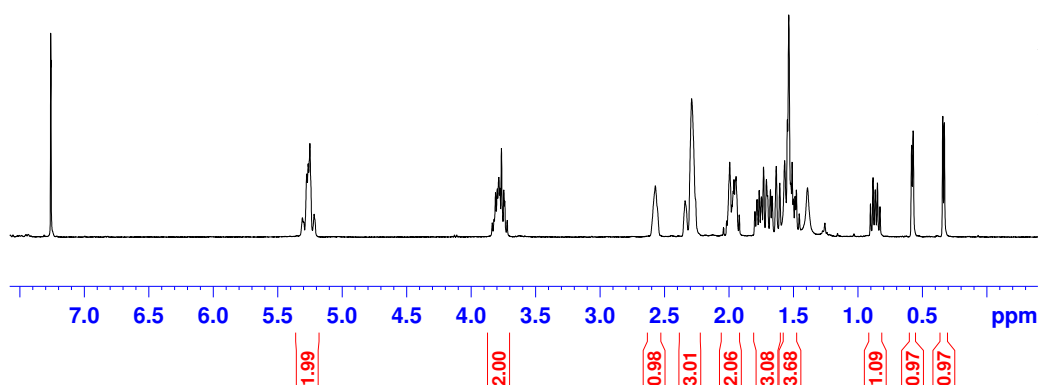
Compound 12



```

NAME      K 795 HF
EXPNO     30
PROCNO    1
Date_     20090615
Time      15.35
INSTRUM   avance400
PROBHD    5 mm BBO BB-1H
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        8278.146 Hz
FIDRES     0.126314 Hz
AQ         3.9584243 sec
RG         256
DW         60.400 usec
DE         6.00 usec
TE         300.2 K
D1         1.00000000 sec
TD0        1

```

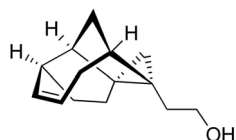


```

===== CHANNEL f1 =====
NUC1      1H
P1         8.75 usec
PL1        -3.00 dB
SFO1      400.1324710 MHz
SI         32768
SF         400.1300095 MHz
WDW        EM
SSB         0
LB         0.30 Hz
GB          0
PC         1.00

```

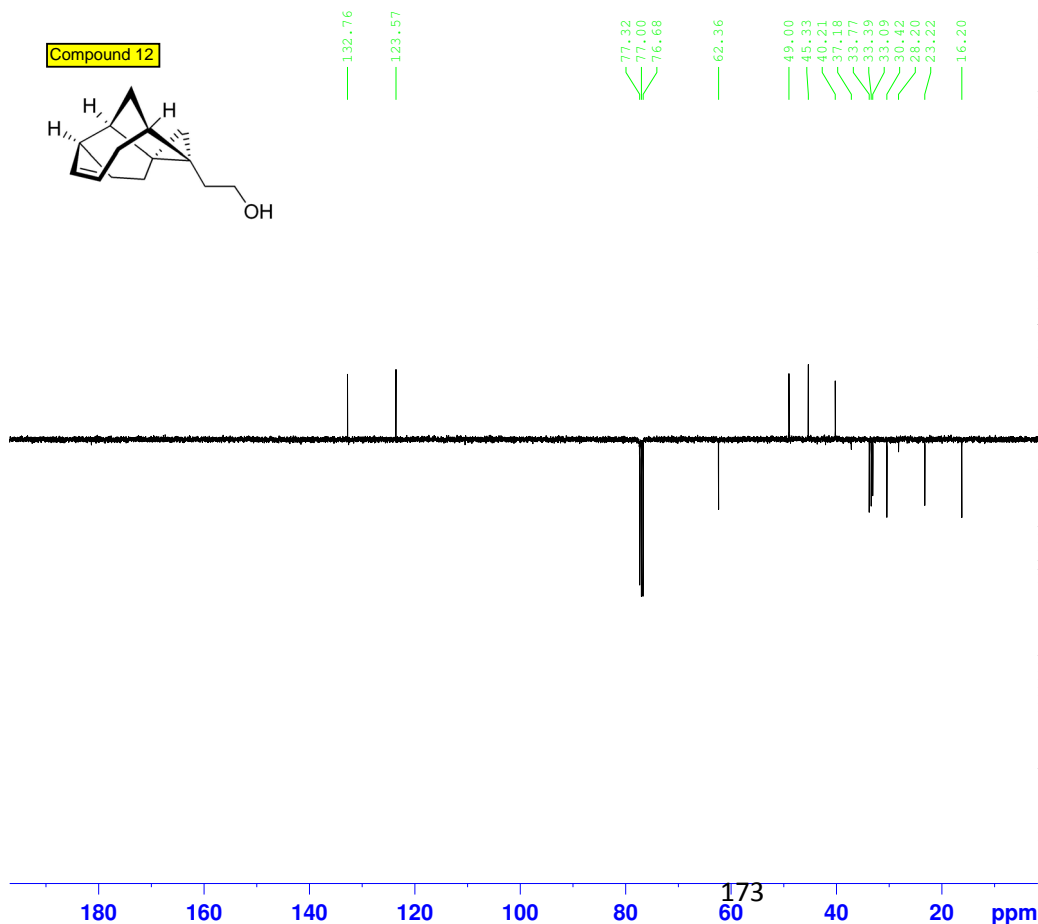
Compound 12



```

NAME      K 795 HF
EXPNO     31
PROCNO    1
Date_     20090615
Time      15.54
INSTRUM   avance400
PROBHD    5 mm BBO BB-1H
PULPROG   jmod
TD         65536
SOLVENT   CDCl3
NS         1200
DS         2
SWH        25062.656 Hz
FIDRES     0.382426 Hz
AQ         1.3074932 sec
RG         7298.2
DW         19.950 usec
DE         6.00 usec
TE         300.2 K
CNST2     145.0000000
CNST11     1.0000000
D1         2.00000000 sec
d20        0.00689655 sec
DELTA      0.00001311 sec
TD0        1

```



```

===== CHANNEL f1 =====
NUC1      13C
P1         10.30 usec
p2         20.60 usec
PL1        -1.00 dB
SFO1      100.6233329 MHz

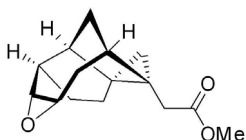
```

```

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2       1H
PCPD2     100.00 usec
PL2        -3.00 dB
PL12       16.00 dB
SFO2      400.1316005 MHz
SI         32768
SF         100.6127697 MHz
WDW        EM
SSB         0
LB         1.00 Hz
GB          0
PC         1.40

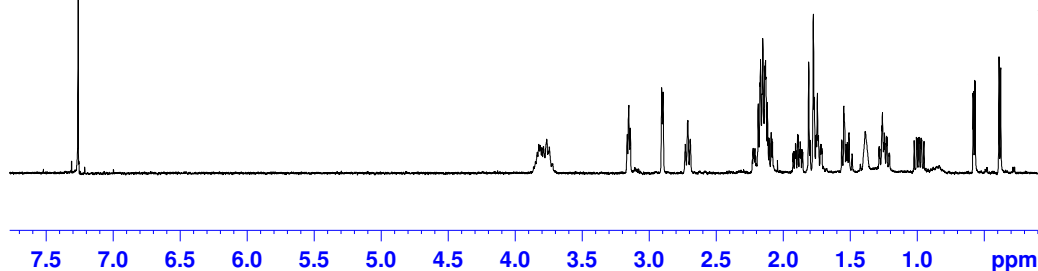
```

Compound 21



NAME THMA E257
EXPNO 370
PROCNO 1
Date_ 20090708
Time 19.18
INSTRUM AVIII400
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 1
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 645
DW 60.800 usec
DE 6.50 usec
TE 298.3 K
D1 1.00000000 sec
TD0 1

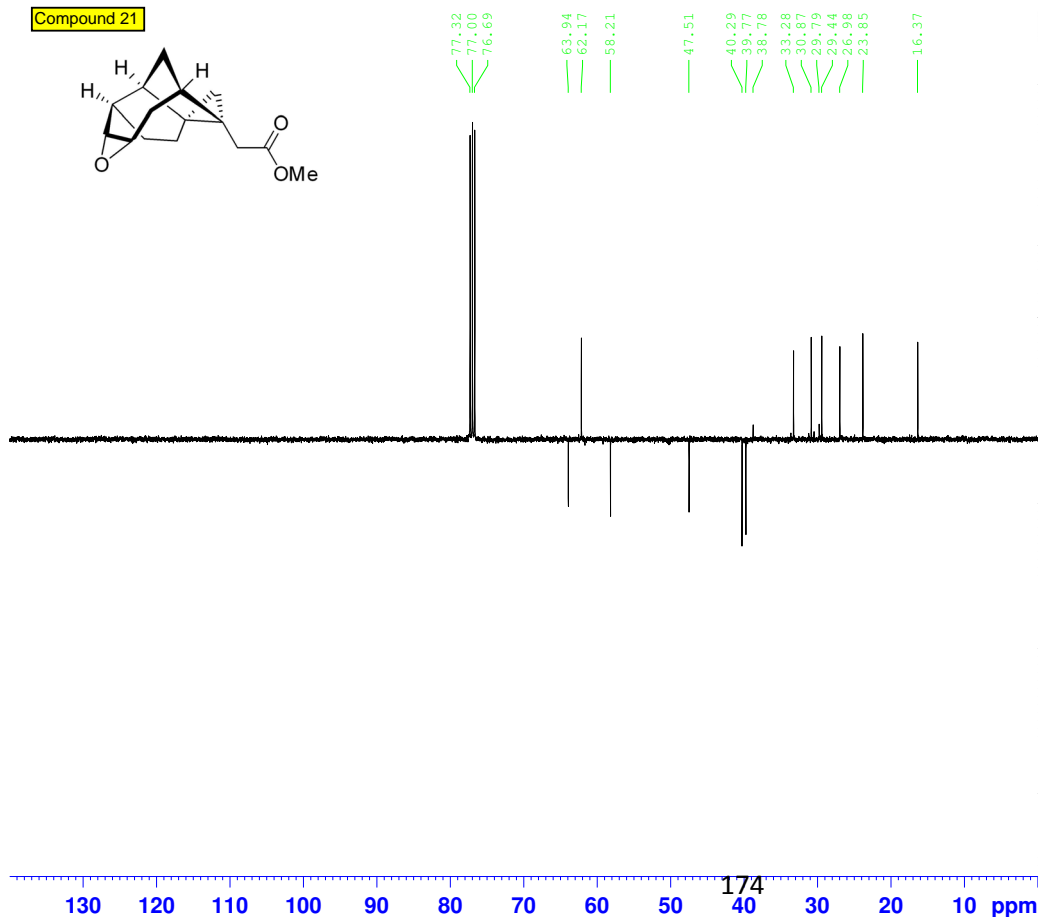
===== CHANNEL f1 =====
NUC1 1H
P1 13.50 usec
PL1 -1.80 dB
PL1W 15.28361320 W
SFO1 400.2724718 MHz
SI 32768
SF 400.2700105 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



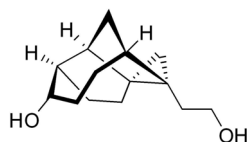
NAME THMA E257
EXPNO 371
PROCNO 1
Date_ 20090709
Time 6.37
INSTRUM AVIII400
PROBHD 5 mm PABBO BB-
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 3000
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 298.1 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
D20 0.00689655 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
P2 20.00 usec
PL1 -1.50 dB
PL1W 47.89980698 W
SFO1 100.6580364 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -1.80 dB
PL12 14.70 dB
PL2W 15.28361320 W
PL12W 0.34215751 W
SFO2 400.2716011 MHz
SI 32768
SF 100.6479720 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 3.00

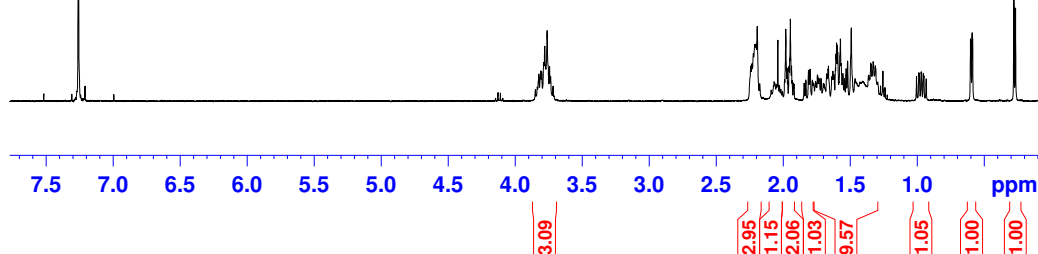


Compound 14

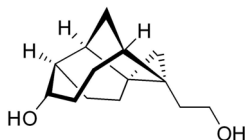


NAME THMA E258
EXPNO 60
PROCNO 1
Date_ 20090709
Time 12.15
INSTRUM AVIII400
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 1
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 456
DW 60.800 usec
DE 6.50 usec
TE 298.2 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.50 usec
PL1 -1.80 dB
PL1W 15.28361320 W
SFO1 400.2724718 MHz
SI 32768
SF 400.2700105 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Compound 14



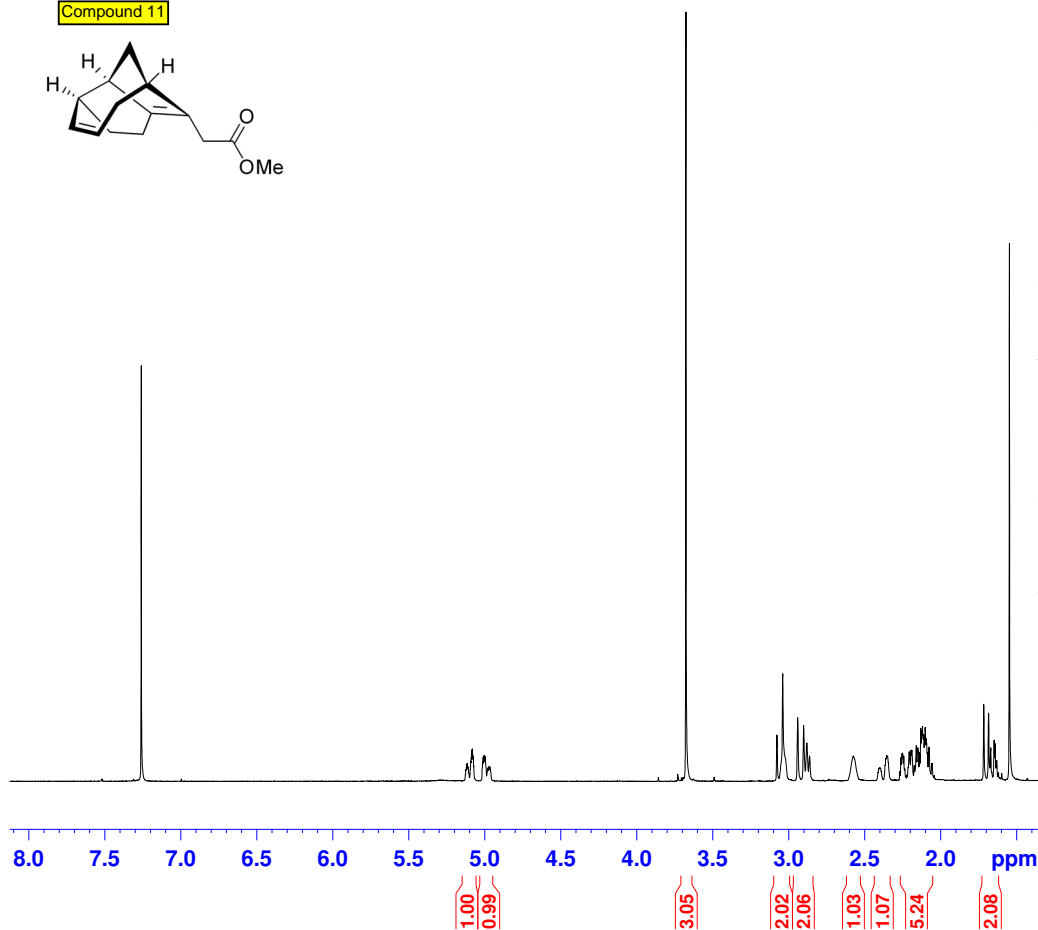
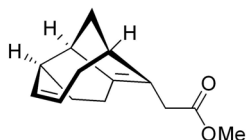
NAME THMA E258
EXPNO 61
PROCNO 1
Date_ 20090710
Time 0.21
INSTRUM AVIII400
PROBHD 5 mm PABBO BB-
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 4000
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 298.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
D20 0.00689655 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
P2 20.00 usec
PL1 -1.50 dB
PL1W 47.89980698 W
SFO1 100.6580364 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -1.80 dB
PL12 14.70 dB
PL2W 15.28361320 W
PL12W 0.34215751 W
SFO2 400.2716011 MHz
SI 32768
SF 100.6479720 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 2.00



Compound 11



```

NAME      K 801 HF
EXPNO     10
PROCNO    1
Date_     20090626
Time      15.18
INSTRUM   avance400
PROBHD    5 mm BBO BB-1H
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         32
DS         2
SWH        8278.146 Hz
FIDRES     0.126314 Hz
AQ         3.9584243 sec
RG         574.7
DW         60.400 usec
DE         6.00 usec
TE         300.2 K
D1         1.00000000 sec
TD0        1

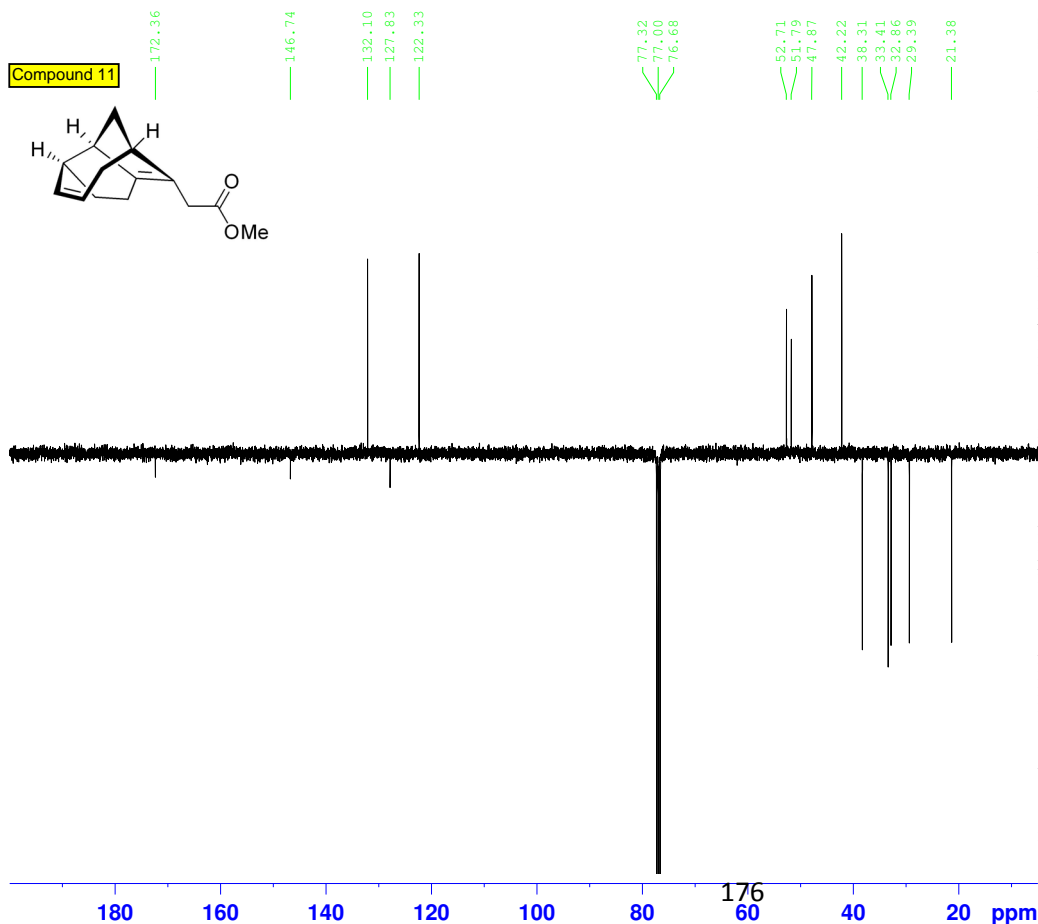
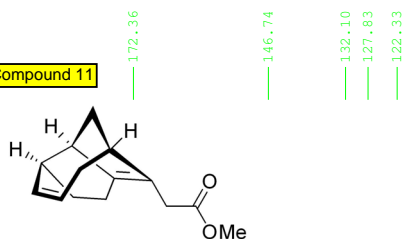
```

```

===== CHANNEL f1 =====
NUC1       1H
P1         8.75 usec
PL1        -3.00 dB
SFO1       400.1324710 MHz
SI         32768
SF         400.1300094 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

```

Compound 11



```

NAME      K 801 HF
EXPNO     15
PROCNO    1
Date_     20090627
Time      0.07
INSTRUM   avance400
PROBHD    5 mm BBO BB-1H
PULPROG   jmod
TD         65536
SOLVENT   CDCl3
NS         5000
DS         2
SWH        25062.656 Hz
FIDRES     0.382426 Hz
AQ         1.3074932 sec
RG         5792.6
DW         19.950 usec
DE         6.00 usec
TE         300.2 K
CNST2     145.0000000
CNST11    1.0000000
D1         2.00000000 sec
d20        0.00689655 sec
DELTA     0.00001311 sec
TD0        1

```

```

===== CHANNEL f1 =====
NUC1       13C
P1         10.30 usec
p2         20.60 usec
PL1        -1.00 dB
SFO1       100.6233329 MHz

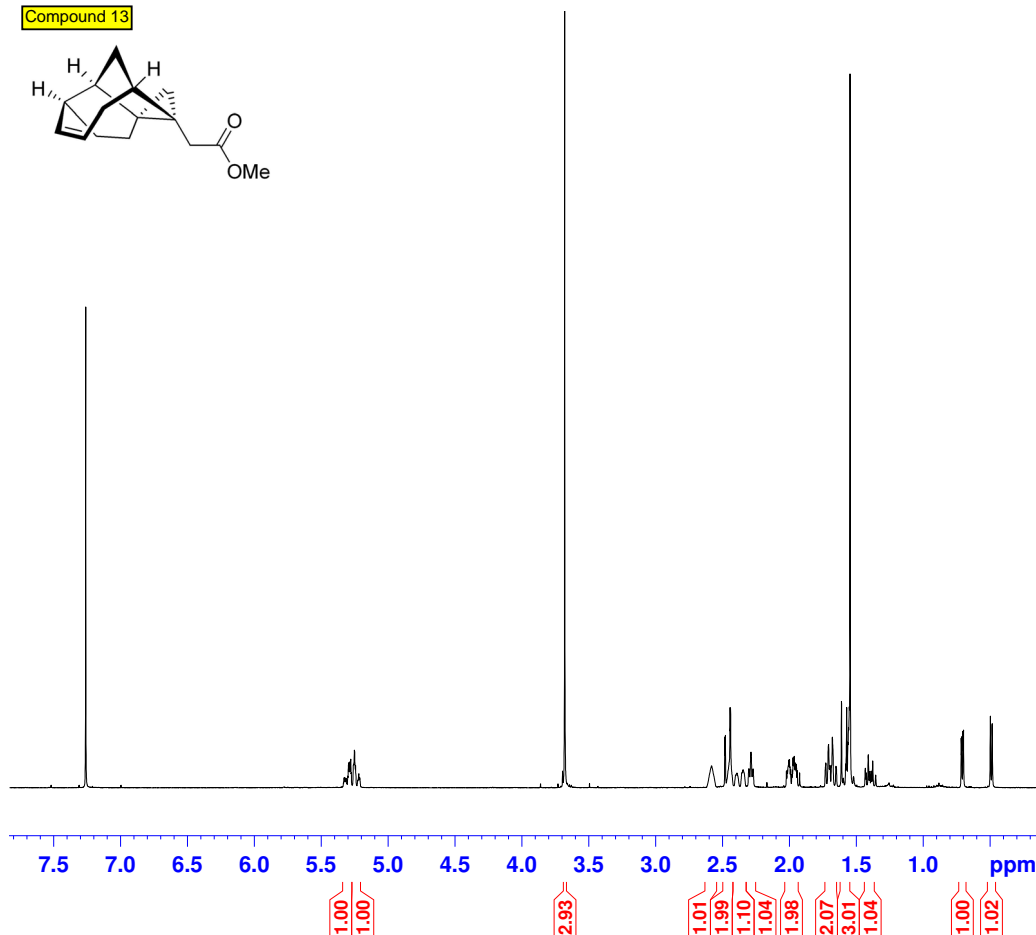
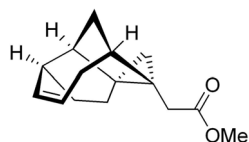
```

```

===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2        1H
PCPD2       100.00 usec
PL2         -3.00 dB
PL12        16.00 dB
SFO2       400.1316005 MHz
SI         32768
SF         100.6127695 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40

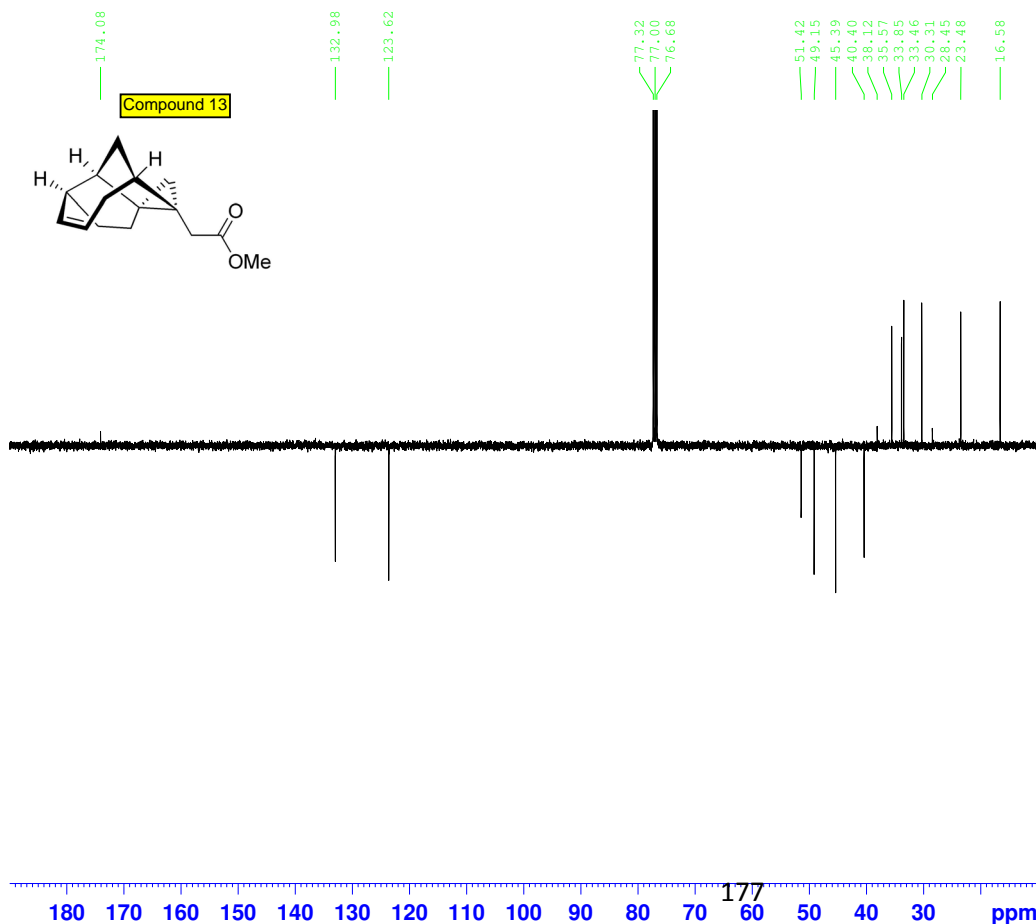
```

Compound 13



NAME K 807 HF
EXPNO 40
PROCNO 1
Date_ 20090709
Time 13.42
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 32
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 512
DW 60.400 usec
DE 6.00 usec
TE 300.2 K
D1 1.00000000 sec
TD0 1

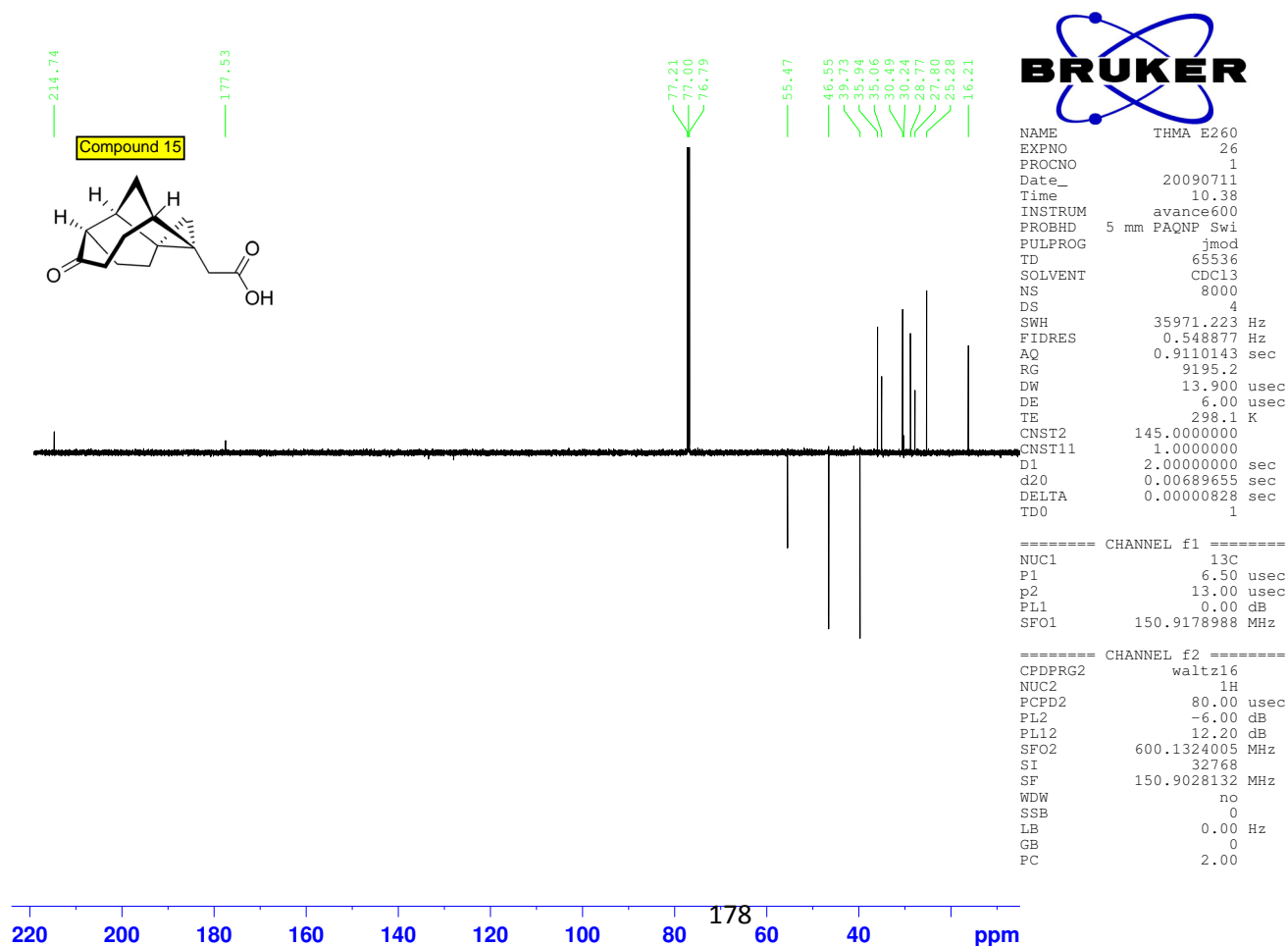
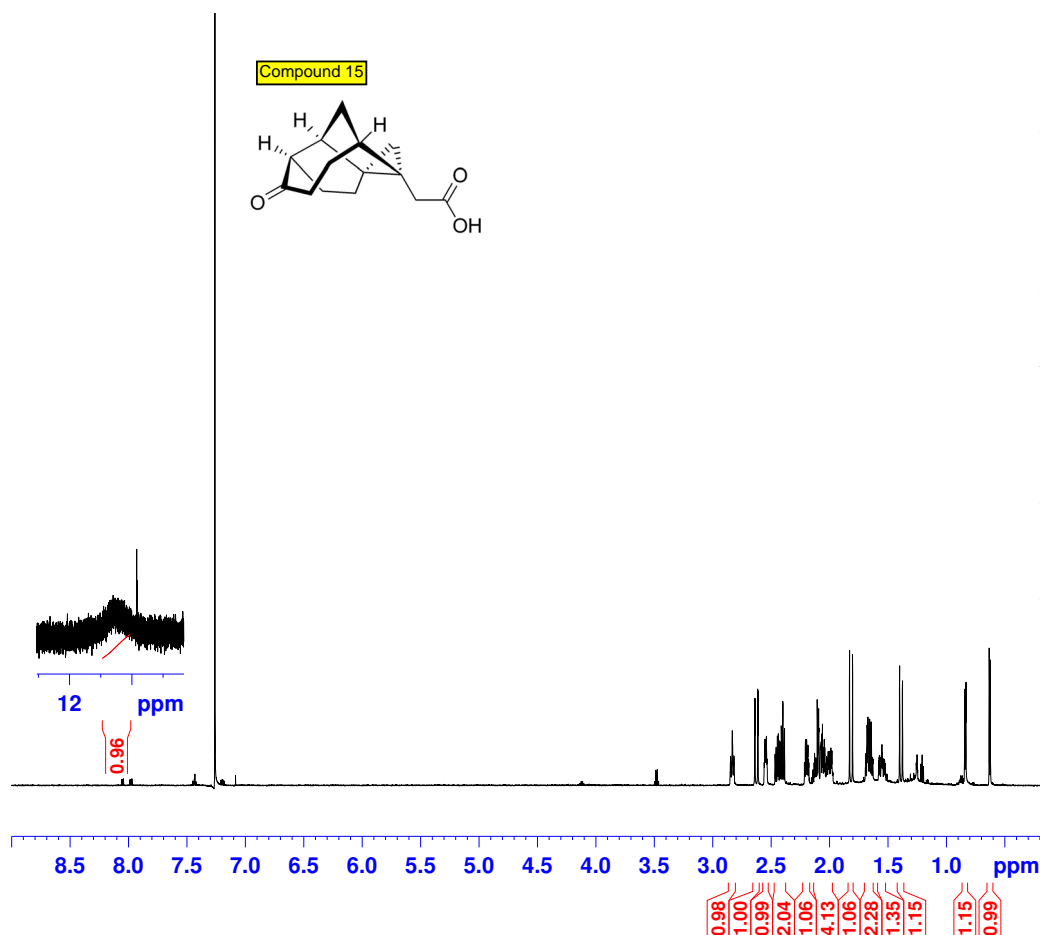
===== CHANNEL f1 =====
NUC1 1H
P1 8.75 usec
PL1 -3.00 dB
SFO1 400.1324710 MHz
SI 32768
SF 400.1300093 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

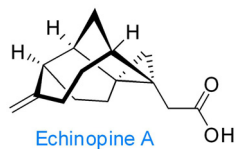


NAME K 807 HF
EXPNO 41
PROCNO 1
Date_ 20090709
Time 14.06
INSTRUM avance400
PROBHD 5 mm BBO BB-1H
PULPROG jmod
TD 65536
SOLVENT CDCl3
NS 6000
DS 2
SWH 25062.656 Hz
FIDRES 0.382426 Hz
AQ 1.3074932 sec
RG 14596.5
DW 19.950 usec
DE 6.00 usec
TE 300.2 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
d20 0.00689655 sec
DELTA 0.00001311 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.30 usec
p2 20.60 usec
PL1 -1.00 dB
SFO1 100.6233329 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -3.00 dB
PL12 16.00 dB
SFO2 400.1316005 MHz
SI 32768
SF 100.6127692 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



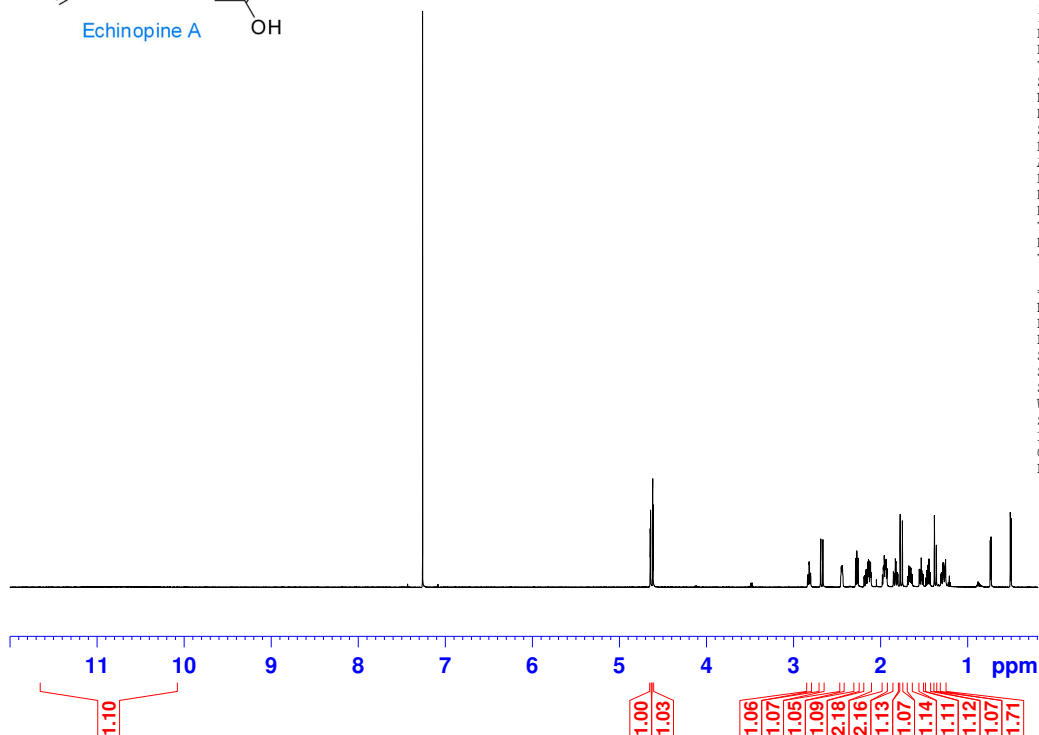


```

NAME      THMA Echinopine A
EXPNO     10
PROCNO    1
Date_     20090710
Time      14.09
INSTRUM   avance600
PROBHD    5 mm PAQNP Swi
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         32
DS         2
SWH        12376.237 Hz
FIDRES     0.188846 Hz
AQ         2.6477449 sec
RG         406.4
DW         40.400 usec
DE         6.00 usec
TE         298.1 K
D1         1.00000000 sec
TD0        1
  
```

```

===== CHANNEL f1 =====
NUC1      1H
P1        13.85 usec
PL1       -6.00 dB
SFO1      600.1337060 MHz
SI        32768
SF        600.1300174 MHz
WDW       no
SSB       0
LB        0.00 Hz
GB        0
PC        1.00
  
```

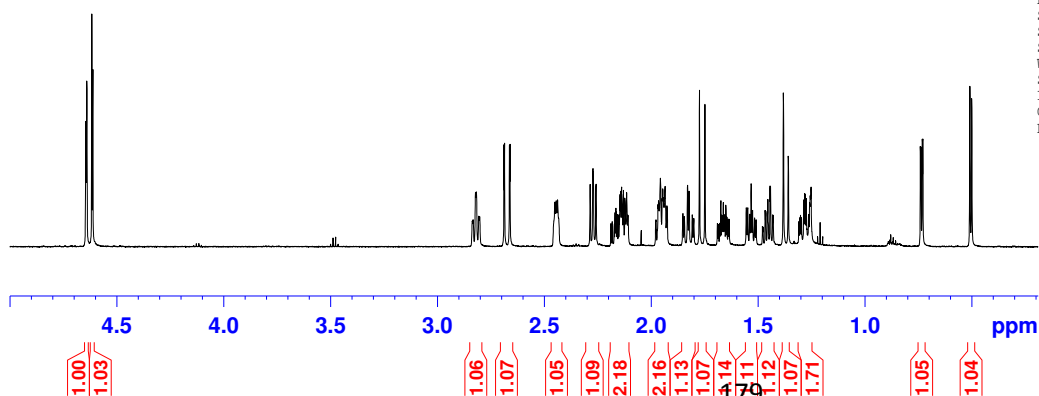


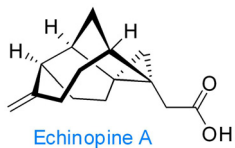
```

NAME      THMA Echinopine A
EXPNO     10
PROCNO    1
Date_     20090710
Time      14.09
INSTRUM   avance600
PROBHD    5 mm PAQNP Swi
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         32
DS         2
SWH        12376.237 Hz
FIDRES     0.188846 Hz
AQ         2.6477449 sec
RG         406.4
DW         40.400 usec
DE         6.00 usec
TE         298.1 K
D1         1.00000000 sec
TD0        1
  
```

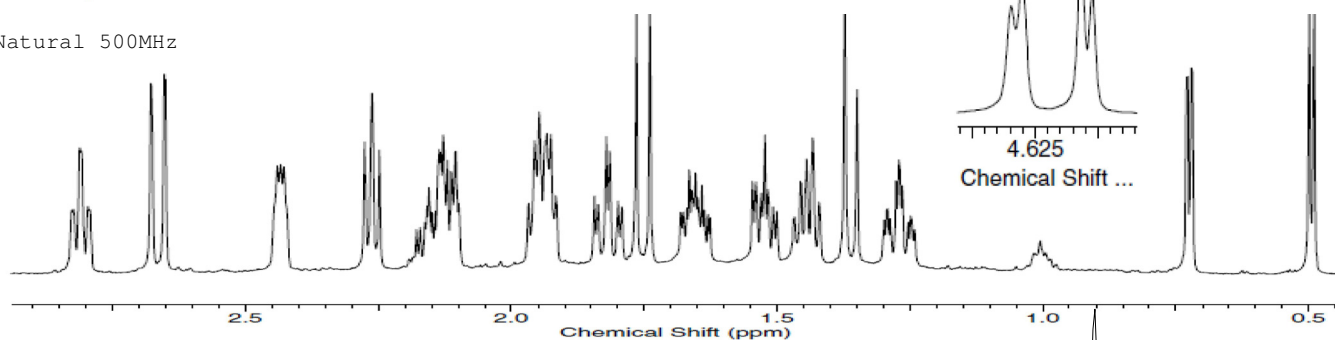
```

===== CHANNEL f1 =====
NUC1      1H
P1        13.85 usec
PL1       -6.00 dB
SFO1      600.1337060 MHz
SI        32768
SF        600.1300174 MHz
WDW       no
SSB       0
LB        0.00 Hz
GB        0
PC        1.00
  
```

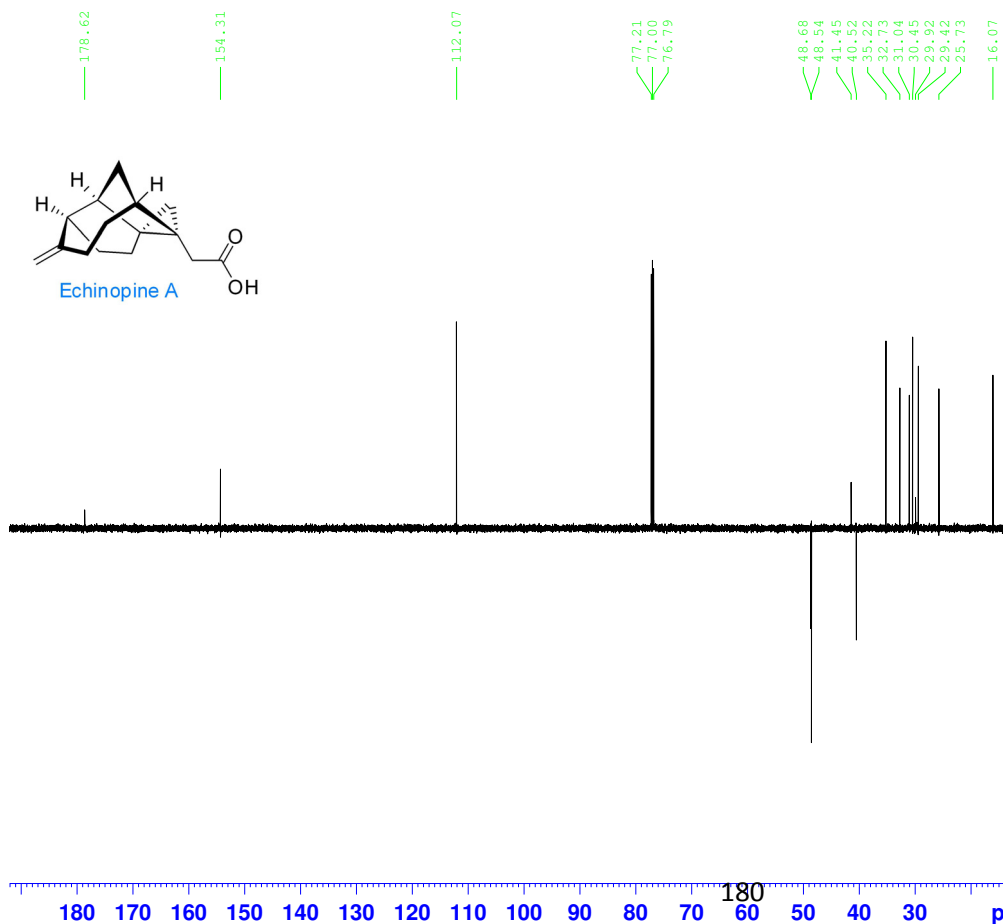
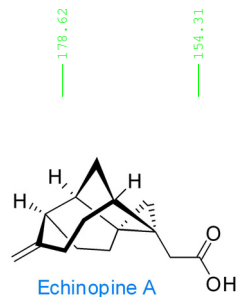
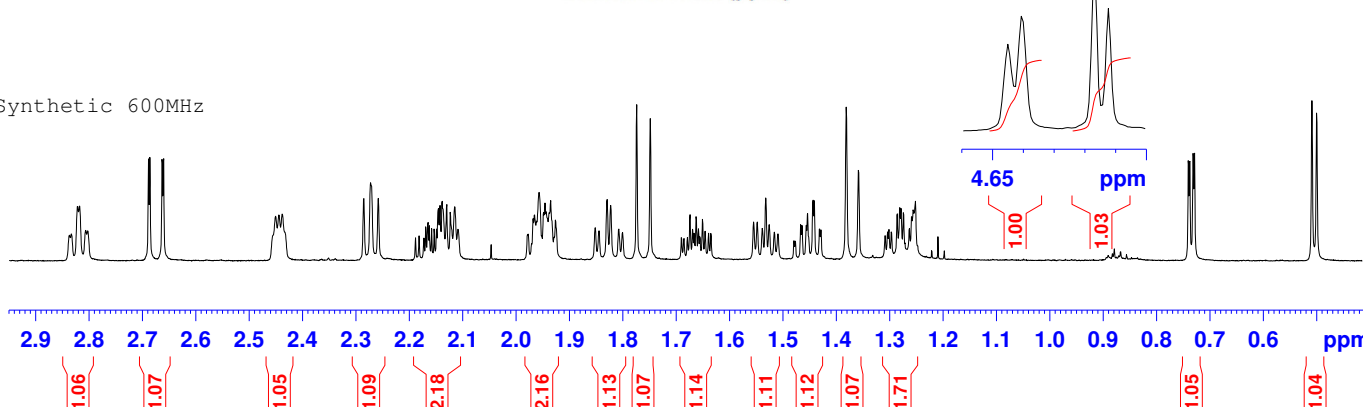




Natural 500MHz



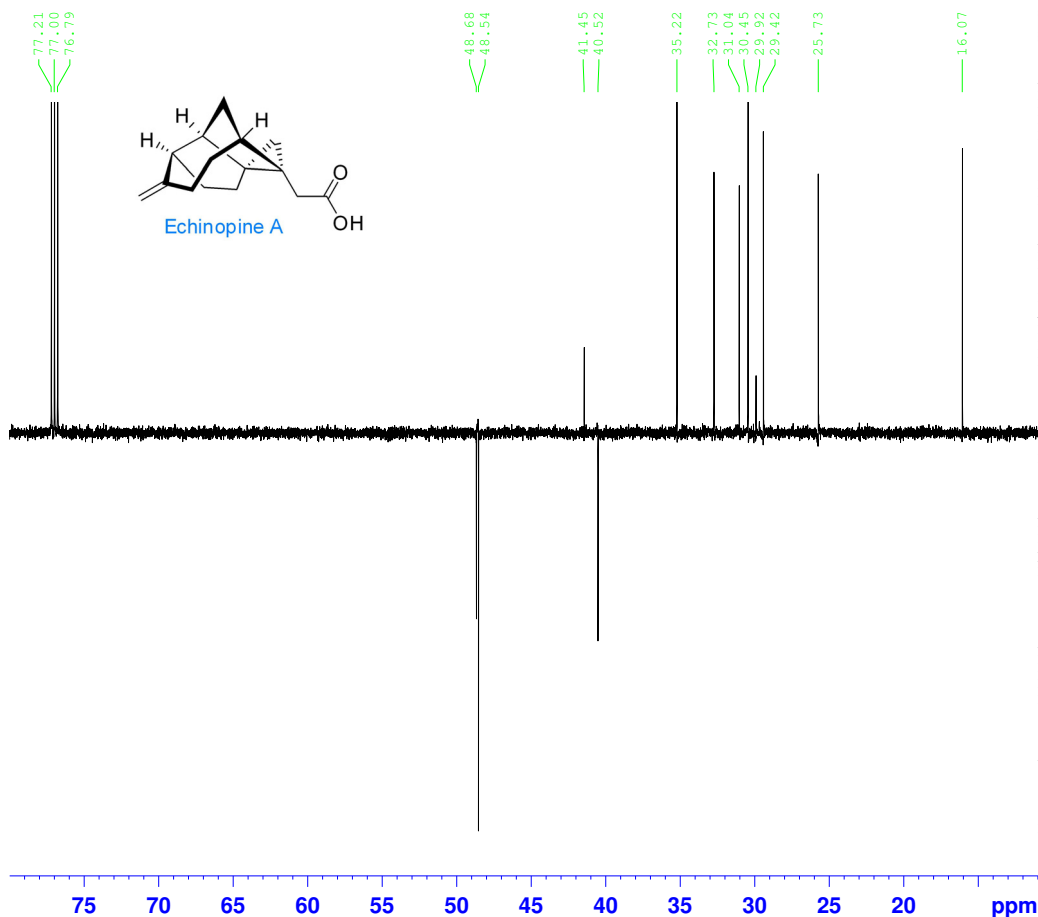
Synthetic 600MHz



NAME THMA Echinopine A
 EXPNO 16
 PROCNO 1
 Date_ 20090710
 Time 20.21
 INSTRUM avance600
 PROBHD 5 mm PAQNP Swi
 PULPROG jmod
 TD 65536
 SOLVENT CDC13
 NS 3000
 DS 4
 SWH 35971.223 Hz
 FIDRES 0.548877 Hz
 AQ 0.9110143 sec
 RG 10321.3
 DW 13.900 usec
 DE 6.00 usec
 TE 298.1 K
 CNST2 145.0000000
 CNST11 1.0000000
 D1 2.0000000 sec
 d20 0.00689655 sec
 DELTA 0.00000828 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 6.50 usec
 p2 13.00 usec
 PL1 0.00 dB
 SFO1 150.9178988 MHz

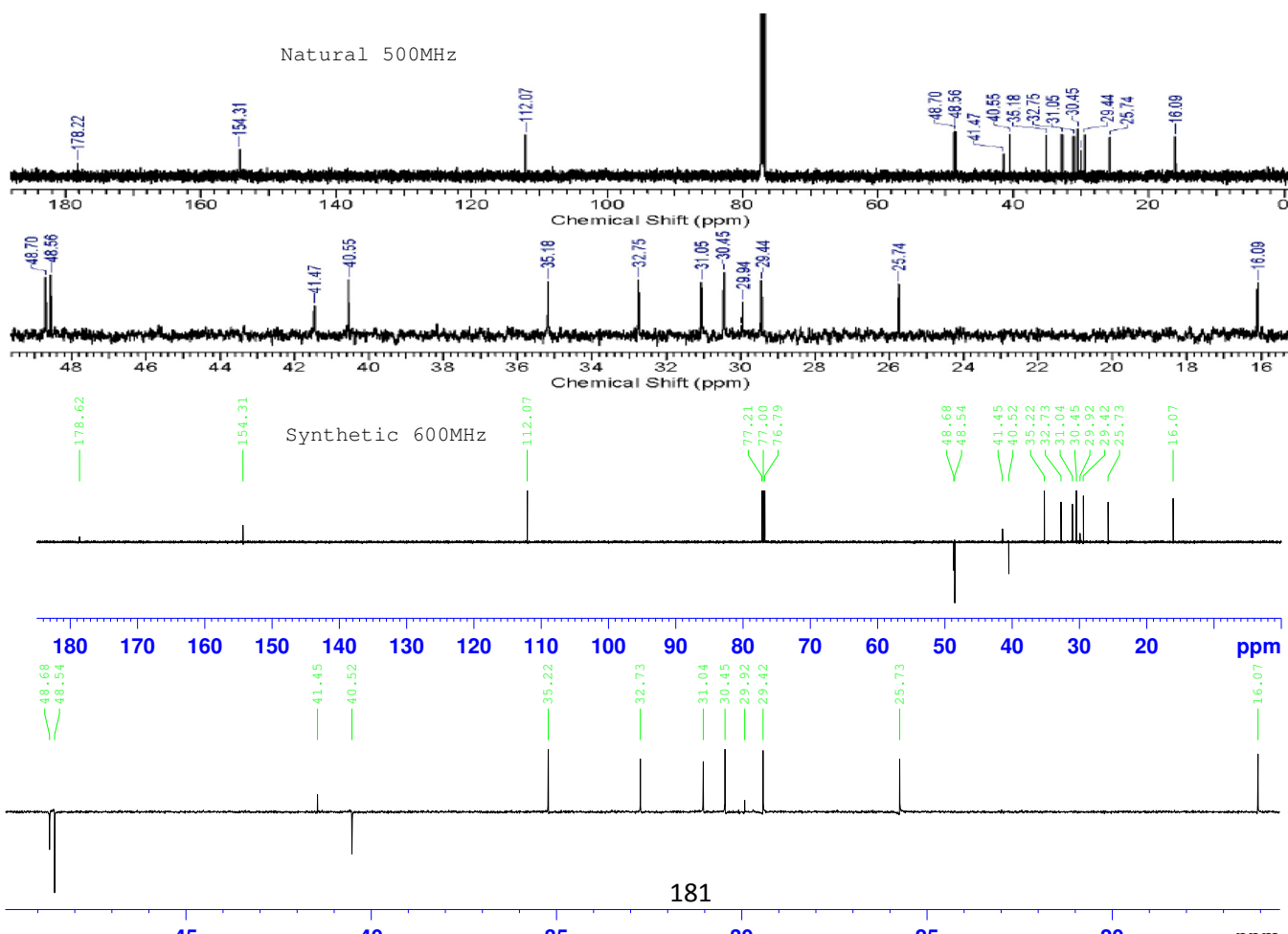
===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -6.00 dB
 PL12 12.20 dB
 SFO2 600.1324005 MHz
 SI 32768
 SF 150.9028132 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 3.00

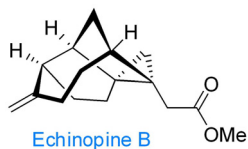


NAME THMA Echinopine A
 EXPNO 16
 PROCNO 1
 Date_ 20090710
 Time 20.21
 INSTRUM advance600
 PROBHD 5 mm PAQNP Swi
 PULPROG jmod
 TD 65536
 SOLVENT CDCl3
 NS 3000
 DS 4
 SWH 35971.223 Hz
 FIDRES 0.548877 Hz
 AQ 0.9110143 sec
 RG 10321.3
 DW 13.900 usec
 DE 6.00 usec
 TE 298.1 K
 CNST2 145.0000000
 CNST11 1.0000000
 D1 2.00000000 sec
 d20 0.00689655 sec
 DELTA 0.00000828 sec
 TD0 1

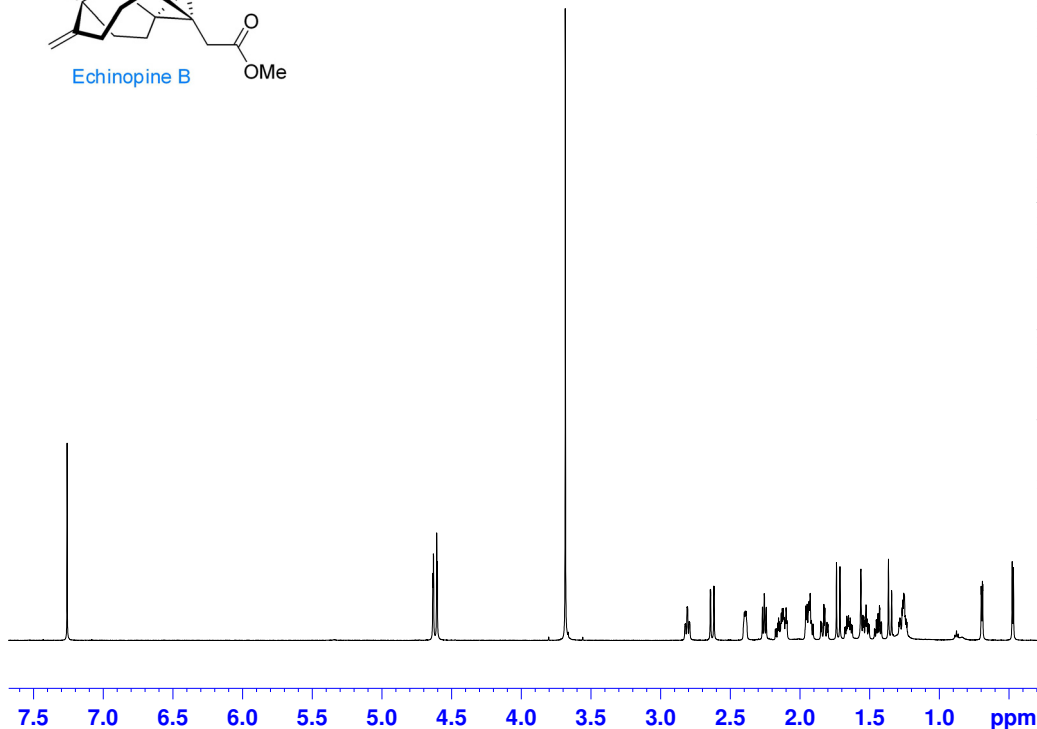
===== CHANNEL f1 =====
 NUC1 13C
 P1 6.50 usec
 p2 13.00 usec
 PL1 0.00 dB
 SFO1 150.9178988 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -6.00 dB
 PL12 12.20 dB
 SFO2 600.1324005 MHz
 SI 32768
 SF 150.9028132 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 3.00

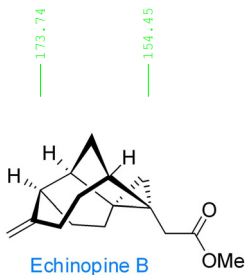




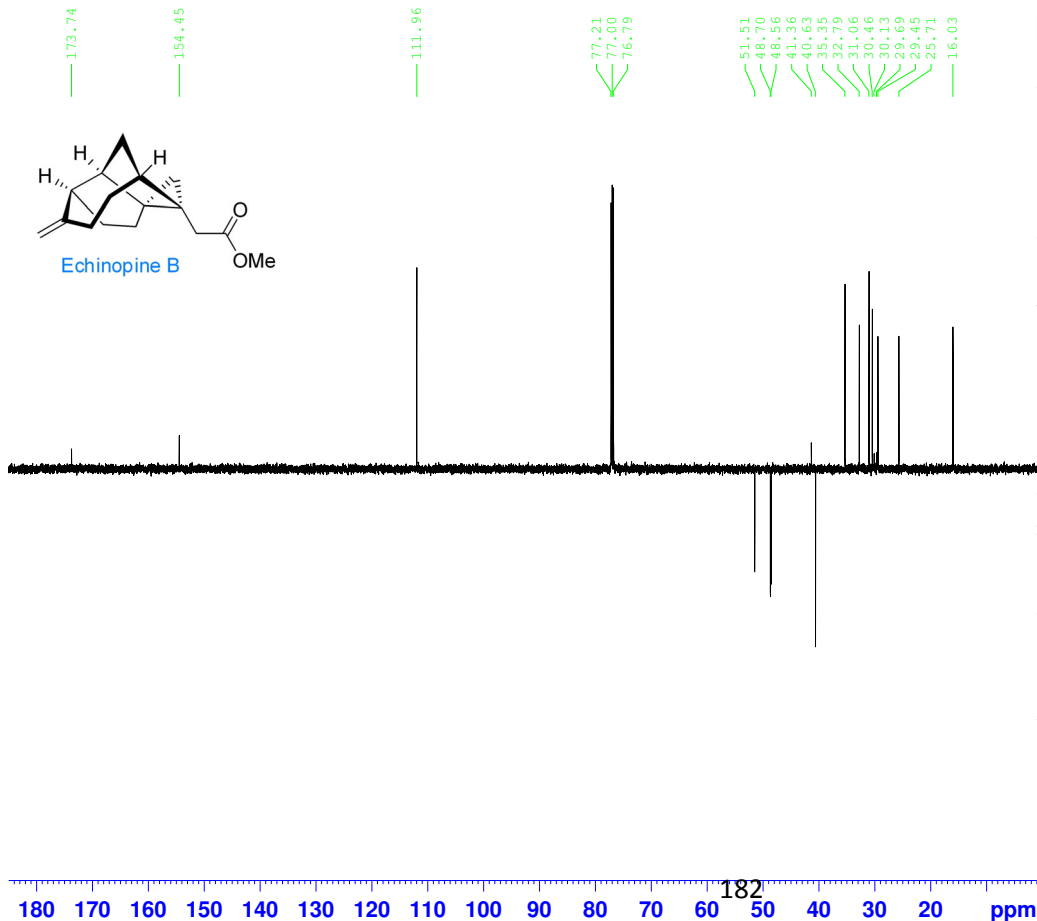
NAME THMA Echinopine B opt
EXPNO 10
PROCNO 1
Date_ 20090713
Time 14.56
INSTRUM advance600
PROBHD 5 mm PAQNP Swi
PULPROG zg30
TD 65536
SOLVENT CDC13
NS 16
DS 2
SWH 12376.237 Hz
FIDRES 0.188846 Hz
AQ 2.6477449 sec
RG 287.4
DW 40.400 usec
DE 6.00 usec
TE 298.1 K
D1 1.00000000 sec
TD0 1



===== CHANNEL f1 =====
NUC1 1H
P1 13.85 usec
PL1 -6.00 dB
SFO1 600.1337060 MHz
SI 32768
SF 600.1300174 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



NAME THMA Echinopine B opt
EXPNO 11
PROCNO 1
Date_ 20090713
Time 15.08
INSTRUM advance600
PROBHD 5 mm PAQNP Swi
PULPROG jmod
TD 65536
SOLVENT CDC13
NS 1158
DS 4
SWH 35971.223 Hz
FIDRES 0.548877 Hz
AQ 0.9110143 sec
RG 5792.6
DW 13.900 usec
DE 6.00 usec
TE 298.1 K
CNST2 145.0000000
CNST11 1.0000000
D1 2.00000000 sec
d20 0.00689655 sec
DELTA 0.00000828 sec
TD0 1



===== CHANNEL f1 =====
NUC1 13C
P1 6.50 usec
p2 13.00 usec
PL1 0.00 dB
SFO1 150.9178988 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -6.00 dB
PL12 12.20 dB
SFO2 600.1324005 MHz
SI 32768
SF 150.9028128 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

8. Summary

In conclusion, we have accomplished the first total syntheses of the sesquiterpenoids (+)-echinopine A and B and the determination of their absolute stereochemistry.

The key features are: (1) The stereoselective installation of the vinyl-moiety on the hindered concave side of the RCM precursor via Myers' [3,3]-sigmatropic rearrangement. (2) The successful equilibration of the C7-substituent to the desired concave face under basic conditions. (3) The installation of the highly strained 7-membered ring via RCM. (4) The extension of the unusual Pd(0)-mediated C2-homologation of aryltriflates with a ketene silyl acetal. (5) 1,5-cyclooctadiene as an inexpensive, readily available starting material. The efficient syntheses, (14)15 linear steps from known enantiopure and easily accessible starting material allows for diversification and the preparation of sufficient material for proper biological testing. In addition, we were able to grow crystals suitable for crystal structure analysis. The X-ray structure of synthetic echinopine A confirmed the proposed structure.

Unfortunately we were not able to realize the originally planned dynamic RCM, which would have saved an additional step, under a variety of conditions. All efforts to install the 7-membered ring bearing a trisubstituted double bond via RCM failed, probably due to the high ring strain.

9. Abbreviations

| | |
|-------|---------------------------------------|
| Ac | acetyl |
| ACP | acyl carrier protein |
| ADP | adenosinediphosphate |
| AIBN | azobisisobutyronitrile |
| AT | acyl transferase |
| ATP | adenosinetriphosphate |
| 9-BBN | 9-borabicyclo[3.3.1]nonane |
| Bn | benzyl |
| CAN | ceric ammonium nitrate |
| CBS | Corey–Bakshi–Shibata reduction |
| CL | CoA–ligase |
| COSY | correlated spectroscopy |
| CSA | camphersulfonic acid |
| CTP | cytidine triphosphate |
| Cy | cyclohexyl |
| DBU | 1,8-diaza-bicyclo[5.4.0]–7-undecene |
| DIBAL | diisobutylaluminium hydride |
| DCC | <i>N,N'</i> -dicyclohexylcarbodiimide |
| DIPEA | diisopropylethylamine (Huenig's base) |
| DH | β -hydroxyacyl dehydratase |
| DMF | <i>N,N'</i> -dimethylformamide |
| DMAP | 4-dimethylaminopyridine |

| | |
|---------------|---|
| DMAPP | dimethylallyl diphosphate |
| DME | dimethoxyethane |
| DMP | Dess–Martin periodinane |
| DMPS | dimethylphenylsilyl |
| DMSO | dimethyl sulfoxide |
| DXP | 1–deoxyxylose–5–phosphate |
| Ee | enantiomeric excess |
| EDA | ethyl diazo acetate |
| ER | enoyl reductase |
| GGP | geranyl–geranyl diphosphate |
| GPP | geranyl diphosphate |
| HMBC | <i>hetero</i> –nuclear multiple bond correlation |
| HMG | 3–hydroxy–3–methylglutaryl |
| HMPA | hexamethylphosphoramide |
| HMTA | hexamethylenetetramine |
| HRMS | high resolution mass spectrometry |
| IBX | 2–iodoxybenzoic acid |
| IPP | isopentenyl diphosphate |
| KR | β –ketoacyl reductase |
| KS | ketoacyl synthase |
| LDA | lithium diisopropylamide |
| LAH | lithium aluminiumhydride |
| LHMDS | lithium bis(trimethylsilyl)amide |
| L–selectride | lithium <i>tri</i> – <i>sec</i> –butylborohydride |
| <i>m</i> CPBA | <i>meta</i> –chloroperoxybenzoic acid |

| | |
|-------|--|
| MEM | methoxyethyl |
| MOM | methoxymethyl |
| MRSA | multi-resistant <i>Staphylococcus Aureus</i> |
| Ms | methanesulfonyl (mesyl) |
| MS | molecular sieve |
| MT | methyl transferase |
| MTPA | methoxy trifluoromethyl phenylacetic acid |
| MVA | mevalonic acid |
| NADPH | nicotinamide adenine dinucleotide phosphate |
| NIS | <i>N</i> -iodosuccinimide |
| NOE | nuclear Overhauser enhancement |
| PKS | polyketide synthase |
| PMB | <i>para</i> -methoxybenzyl |
| PPTS | pyridinium <i>para</i> -toluenesulfonate |
| PTC | phase transfer catalyst |
| RCM | ring closing metathesis |
| rfx | reflux |
| r.t. | room temperature |
| TBAF | tetrabutylammonium fluoride |
| TBS | <i>tert</i> -butyldimethylsilyl |
| TBDPS | <i>tert</i> -butyldiphenylsilyl |
| TE | thioesterase |
| TEA | triethylamine |
| TES | triethylsilyl |
| Tf | trifluoromethanesulfonyl |

Abbreviations

| | |
|------|------------------------------|
| TFA | trifluoroacetic acid |
| THF | tetrahydrofuran |
| THP | tetrahydropyran |
| TIPS | triisopropylsilyl |
| TMS | trimethylsilyl |
| TPP | thiamine pyrophosphate |
| Ts | <i>para</i> -toluenesulfonyl |
| TS | transition state |

D. Curriculum Vitae

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Education

Since November 2007 University of Vienna – Ph.D. research: “*Total Syntheses of (-)-Kendomycin and (+)-Echinopine A and*” in the group of Prof. Dr. Johann Mulzer

23.10.2007 University of Vienna – Graduation in minimum time (10 terms) and with distinction (*Magister rer. nat.*, comparable to M.Sc.)

Feb. 2007-Oct. 2007 University of Vienna - Diploma thesis: “*Towards the Synthesis of the ansa-Macrocycle Kendomycin*” in the research group of Prof. Dr. Johann Mulzer

2002-2007 University of Vienna – Study of Chemistry, specialized in Organic Chemistry, Spectroscopy and Analytical Chemistry

2002 Military service at HUAK Enns.

1993-2001 Gymnasium Werndlpark, Leopold-Werndl Straße 5, 4400 Steyr, focusing on math and science.

1989-1993 Primary school Karl-Punzer-Straße 73-75, 4400 Steyr

Internships

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| Summer 1999 | Plumber Schlossgangl, trainee |
| Autumn 2001 | BMW Automobiles, trainee for 3 months |
| Summer 2003 | Company SLR-Gusswerk II, casting trainee |
| Summer 2004 | Company Naintsch-Luzenac, R & D laboratory |
| Summer 2005 | Company MAN Steyrer Werke, Dept. of environment |
| Summer 2005 | BMW Automobiles, R & D laboratory |
| Summer term 2007 | University of Vienna, tutor of undergraduate students in the organic chemistry lab |
| February to October 2007 | University of Vienna, Research assistant. |
| Since November 2007 | University of Vienna, Assistant position. Teaching, supervision, tutoring and grading students in organic chemistry |

Awards

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| 2005 | Leistungsstipendium (University of Vienna) |
| 2008 | Gesellschaft Österreichischer Chemiker (GÖCH) Diploma Thesis Prize (Best Diploma Thesis) |
| 2009 | Thieme Chemistry - SYNFACTS Poster Prize at the "Synthesefest of the Ludwig-Maximilian University". |

Language and other skills

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| Languages | <ul style="list-style-type: none">German – mother languageEnglish – spoken - excellent, written - excellent, comprehension - excellentFrench – spoken – basic, written – basic, comprehension – basic |
| Technical | <ul style="list-style-type: none">Skilled operator of NMR, IR and HPLC instruments and software (Topspin)Extensive knowledge of office tools and operating systems (MacOS X Snow Leopard, Windows, Ms Office)Experienced with a variety of chemistry software (Beilstein, CS ChemOffice, IsisBase, IsisDraw, SciFinder) |

Social/Teamwork Interests

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| Sports | Rock Climbing, Hiking, Snowboarding, Beach Volleyball, Soccer, Swimming, Basketball, Gym, Cycling |
| Other | Camping, Music, Cooking |